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Strategies for targeting T-cells in allergic diseases and asthma

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Abstract

T helper (Th) 2 lymphocytes play a crucial role in the initiation, progression and persistence of allergic diseases, including asthma. Drugs that interfere with the activation of T-cells or more selectively Th2-specific signaling molecules and drugs that prevent the selective migration into lung tissue are promising novel strategies for the treatment of allergic asthma. Although the mainstay asthma therapy of inhaled glucocorticoids is rather effective, targeting Th2 cells may be an important alternative in childhood. Regulatory T-cells (Treg cells) have a physiological role in protection of unwanted immune responses to auto-antigens and allergens. Literature data indicate that an imbalance between Th2 and Treg cells may underlie development and disease expression of allergic asthma. Drugs or immunotherapies that stimulate these counter-Treg cells may limit aberrant Th2 responses leading to suppression of symptoms. Furthermore, these types of treatments may offer the perspective of disease modification and long-term relief of symptoms.

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Abbreviations: AHR, airway hyperreactivity; AP, activator protein; APC, antigen-presenting cell; BAL, bronchoalveolar lavage; cAMP, cyclic adenosine monophosphate; CBP, CREB-binding protein; CpG DNA, CpG-containing immunostimulatory deoxyribonucleic acid; CsA, cyclosporin A; CTLA4, cytotoxic T lymphocyte antigen 4; DC, dendritic cell; ERK, extracellular-regulated kinase; GR, glucocorticoid receptor; GRE, glucocorticoid response elements; ICAM, intercellular adhesion molecule; Ig, immunoglobulin; IL, interleukin; ITAM, immunoreceptor tyrosine-based activation motif; JNK, Jun N-terminal kinase; LFA, leukocyte function-associated antigen; MAPK, mitogen-activated kinase; NF-AT, nuclear factor for activated T-cells; PAMP, pathogen-associated molecular patterns; PBMC, peripheral blood mononuclear cells; PDE, phosphodiesterase; PGD₂, prostaglandin D₂; PI3-K, phosphoinositide-3 phosphate; SAPK, stress activated protein kinase; STAT, signal transducer and activator of transcription; TARC, thymus and activation-regulated chemokine; TCR, T-cell receptor; TGF- β , transforming growth factor β ; Th, T helper; TLR, toll-like receptor; Treg cells, regulatory T-cell; VCAM, vascular cell adhesion molecule; VLA, very late antigen.

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1. Introduction

High serum levels of immunoglobulin E (IgE) antibodies to common environmental allergens like house-dust or pollen is a key determinant of allergic diseases like allergic asthma, allergic rhinitis and atopic dermatitis. IgE mediates the type-I immediate hypersensitivity reaction that is characterized by crosslinking of receptor-bound IgE on mast cells with allergen, inducing the release of preformed and newly generated mediators that elicit the symptoms of allergic disease. The prevalence of allergic diseases is very high and has shown a considerable increase during the last decades, especially in children, although this increase appears to level off (van Schayck & Smit, 2005). Allergic rhinitis alone affects more than 155 million people worldwide and over 80 million people in Europe have some form of allergic disease. Asthma is one of the most common chronic diseases with over 300 million people worldwide (GINA report 2004). While approximately 90% of children with asthma are allergic, only 50–60% of adult asthmatics display elevated serum levels of allergen-specific IgE. Asthma is further characterized by reversible airway obstruction, chronic eosinophilic airway inflammation, airway remodelling, mucus hypersecretion, and airway hyperresponsiveness (AHR) to bronchospasmogenic stimuli. T-helper (Th) 2 lymphocytes play a critical role in the initiation, progression and persistence of allergic diseases, asthma included. Initially, a disturbed balance between Th1- and Th2-mediated immune responses has been postulated to underlie aberrant Th2 reactions to harmless inhaled allergens. Indeed, allergen-specific T-cell clones isolated from the blood of allergic individuals express a typical Th2 cytokine profile secreting interleukin (IL)-4, IL-5 and minimal IFN- γ and IL-2, whereas those clones from non-atopic individuals displayed a Th1 profile (Kapsenberg et al., 1992). Furthermore, allergic asthma is associated with expression of IL-3, IL-4, IL-5 and GM-CSF in bronchoalveolar cells, strongly supporting Th2 activation (Robinson et al., 1992). Nowadays, Th2-type cytokines IL-4, IL-5, IL-13 are known to be critical for IgE production, airway eosinophilia, mucus hypersecretion and non-specific airway hyperreactivity (AHR). However, it appears that susceptibility to allergic diseases cannot solely be explained by an imbalance between Th1 and Th2 responses (Wills-Karp et al., 2001; Herrick & Bottomly, 2003). Recently, an important immunoregulatory role for regulatory T-cells (Treg cells) has been put forward, capable

of suppressing both Th1- and Th2-mediated adaptive immune responses (van Oosterhout & Bloksma, 2005). Targeting these different T-cell subsets for the treatment of allergic asthma is an interesting strategy that has not yet been widely explored. Interestingly, some T-cell-directed therapies harbour the potential to induce long-lasting suppression or even complete remission of disease.

2. Modulation of T-cell receptor-induced signal transduction during T-cell activation

Activation of T-cells is initiated by processed antigen-derived peptides presented by antigen-presenting cells (APC) to the T-cell receptor (TCR)/CD3 complex. An accessory signal provided by co-stimulatory molecules on APC leads to full T-cell activation and this prevents the induction of T-cell tolerance (T-cell unresponsiveness, also called anergy), which normally occurs when T-cells are stimulated by antigen-derived peptide in absence of an appropriate accessory signal. The most potent accessory signal is provided by CD80/CD86, which interacts with CD28 on the T-cell. TCR/CD3 protein complex activation induces a cascade of phosphorylation reactions. First, non-receptor tyrosine kinases, for example, the Src family kinases Lck and Fyn, phosphorylate immunoreceptor tyrosine-based activation motifs (ITAM) located in the CD3 complex (Pitcher and van Oers, 2003), which serve as a docking site for downstream adaptor molecules containing SH2 and phosphotyrosine domains. This induces recruitment of Syk family tyrosine kinases and activation of adaptor molecules, leading to activation of more downstream signaling molecules, for example, phospholipase C and phosphoinositide-3 phosphate kinase (PI3-K) (Nel, 2002). These molecules can activate multiple signaling cascades, including the Ca²⁺ mobilization/calmodulin pathway, the mitogen-activated kinases (MAPK) extracellular-regulated kinase (ERK) pathway, Jun N-terminal kinase (JNK)/stress activated protein kinase (SAPK) pathway and the p38/Mpk2 MAPK pathway, finally resulting in activation of the transcription factors, nuclear factor for activated T-cells (NF-AT) and activator protein (AP)-1. NF-AT and AP-1 can bind to the promoter of many T-cell cytokine genes and enhance their transcription. Full activation of AP-1 and NF-AT requires co-stimulation of the TCR-induced signal by CD28. Similar to TCR/CD3 signaling, CD28 signaling is mediated by tyrosine kinases (e.g. Lck and Ltk) and subsequent

activation of the PI3K, JNK and Rac-1/p38 MAPK pathways (Zhang et al., 2001). In addition to the co-stimulatory role of CD28, CD28-mediated activation of PI3-kinase is thought to induce downregulation of transcription factors involved in the suppression of T-cell activity. This has for instance been described for the expression of cyclin-dependent kinase inhibitor p27^{kip}, a factor that is involved in tolerance induction (Boussiotis et al., 2000; Appleman et al., 2002; Heijink et al., 2003; Kubsch et al., 2003).

2.1. Inhibition of signal transduction in T-cells

Many novel strategies of asthma therapy are directed towards the inhibition of TCR/CD3-induced pathways. Targeting of the signal transduction pathways involved in the transcription of cytokines genes, in particular Th2-type cytokines, is of interest because of the central role of these cytokines in role in IgE synthesis, mucus production, activation and attraction of eosinophils and airway hyperresponsiveness. It is important to note that the kinase signaling pathways described below not only mediate TCR activation, but also activation of B cell receptor signaling, FcεRI activation, cytokine receptors, G protein-coupled receptors and receptor tyrosine kinases.

Activation of non-receptor tyrosine kinase Syk is involved in TCR signaling and inhibition of this molecule has potential interest in the treatment of asthma. Syk inhibitor BAY61-3606 as well as Syk-selective oligonucleotides have been demonstrated to reduce airway eosinophilia in a rat model of acute asthma (Stenton et al., 2002; Yamamoto et al., 2003; Ulanova et al., 2005). In addition, inhibition of Lyn, a tyrosine kinase that acts upstream of Syk, resulted in a reduction of eosinophilic inflammation in a murine model of asthma (Adachi et al., 1999). Inhibitors of more downstream molecules of TCR signaling, that is, the PI3-K inhibitor LY294002, p38 MAPK inhibitor SB203580 and ERK-1/2 inhibitor PD98059 may also have relevant effects for the treatment of asthma. All these inhibitors reduce α-CD3/α-CD28-induced IL-5 secretion in freshly isolated human T-cells (Heijink et al., 2002). In addition, the use of LY924002 and ERK inhibitor U0126 has been demonstrated to reduce airway infiltration of inflammatory cells, IL-4, IL-5 and IL-13 production and airway hyperresponsiveness in mouse models of asthma (Kwak et al., 2003; Duan et al., 2004; Chialda et al., 2005; Duan et al., 2005). Targeting of JNK by selective inhibitor SP600125 has revealed anti-inflammatory effects in a rat model of asthma (Eynott et al., 2004; Chialda et al., 2005). The drugs SB203850, SB239063 and RWJ67657 (Barnes, 2004) block activity of p38 and are of particular interest in the treatment of asthma, given their potential preference to inhibit Th2-type cytokines (Schafer et al., 1999). Although p38 inhibitor SB203580 antagonized the beneficial effect of ERK-1/2 inhibitor U0126 on lung inflammation and cytokine mRNA levels in bronchoalveolar lavage (BAL) in a murine model of asthma (Chialda et al., 2005), p38 inhibition has also been described to reduce airway inflammation in another mouse study (Underwood et al., 2002). In addition, p38-selective antisense oligonucleotides showed significant beneficial effects in mice, through reduction

of airway inflammatory cell infiltration, IL-4, IL-5 and IL-13 production, mucus secretion and airway hyperresponsiveness (Duan et al., 2005). Several p38 MAP kinase inhibitors are now in Phase II development (Barnes, 2004). Because of the wide distribution of the above described signaling cascades in the immune system, the safety of these selective inhibitors is still of concern.

T-cell selective immunosuppressive drugs have also proven beneficial in asthma. For instance, cyclosporin A (CsA), which blocks activation of NF-AT by inhibiting its dephosphorylation, has strong anti-inflammatory effects and inhibits IL-5 production in peripheral blood mononuclear cells (PBMC) of asthma patients (Mori et al., 1995). CsA has established clinical efficacy in improving lung function in chronic, severe, glucocorticoid-dependent asthma (Powell et al., 2001). FK506, which also inhibits dephosphorylation of NF-AT, has been described to reduce asthma symptoms in a guinea pig model of asthma (Fukuda et al., 1991). However, both CsA and FK506 have serious side effects, including nephrotoxicity, which limits their use for chronic asthma therapy.

2.2. Effects of glucocorticoids and cyclic AMP elevating drugs

At the moment, glucocorticoids are the mainstay of asthma therapy. They have broad anti-inflammatory effects, including suppression of cytokine genes. One of the mechanisms by which this is mediated is the binding to elements in cytokine gene promoters that are recognized by the NF-AT/AP-1 complex. Repression of the IL-5 gene has been shown to involve recruitment of the glucocorticoid receptor (GR) to the NF-AT and AP-1 binding site (Jee et al., 2005). In addition, activated GR can bind to glucocorticoid response elements (GRE) in the promoter of several genes and regulate their expression. Another way of GR to modulate gene transcription is by competitive binding to cofactor CREB-binding protein (CBP)/p300, which is also involved in AP-1-dependent transcription. GR binding to CBP results in histone deacetylation, leading to restricted access of transcription factors to promoter regions of proinflammatory genes (Popescu, 2003). Histone deacetylation is at least in part involved in the repression of transcription of the IL-5 gene by glucocorticoids (Jee et al., 2005). In contrast, however, it has recently been demonstrated that histone deacetylase trichostatin A has T-cell suppressive effects and reduces airway inflammation in a mouse model of asthma by reducing T-cell infiltration, IL-4, IL-5 and IgE levels in BAL fluid (Choi et al., 2005). In addition to histone acetylase inhibition, the upregulatory effect that is exerted by GR on β₂-adrenoreceptor (β₂-AR) expression might be an interesting feature of corticosteroids with respect to the inhibition of T-cell activity. β₂-AR activation results in cyclic adenosine monophosphate (cAMP) production, a 2nd messenger with many anti-inflammatory effects. Although inhaled β₂-agonists have no proven efficacy in reducing airway inflammation in asthma (Caramori & Adcock, 2003), inhibitory effects of β₂-agonists have been observed on TCR/CD28-dependent signaling pathways, Th2-type cytokine production and Th2 cell migration (Staples et al., 2001; Heijink et al., 2004;

Loza et al., 2005). Possibly, the inability of β_2 -agonists to efficiently inhibit airway inflammation in asthma is a consequence of the allergen-induced desensitization of the β_2 -adrenergic receptor on Th2 cells that has been observed in asthma (Meurs et al., 1982; Heijink et al., 2004). Improvement of β_2 -adrenergic function by glucocorticoid treatment may enhance the anti-inflammatory effects of β_2 -agonists. Similar to β_2 -agonists, phosphodiesterase (PDE) 4 inhibitors increase cAMP accumulation and exert anti-inflammatory effects, including inhibition of Th2 cytokine production (Staples et al., 2001; Heijink et al., 2003). There are now emerging preliminary data on the beneficial effect of PDE4 inhibitors on lung function in asthma patients (Lipworth, 2005). Also the non-selective PDE inhibitor Theophylline is used for the treatment and has been described to reduce the number of IL4⁺ and IL-5⁺ cells in bronchial biopsies of asthma patients (Djukanovic et al., 1995; Finnerty et al., 1996). Similar to glucocorticoids, Theophylline has been described to exert anti-asthma effects by increasing the activation of histone deacetylase, which is subsequently recruited by glucocorticoids to suppress inflammatory genes (Ito et al., 2002).

Together, therapies directed towards antigen activation of T-cells and downstream signal transduction pathways may be promising, although the broad immunologic activity might limit their use in some cases.

3. Modulation of T-cell differentiation into functional subsets and expression of Th2 specific transcription factors

While NF-AT and AP-1 are involved in the expression of both Th1 and Th2 cytokines, the production of a restricted Th1 or Th2 cytokine pattern requires expression of specific transcription factors. Differentiation of uncommitted Th cells into Th1 or Th2 cells is induced when T-cells undergo cell cycle progression a specific cytokine environment. The best characterized cytokine to induce Th2 differentiation is IL-4, while IL-12 is well known to induce differentiation towards a Th1 phenotype. Specific transcription factors induced by the Th1- and Th2-directing cytokines IL-12 and IL-4 are signal transducer and activator of transcription (STAT) 4 and STAT6, respectively. STAT6 is essential for the development of Th2 cells in response to IL-4 and has been described to negatively regulate a Th1-specific IL-4 silencer (Kubo et al., 1997). Other transcription factors that are selective for differentiated Th2 cells and Th2 clones are NF-IL-6 (C/EBP- β), c-Maf and GATA-3. GATA-3 may be a key regulator of Th2 cytokine production. GATA-3 expression is enhanced in polarized Th2 cells and downregulated in Th1 cells. Moreover, forced expression of GATA-3 in polarized Th1 cells is sufficient to initiate Th2 cytokine expression (Zhang et al., 1997). GATA-3 is thought to play an important role in the expression of Th2 cytokines in asthma. Enhanced expression of GATA-3 has been observed in the airways of asthma patients, with a further increase upon segmental allergen challenge (Nakamura et al., 1999; Erpenbeck et al., 2003). A Th1-specific transcription factor is T-bet, which may be key in the development of Th1

cells, since T-cells from T-bet^{-/-} mice show defective IFN- γ production. Vice versa, retroviral expression of T-bet has been described to suppress expression of the Th2-type cytokines IL-4 and IL-5 (Szabo et al., 2000).

3.1. Inhibition of Th2 specific effector molecules

Modulating the expression of Th1-, Th2- or Treg-specific transcription factors may become an important tool in the treatment of Th1- or Th2-deviated immune responses, including asthma. New asthma drugs that act on Th2 effector molecules include peroxisome proliferator-activated receptor (PPAR) agonists (e.g. cyclopentenone prostaglandins and thiazolidinediones) and antisense oligonucleotides (Popescu, 2003). Most studies are directed towards inhibition of Th2-specific transcription factor GATA-3. In a murine model of asthma, PPAR-treatment downregulated expression of GATA-3, leading to decreased antigen-induced airway hyperresponsiveness, lung inflammation, eosinophilia, cytokine production as well as serum levels of antigen-specific IgE (Woerly et al., 2003). In a similar model, treatment with PPAR agonists inhibited GATA-3 and decreased Th2-driven IgE production (Woerly et al., 2003). In addition, PPAR γ ligands have been described to reduce IL-5 production in in vitro activated T-cells (Mueller et al., 2003). Both PPAR agonists and glucocorticoids may exert their anti-inflammatory effects through inhibition of histone acetylation (Nie et al., 2005) and similar to PPAR, GR activation has been described to repress IL-5 transcription by histone deacetylase activity and inhibition of GATA-3 transcriptional activation (Jee et al., 2005). Also, treatment with antisense oligonucleotides to GATA-3 has been reported to reduce Th2 cytokine production, lung inflammation and airway hyperresponsiveness in OVA-sensitized mice (Finotto et al., 2001). These data indicate that drugs acting on GATA-3 are promising in the treatment of asthma. In addition, a thiazazole inhibitor of the Th2-specific transcription factor STAT6 may be a potential drug for asthma treatment, as it has been described in vitro to repress IL-4-induced genes (Popescu, 2003). No data on selective inhibitors for Th2-specific transcription factor c-Maf have been published. Although the inhibition of Th2 activity, in particular inhibition of GATA-3, may be an important tool to relieve asthma symptoms and even prevent the disease, risks to develop Th1-mediated immune diseases, such as Crohn's disease and autoimmune diseases should be considered.

4. Targeting regulatory T-cells

With the initial discovery of Th1 and Th2 cells in the mouse that exert mutual inhibitory effects (Mosmann & Coffman, 1989), an imbalance between these 2 arms of the immune response has been postulated to underlie both Th1-mediated autoimmune diseases as well as Th2-mediated allergic diseases and asthma (Kapsenberg et al., 1992; Romagnani, 1992). The balance between Th1 and Th2 cells as an immunoregulatory system to control immune responses to self or foreign antigens, however, appears insufficient to explain many experimental observations (Wills-Karp et al., 2001; Gor et al., 2003; Herrick

& Bottomly, 2003; van Oosterhout & Motta, 2005). CD4⁺ Treg cells are crucial immunoregulatory cells that suppress Th1- and Th2-mediated adaptive immune responses. Treg cells are subdivided in naturally occurring Treg cells and adaptive Treg cells, that are postulated to prevent immune responses against self-antigens and adaptive immune responses, respectively (Bluestone & Abbas, 2003; Sakaguchi, 2005). Adaptive Treg cells are further subdivided into type-1 regulatory T-cells (Tr1 cells) and T-helper type-3 cells (Th3 cells) that mediate suppression via the cytokines IL-10 and transforming growth factor β (TGF- β), respectively. Natural Treg cells, unlike adaptive Treg cells, exert their immunosuppressive effects through T-cell:T-cell/APC contact, but the exact molecular mechanism of suppression has remained incompletely understood (Bluestone & Tang, 2005). As previously reviewed in detail (Hawrylowicz, 2005; van Oosterhout & Bloksma, 2005), literature data indicate that different Treg cell subsets interfere with the development of asthma and other allergic diseases at different stages, like allergic sensitisation, progression to established allergic disease and asthma and severity and persistence of disease.

4.1. Drugs that stimulate regulatory T-cells

The transcription factor FOXP3 appears to be selectively expressed by natural Treg cells both in mice and humans and is a master regulatory gene for the development and function of natural Treg cells (Fontenot et al., 2003; Hori et al., 2003; Walker et al., 2003; Yagi et al., 2004). This makes FOXP3 an interesting therapeutic target protein and may open novel therapeutic strategies aimed at the induction of FOXP3 to convert conventional CD4⁺ T-cells into natural Treg cells with the ultimate goal to reverse aberrant Th2-mediated allergic asthmatic responses (Fig. 1).

As mentioned above, glucocorticoids are widely used in the treatment of allergic asthma for their potent anti-inflammatory effect. Interestingly, glucocorticoids potentiate the suppressive effects of natural Treg cells on allergen-stimulated conventional (CD4⁺ CD25⁻) T-cells from atopic as well as non-atopic donors (Dao Nguyen & Robinson, 2004). The glucocorticoid-induced potentiation of suppressive activity was mediated by enhanced IL-10 production. This mechanism may also be operative in vivo, since Karagiannidis et al. (2004) demonstrated that inhalation or systemic glucocorticoid treatment of asthma patients increased the levels of FOXP3 as well as IL-10 mRNA in peripheral blood CD4⁺ T-cells. In addition, synergistic effects of glucocorticoids and β_2 -adrenoceptor agonists on IL-10 secretion by T-cells have been demonstrated (Peek et al., 2005). Furthermore, glucocorticoids not only enhance IL-10 production in T lymphocytes but also generate new IL-10 producing Treg cells when given in combination with the active form of vitamin D3, 1,25(OH)₂ vitamin D3 (Barrat et al., 2002). Recently, an interesting interaction between glucocorticoids and 1,25(OH)₂ vitamin D3 has been shown (Xystrakis et al., 2005). The failure of T lymphocytes from steroid-resistant asthma patients to upregulate IL-10 production by glucocorticoids in vitro could be restored by 1,25(OH)₂ vitamin D3 leading to levels of IL-10 production comparable to those observed in steroid-sensitive asthma patients.

Altogether, the data demonstrate that glucocorticoid treatment either alone or in combination with β_2 -agonists or 1,25(OH)₂ vitamin D3 potentiates the immunosuppressive effects of Treg cells by increasing IL-10 production and/or FOXP3 expression. Besides glucocorticoids, other hormones, including estrogen and dehydroepiandrosterone and the immunosuppressant rapamycin, can induce FOXP3 in T lymphocytes (Polanczyk et al., 2004; Battaglia et al., 2005; Coenen et al., 2005; Coles et al., 2005). It remains to be established whether

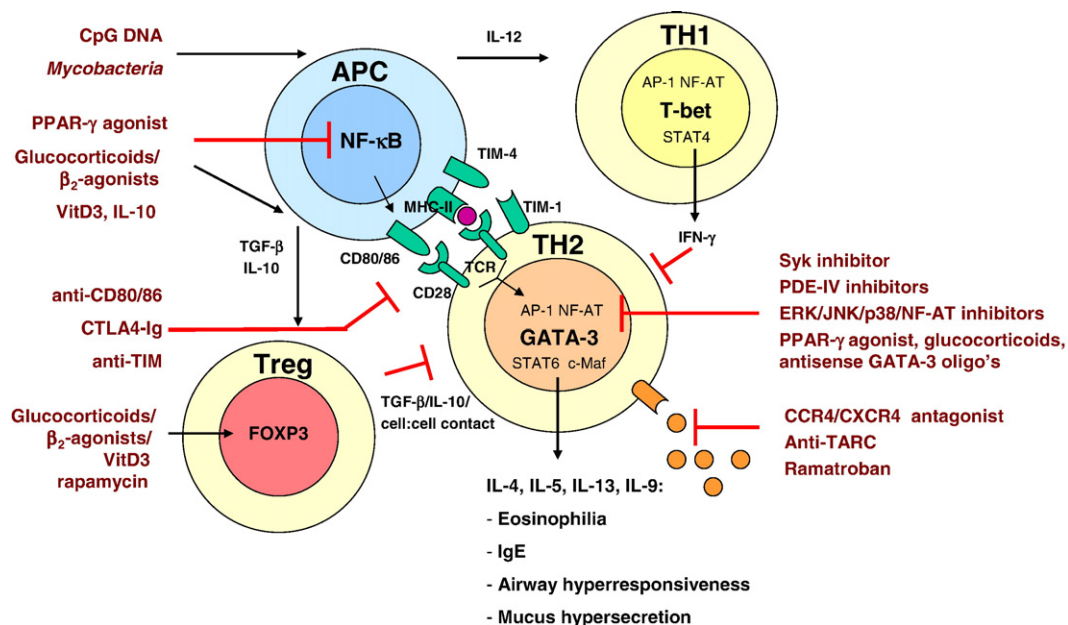


Fig. 1. Overview of therapeutic strategies to limit Th2 activity in allergic diseases, including asthma. Drugs can directly inhibit Th2 effector function or modulate APC function. Otherwise, drugs may act to promote Th1 or Treg cell activities. See text for further details.

failure of different Treg cell subsets in allergic asthma can be restored by these agents. Nonetheless, novel therapeutic strategies aiming at transient or stable induction of Treg cell subsets or potentiation of their immunosuppressive capacities may offer new perspectives for the treatment of allergic asthma.

5. Targeting T-cell costimulation

As described above, CD4⁺ T-cells require 2 independent signals for optimal activation, one through the TCR provided by engagement of the peptide-MHC class II complex on APC and a second costimulatory signal. Although various receptor-ligand pairs for T-cell costimulation have been identified, the CD28 pathway is crucial for primary activation of naive T-cells (Riley & June, 2005). CD28 is constitutively present on the cell-surface of T-cells. The 2 ligands of CD28, CD80 and CD86, are expressed by APCs. On most APC populations, CD86 is expressed constitutively at low levels and is rapidly upregulated upon APC activation, whereas CD80 is inducibly expressed later after activation. CD28 signals promote T-cell activation by augmenting and sustaining T-cell responses initiated by TCR signaling. Blockade of CD28 during activation of naive T-cells renders these cells anergic to secondary activation, even when restimulated in the presence of CD28 ligation. Furthermore, costimulation with CD28 has been described to prevent the induction of Foxp3 expression and suppressive function of Tregs in mice (Fu et al., 2004). Thus, prevention of CD28 costimulation may contribute to tolerance induction and lead to suppressed cytokine expression in T-cells. Another receptor for the CD80 and CD86 is called cytotoxic T lymphocyte antigen 4 (CTLA4; CD152) and delivers inhibitory signals. In contrast to CD28, CTLA4 is only expressed on activated T-cells and is a powerful negative regulator of T-cell activation. Thus, CD28 and CTLA4 compete for the same ligands, CD80 and CD86, but have counterregulatory effects on T-cell activation.

In addition to the CD28 pathway, many different receptor-ligand pairs of T-cell co-stimulation have been identified and are implicated in mouse models of allergic asthma (Deurloo & van Oosterhout, 2004). These costimulatory interactions can be divided into stimulatory signals for T lymphocytes including inducible costimulator (ICOS):ICOS ligand, OX40:OX40 ligand and CD40:CD40 ligand and inhibitory signals including programmed death (PD)-1:PD-1 ligand-1 and -2 and B and T lymphocyte attenuator (BTLA):herpes virus entry mediator (HVEM) (Deurloo & van Oosterhout, 2004; Sedy et al., 2005). However, the CD28 pathway is the only pathway that is currently therapeutically targeted in the clinic and is further discussed in detail (Riley & June, 2005).

5.1. CTLA4-Ig

CTLA4-Ig is a chimeric fusion protein that consists of the extracellular domain of CTLA4 and the heavy-chain constant region of IgG. CTLA4 has a higher affinity for CD80/CD86 when compared to CD28, which makes CTLA4-Ig a potent inhibitor of CD28-mediated T-cell costimulation by blocking of CD80/CD86 on APC. CTLA4-Ig (Abatacept, Orencia®) is

currently in clinical development for the treatment of rheumatoid arthritis, multiple sclerosis and systemic lupus erythematosus, which are also immunological disorders maintained by T-cell activation (Kremer et al., 2005). Moreover, a variant of CTLA4-Ig (Belatacept®) has been developed with 10-fold higher binding avidity for CD80/CD86 as the standard CTLA4-Ig (Larsen et al., 2005). So far, both compounds have shown little apparent toxicity and exhibit substantial clinical activity in these autoimmune diseases as well as renal transplant rejection (Riley & June, 2005).

5.2. CTLA4-Ig in allergic asthma

The therapeutic potential of CTLA4-Ig in the development of Th2-dominated allergic inflammatory responses and AHR in mice has been unequivocally demonstrated (Tsuyuki et al., 1997; van Oosterhout et al., 1997; Keane-Myers et al., 1998). In all of these studies decreased Th2 cytokine production was observed in vivo as shown in BAL fluid or in vitro in supernatants of antigen re-stimulated lung T-cells. The importance of the CD28 costimulatory pathway in allergic asthma has been studied in vitro using PBMC, BAL T-cells and bronchial tissue explants. Larche and colleagues (Larche et al., 1998) demonstrated that, in contrast to peripheral blood CD4⁺ T-cell lines, allergen-induced proliferation and IL-5 production of PBMC were inhibited by CTLA4-Ig. Similarly, van Neerven et al. (1998) demonstrated that allergen- and antigen-specific T-cells of allergic patients and non-allergic control persons are equally dependent on costimulation via the CD28 pathway for their proliferation and cytokine mRNA production. The group of Holgate compared the role of CD28 costimulation in allergen-induced IL-5 production between PBMC cultures and ex vivo bronchial explants. CTLA4-Ig inhibited allergen-induced IL5 production by bronchial tissue explants from mild atopic asthmatic subjects (Jaffar et al., 1999), but not by those of subjects with moderately severe asthma (Lordan et al., 2001). In contrast, blocking of CD28 costimulation by CTLA4-Ig effectively inhibited allergen-induced IL-5 production in PBMC cultures from the same subjects. In agreement herewith, the suppressive effects of CTLA4-Ig on serum IgE levels and airway eosinophilia are more pronounced in a “mild” compared to a more “severe” mouse model of established antigen-induced asthma manifestations (Deurloo et al., 2001).

Together, these data suggest that CTLA4-Ig may have therapeutic potential at least for the treatment of mild forms of asthma. However, to the best of our knowledge no clinical trials on the use of CTLA4-Ig in asthma are currently reported.

5.3. T-cell immunoglobulin and mucin-containing molecules

T-cell immunoglobulin (Ig) and mucin-containing molecules (TIM) comprise a recently described family of molecules expressed predominantly on T-cells (Kuchroo et al., 2003). Th1 cells selectively express TIM-3, while Th2 cells selectively express TIM-1 (Khademi et al., 2004; Chakravarti et al., 2005). Ligation of TIM-3 with its ligand, galectin 9, provides a

negative costimulatory signal to the T-cell leading to down-regulation of Th1 responses (Zhu et al., 2005). TIM-1 was previously identified as the hepatitis A virus receptor and is genetically linked to asthma in humans as well as to experimental asthma in a mouse model (McIntire et al., 2001; McIntire et al., 2004). Recently, TIM-4, which is expressed by APC, was discovered as the endogenous ligand for TIM-1 (Meyers et al., 2005). TIM-1:TIM-4 interaction behaves as a costimulatory signal to T-cells leading to enhanced proliferation and cytokine production (Meyers et al., 2005; Umetsu et al., 2005). Interestingly, blockade of TIM-1 by a monoclonal antibody at the time of allergen challenge inhibits Th2 mediated allergic airway inflammation, mucus hypersecretion and Th2 cytokine production in a mouse model of asthma (Encinas et al., 2005). In contrast, blockade of TIM-1 in a model of respiratory tolerance reversed tolerance induction and increased allergic inflammation (Umetsu et al., 2005). More research is definitely needed to unravel these apparent discrepancies before TIM-1 and its ligand, TIM-4, can be regarded as a novel therapeutic target protein for allergic asthma.

6. Modulation of T-cell trafficking

Chemokines orchestrate immune responses by the attraction of inflammatory cells to target organ. Chemokines are small chemotactic cytokines that induce adhesion and transmigration of leukocytes through the endothelium. Arrest on the endothelium is a prerequisite for the transmigration of leukocytes. A process of rolling precedes this and specialized integrins are involved in the adhesion of T-cells to the endothelium (Luster et al., 2005). These include very late antigen (VLA)-4 and leukocyte function-associated antigen (LFA)-1 (integrin $\alpha 4\beta 1$ and integrin $\alpha L\beta 2$, respectively), which are ligands for the adhesion molecules, vascular cell adhesion molecule (VCAM-1) and intercellular adhesion molecule (ICAM)-1, respectively. VLA-4 and LFA-1 are expressed on memory T-cells and their expression is induced during antigenic activation of T-cells in the lymph nodes. High affinity adhesive interactions have been described between VCAM-1 and VLA-4 expressed on differentiated Th2 cells, whereas VLA-4 expressing Th1 were less capable of sustained interactions with VCAM-1 (Lim et al., 2000). Another adhesion molecule expressed on the endothelium and involved in the transendothelial migration of T-cells is P-selectin. Unlike VLA-4, P-selectin glycoprotein ligand-1 (PSGL-1) appears to be selectively involved in the migration of Th1 cells, since PSGL-1 on Th1 cells, but not Th2 cells, could bind to P-selectin and support migration (Borges et al., 1997). Another study demonstrated that PSGL-1 is functional on both Th1 and Th2 cells, but is expressed more abundantly on Th1 cells (Mangan et al., 2005). Thus, differences in adhesion molecule signaling may contribute to the selective recruitment of T-cell subsets to the lung. In addition to integrins, chemokine receptors are expressed in different patterns on T-cell subsets. While the lymph node-homing chemokine receptor CCR7 is expressed on both naive and memory T-cells, most chemokine receptors (e.g. CCR2, CCR3, CCR4, CCR5, CXCR3 and CXCR5) are induced by antigenic stimulation and only expressed

on memory T-cells. Th0 cells and nonpolarized Th cells preferentially express the chemokine receptors CCR2, CCR4, CCR7 and CXCR3. Instead, CCR5 may be a marker of Treg cells and it has been suggested that ligands for this receptor play a central role in the recruitment of Treg cells (Bystry et al., 2001). Th1 cells abundantly express CCR2, CCR5, CCR7 and CXCR3, with the latter being present on almost all Th1 cells. A substantial percentage of Th2 cells also expresses CCR7 and CCR2. Receptors that are preferentially expressed on Th2 cells are CCR4, CCR8, CXCR4 and (on a low percentage) CCR3 (Annunziato et al., 1999; Kim et al., 2001). Infiltration of CCR4⁺ T-cells has been observed in bronchial tissue after allergen challenge in asthma patients (Panina-Bordignon et al., 2001). There is growing evidence that one of the ligands for CCR4, that is, thymus and activation-regulated chemokine (TARC)/CCL17, plays a key role in recruitment of Th2 cells to the asthmatic airways (Sekiya et al., 2000; Berin et al., 2001; Panina-Bordignon et al., 2001; Sekiya et al., 2002; Bochner et al., 2003; Morgan et al., 2005).

6.1. Inhibition of T-cell lung infiltration

Strategies to inhibit T-cell infiltration in the asthmatic lung are directed to T-cell adhesion or chemoattraction. T-cell adhesion can be targeted by inhibiting the interaction between adhesion molecules and their ligands. In this respect, the use of VLA-4 inhibitors may be relevant in asthma, as demonstrated by their effects in allergen-induced inflammatory responses in sheep (Lin et al., 1999; Singh et al., 2004). However, VLA-4 antagonist IVL745 did not affect the early and late response to inhaled allergen or markers of airway inflammation in patients with mild-to-moderate atopic asthma, except for a modest reduction in sputum eosinophils (Diamant et al., 2005; Norris et al., 2005). The clinical development of inhibitors of VLA-4 or the treatment of asthma was discontinued. While OVA-sensitized/challenged ICAM-1-deficient mice have a dramatically reduced inflammatory influx into the airway/lung and a corresponding attenuation of AHR as compared to wild-type controls (Tang & Fiscus, 2001), no clinical data on blocking integrins or antibodies to ICAM-1 or VCAM-1 are available for asthma.

Treatment with specific antagonists to chemokine receptors is in development for asthma. Trafficking of Th2 cells into the lung is mediated by G_α_i-coupled chemoattractant receptors. G_α_i-signaling is inhibited by pertussis toxin. Pertussis toxin-treated Th2 cells have been described to be unable to traffic into the airways and to induce eosinophilic airway inflammation following OVA challenge in a mouse model of asthma (Mathew et al., 2002). Thus, chemokine receptor antagonism may become an important approach in the treatment of asthma. CCR3 antagonists are likely to be safe and efficient in allergic diseases and their pharmacology has already been defined in phase I/II studies in patients (Erin et al., 2002). However, anti-inflammatory activity of CCR3 antagonists may be predominantly due to reduction in eosinophilic infiltration, since the majority of eosinophils but only a minority of T-cells expresses CCR3. Moreover, in CCR3 deficient mice Th2 trafficking

seems to be intact and these mice can even mount a tissue-specific allergic response (Humbles et al., 2002; Ma et al., 2002). Increased expression of 2 other Th2 specific chemokine receptors, that is, CCR8 and CCR4, has been observed in the asthmatic airway upon allergen challenge. However, CCR8 ligand I-309 was hardly detectable in BAL and serum of asthma patients and knockout of CCR8 did not affect allergic inflammation in mice (Bochner et al., 2003; Chung et al., 2003). In contrast, although blocking of CCR4 failed to reduce Th2 inflammation in the airways of guinea pigs, CCR4 deficiency and monoclonal antibodies against TARC have been described to do so in mouse models of asthma (Kawasaki et al., 2001; Schuh et al., 2002; Conroy et al., 2003). Thus, blocking of CCR4-mediated chemotaxis may be promising in the treatment of asthma. In addition, a CXCR4 agonist has been demonstrated to inhibit allergen-induced inflammation in a murine model of asthma, although increased CXCR4 expression in the asthmatic lung has only been described for eosinophils. Another chemokine receptor that characterizes Th2 cells is CRTH2 (Caramori et al., 2004). Prostaglandin D2 (PGD2), which is produced by mast cells, is believed to be one of the ligands of CRTH2 (Hirai et al., 2001). Ramatroban, an orally active antagonist of receptors activated by PGD2, is marketed in Japan for the treatment of rhinitis (Sugimoto et al., 2003). This drug has also been shown to inhibit airway hyperresponsiveness in humans, but no studies in asthma patients are available yet. Thus, although drugs interfering with chemokines and chemokine receptors have been proven beneficial in animal models, no clinical data on these compounds in asthma have been reported so far (Caramori et al., 2004; Garcia et al., 2005).

7. Indirect modulation of T-cells by targeting dendritic cells

Dendritic cells (DC) are considered to be essential for the priming of naïve CD4⁺ T lymphocytes by delivering signal 1 (T-cell receptor) and 2 (costimulatory). Moreover, they also provide T-cell polarizing signal 3, of which IL-12 family members (IL-12, IL-23 and IL-27) for Th1 generation and IL-10 and TGF- β for respectively Tr1 and Th3 generation, are well-known examples (Kapsenberg, 2003). Herewith, DCs play a crucial role in generation of different effector T-cell subsets, Th1 and Th2, and adaptive Treg cells, Tr1 and Th3 cells (Kapsenberg, 2003; De Jong et al., 2005). DCs that promote Treg cells are generally immature or semi-mature and display lower antigen uptake capacity, lower expression of costimulatory molecules and lower T-cell stimulatory capacity (Lutz & Schuler, 2002). Inhibition of DC maturation by inhibiting NF- κ B activation with 1,25(OH)₂ vitamin D₃, glucocorticoids, or PPAR- γ agonist induces Tr1-like cells in vitro and tolerance in mouse models of transplantation, autoimmune diseases and asthma in vivo (Barrat et al., 2002; Gregori et al., 2002; Hammad et al., 2004; Pedersen et al., 2004). The induction of Tr1 or Th3 cells is related to the production of IL-10 or TGF- β , respectively, by these DC (Lutz & Schuler, 2002). Many pathogens, including viruses, parasites, fungi and bacteria, and isolated, so-called regulato-

ry-type pathogen-associated molecular patterns (PAMP), have been shown to induce the production of either one or both of these cytokines by DC and to facilitate induction of Treg cells in this way (Mills, 2004). It will be of interest to know whether some of these PAMP can be therapeutically exploited for the treatment of allergic asthma.

7.1. Mycobacteria

The hygiene hypothesis implies that there is an inverse relation between the extent of exposure to microbial agents during early childhood and the prevalence of allergic diseases, asthma included. One of the early observations that spurred research in this area was the strong inverse association between Th1-mediated delayed hypersensitivity to *Mycobacterium tuberculosis* and atopy in Japanese schoolchildren (Shirakawa et al., 1997). Consequently, it was anticipated that Th2-dominated allergic responses could be downregulated by exposure to specific bacteria such as heat-killed or live *Mycobacteria* (Rook & Stanford, 1998; Matricardi et al., 2003). It has been shown that prophylactic treatment with *Mycobacterium bacillus calmette-guerin* (BCG) or *Mycobacterium vaccae* can suppress AHR and inflammation in mouse models. The mechanism of action may not be related to the induction of Th1 responses, but rather the production of IL-10 and TGF- β by antigen-specific Treg cells (Zuany-Amorim et al., 2002). Despite these encouraging preclinical data, clinical trials using BCG vaccination or injection of the non-pathogenic mycobacterium *M. vaccae* (SRP299) for the treatment of allergic asthma have been disappointing so far (Renz, 2004). It remains to be seen whether treatment in early childhood, in particular before the onset of allergic disease, offers more perspective for this type of treatment.

7.2. CpG-containing immunostimulatory deoxyribonucleic acid

CpG-containing immunostimulatory deoxyribonucleic acid (CpG DNA) are derived from bacterial genomic DNA and exert their biological effects through interaction with toll-like receptor (TLR) 9. In mouse models, CpG DNA has been shown to inhibit antigen-induced AHR, acute and persistent airway inflammation, airway remodelling, goblet cell hyperplasia and Th2-type cytokine production (Jain et al., 2002; Ikeda et al., 2003; Hessel et al., 2005). The precise mechanisms of action are still incompletely understood, but include the promotion of Th1 or Treg cell responses, inhibition of APC-mediated Th2 cell activation and inhibition of IgE-dependent release of Th2 cytokines from mast-cells. In addition to this direct therapeutic intervention, another successful approach is the combination of CpG DNA with specific allergen immunotherapy as demonstrated in a mouse model (Kline et al., 2002) and patients with allergic rhinitis (Tulic et al., 2004). Currently at least 2 companies have TLR-9 ligands in clinical development for the treatment of allergy and asthma, either alone or in combination with specific allergen immunotherapy (Aventis: AVE7279 and AVE0675 and Dynavax: 1018 ISS).

8. Concluding remarks

Considering the central role of T lymphocytes in the regulation of disease manifestation in allergic asthma, drugs targeting disease-inducing Th2 cells or Treg cells are promising therapeutic strategies (see Fig. 1). Th2 cells can be targeted by inhibition of their activation through inhibition of TCR-induced signalling or inhibition of specific transcription factors as well as by prevention of their migration into the lung tissue. Although these interventions may not lead to long-term beneficial effects, they may be of particular interest when given at the time of disease progression in childhood. A more promising approach to limit aberrant Th2 responses and provide long-term relief of symptoms may be the induction of disease-regulatory T lymphocytes, in particular Treg cells. Although mouse models of allergic asthma suggest that this is feasible, the translation of these types of studies for the treatment of human asthma remains poor. Furthermore, whether these strategies can compete with the mainstay asthma therapy of inhaled glucocorticoids remains to be seen in the future.

References

- Adachi, T., Stafford, S., Sur, S., & Alam, R. (1999). A novel lyn-binding peptide inhibitor blocks eosinophil differentiation, survival, and airway eosinophilic inflammation. *J Immunol* 163, 939–946.
- Annunziato, F., Cosmi, L., Galli, G., Beltrame, C., Romagnani, P., Manetti, R., et al. (1999). Assessment of chemokine receptor expression by human Th1 and Th2 cells in vitro and in vivo. *J Leukoc Biol* 65, 691–699.
- Appleman, L. J., van Puijenbroek, A. A., Shu, K. M., Nadler, L. M., & Boussiotis, V. A. (2002). CD28 costimulation mediates down-regulation of P27kip1 and Cell Cycle progression by activation of the PI3K/PKB signaling pathway in primary human T-cells. *J Immunol* 168, 2729–2736.
- Barnes, P. J. (2004). New drugs for asthma. *Nat Rev Drug Discov* 3, 831–844.
- Barrat, F. J., Cua, D. J., Boonstra, A., Richards, D. F., Crain, C., Savelkoul, H. F., et al. (2002). In vitro generation of interleukin 10-producing regulatory CD4 (+) T-cells is induced by immunosuppressive drugs and inhibited by T helper type 1 (Th1)- and Th2-inducing cytokines. *J Exp Med* 195, 603–616.
- Battaglia, M., Stabilini, A., & Roncarolo, M. G. (2005). Rapamycin selectively expands CD4+CD25+FOXP3+ regulatory T-cells. *Blood*.
- Berin, M. C., Eckmann, L., Broide, D. H., & Kagnoff, M. F. (2001). Regulated production of the T helper 2-Type T-cell chemoattractant TARC by human bronchial epithelial cells in vitro and in human lung xenografts. *Am J Respir Cell Mol Biol* 24, 382–389.
- Bluestone, J. A., & Abbas, A. K. (2003). Opinion-regulatory lymphocytes: natural versus adaptive regulatory T-cells. *Nat Rev Immunol* 3, 253–257.
- Bluestone, J. A., & Tang, Q. (2005). How do CD4(+)/CD25(+) regulatory T-cells control autoimmunity? *Curr Opin Immunol*.
- Bochner, B. S., Hudson, S. A., Xiao, H. Q., & Liu, M. C. (2003). Release of both CCR4-active and CXCR3-active chemokines during human allergic pulmonary late-phase reactions. *J Allergy Clin Immunol* 112, 930–934.
- Borges, E., Tietz, W., Steegmaier, M., Moll, T., Hallmann, R., Hamann, A., et al. (1997). P-selectin glycoprotein ligand-1 (PSGL-1) on T helper 1 but not on T helper 2 cells binds to P-selectin and supports migration into inflamed skin. *J Exp Med* 185, 573–578.
- Boussiotis, V. A., Freeman, G. J., Taylor, P. A., Berezovskaya, A., Grass, I., Blazar, B. R., et al. (2000). P27kip1 functions as an energy factor inhibiting interleukin 2 transcription and clonal expansion of alloreactive human and mouse helper T lymphocytes. *Nat Med* 6, 290–297.
- Bystry, R. S., Aluvihare, V., Welch, K. A., Kallikourdis, M., & Betz, A. G. (2001). B cells and professional APC recruit regulatory t cells via CCL4. *Nat Immunol* 2, 1126–1132.
- Caramori, G., & Adcock, I. (2003). Pharmacology of airway inflammation in asthma and COPD. *Pulm Pharmacol Ther* 16, 247–277.
- Caramori, G., Ito, K., & Adcock, I. M. (2004). Targeting Th2 cells in asthmatic airways. *Curr Drug Targets Inflamm Allergy* 3, 243–255.
- Chakravarti, S., Sabatos, C. A., Xiao, S., Illes, Z., Cha, E. K., Sobel, R. A., et al. (2005). Tim-2 regulates T helper type 2 responses and autoimmunity. *J Exp Med*.
- Chialda, L., Zhang, M., Brune, K., & Pahl, A. (2005). Inhibitors of mitogen-activated protein kinases differentially regulate costimulated T-cell cytokine production and mouse airway eosinophilia. *Respir Res* 6, 36.
- Choi, J. H., Oh, S. W., Kang, M. S., Kwon, H. J., Oh, G. T., & Kim, D. Y. (2005). Trichostatin A attenuates airway inflammation in mouse asthma model. *Clin Exp Allergy* 35, 89–96.
- Chung, C. D., Kuo, F., Kumer, J., Motani, A. S., Lawrence, C. E., Henderson, W. R., Jr., et al. (2003). CCR8 is not essential for the development of inflammation in a mouse model of allergic airway disease. *J Immunol* 170, 581–587.
- Coenen, J. J., Koenen, H. J., Rijssen, E. V., Hilbrands, L. B., & Joosten, I. (2005). Rapamycin, and not cyclosporin A, Preserves the highly suppressive CD27+ subset of human CD4+CD25+ regulatory T-cells. *Blood*.
- Coles, A. J., Thompson, S., Cox, A. L., Curran, S., Gurnell, E. M., & Chatterjee, V. K. (2005). Dehydroepiandrosterone replacement in patients with addison's disease has a bimodal effect on regulatory (CD4(+)/CD25(Hi) and CD4(+)/FoxP3(+)) T-cells. *Eur J Immunol*.
- Conroy, D. M., Jopling, L. A., Lloyd, C. M., Hodge, M. R., Andrew, D. P., Williams, T. J., et al. (2003). CCR4 blockade does not inhibit allergic airways inflammation. *J Leukoc Biol* 74, 558–563.
- Dao Nguyen, X., & Robinson, D. S. (2004). Fluticasone propionate increases CD4CD25 T regulatory cell suppression of allergen-stimulated CD4CD25 T-cells by an IL-10-dependent mechanism. *J Allergy Clin Immunol* 114, 296–301.
- De Jong, E. C., Smits, H. H., & Kapsenberg, M. L. (2005). Dendritic cell-mediated T-cell polarization. *Springer Semin Immunopathol* 26, 289–307.
- Deurloo, D. T., & van Oosterhout, A. J. (2004). Role of T-cell co-stimulation in murine models of allergic asthma. *Clin Exp Allergy* 34, 17–25.
- Deurloo, D. T., van Esch, B. C., Hofstra, C. L., Nijkamp, F. P., & van Oosterhout, A. J. (2001). CTLA4-IgG reverses asthma manifestations in a mild but not in a more "Severe" ongoing murine model. *Am J Respir Cell Mol Biol* 25, 751–760.
- Diamant, Z., Kuperus, J., Baan, R., Nietzmann, K., Millet, S., Mendes, P., et al. (2005). Effect of a very late antigen-4 receptor antagonist on allergen-induced airway responses and inflammation in asthma. *Clin Exp Allergy* 35, 1080–1087.
- Djukanovic, R., Finnerty, J. P., Lee, C., Wilson, S., Madden, J., & Holgate, S. T. (1995). The effects of theophylline on mucosal inflammation in asthmatic airways: biopsy results. *Eur Respir J* 8, 831–833.
- Duan, W., Chan, J. H., Wong, C. H., Leung, B. P., & Wong, W. S. (2004). Anti-inflammatory effects of mitogen-activated protein kinase inhibitor U0126 in an asthma mouse model. *J Immunol* 172, 7053–7059.
- Duan, W., Chan, J. H., McKay, K., Crosby, J. R., Choo, H. H., Leung, B. P., et al. (2005). Inhaled P38alpha mitogen-activated protein kinase antisense oligonucleotide attenuates asthma in mice. *Am J Respir Crit Care Med* 171, 571–578.
- Encinas, J. A., Janssen, E. M., Weiner, D. B., Calarota, S. A., Nieto, D., Moll, T., et al. (2005). Anti-T-Cell Ig and mucin domain-containing protein 1 antibody decreases T(H)2 airway inflammation in a mouse model of asthma. *J Allergy Clin Immunol* 116, 1343–1349.
- Erin, E. M., Williams, T. J., Barnes, P. J., & Hansel, T. T. (2002). Eotaxin receptor (CCR3) antagonism in asthma and allergic disease. *Curr Drug Targets Inflamm Allergy* 1, 201–214.
- Erpenbeck, V. J., Hohlfeld, J. M., Discher, M., Krentel, H., Hagenberg, A., Braun, A., et al. (2003). Increased messenger RNA expression of C-Maf and GATA-3 after segmental allergen challenge in allergic asthmatics. *Chest* 123, 370S–371S.
- Eynott, P. R., Xu, L., Bennett, B. L., Noble, A., Leung, S. Y., Nath, P., et al. (2004). Effect of an inhibitor of Jun N-Terminal protein kinase, SP600125, in single allergen challenge in sensitized rats. *Immunology* 112, 446–453.

- Finnerty, J. P., Lee, C., Wilson, S., Madden, J., Djukanovic, R., & Holgate, S. T. (1996). Effects of theophylline on inflammatory cells and cytokines in asthmatic subjects: a placebo-controlled parallel group study. *Eur Respir J* 9, 1672–1677.
- Finotto, S., De Sanctis, G. T., Lehr, H. A., Herz, U., Buerke, M., Schipp, M., et al. (2001). Treatment of allergic airway inflammation and hyperresponsiveness by antisense-induced local blockade of GATA-3 expression. *J Exp Med* 193, 1247–1260.
- Fontenot, J. D., Gavin, M. A., & Rudensky, A. Y. (2003). Foxp3 programs the development and function of CD4(+)/CD25(+) regulatory T-cells. *Nat Immunol* 3, 3.
- Fu, S., Zhang, N., Yopp, A. C., Chen, D., Mao, M., Chen, D., et al. (2004). TGF-beta induces Foxp3 + T-regulatory cells from CD4 + C. *Am J Transplant* 4, 1614–1627.
- Fukuda, T., Akutsu, I., Motojima, S., & Makino, S. (1991). Inhibition of antigen-induced late asthmatic response and bronchial hyperresponsiveness by Cyclosporin and FK 506. *Int Arch Allergy Appl Immunol* 94, 259–261.
- Garcia, G., Godot, V., & Humbert, M. (2005). New chemokine targets for asthma therapy. *Curr Allergy Asthma Rep* 5, 155–160.
- Gor, D. O., Rose, N. R., & Greenspan, N. S. (2003). TH1-TH2: a procrustean paradigm. *Nat Immunol* 4, 503–505.
- Grogari, S., Giarratana, N., Smirondo, S., Uskokovic, M., & Adorini, L. (2002). A 1alpha,25-Dihydroxyvitamin D(3) analog enhances regulatory T-cells and arrests autoimmune diabetes in NOD mice. *Diabetes* 51, 1367–1374.
- Hammad, H., de Heer, H. J., Soullie, T., Angeli, V., Trottein, F., Hoogsteden, H. C., et al. (2004). Activation of peroxisome proliferator-activated receptor-gamma in dendritic cells inhibits the development of eosinophilic airway inflammation in a mouse model of asthma. *Am J Pathol* 164, 263–271.
- Hawrylowicz, C. M. (2005). Regulatory T-cells and IL-10 in allergic inflammation. *J Exp Med* 202, 1459–1463.
- Heijink, I. H., Vellenga, E., Borger, P., Postma, D. S., de Monchy, J. G., & Kauffman, H. F. (2002). Interleukin-6 promotes the production of interleukin-4 and interleukin-5 by interleukin-2-dependent and -independent mechanisms in freshly isolated human T-cells. *Immunology* 107, 316–324.
- Heijink, I. H., Kauffman, H. F., Postma, D. S., de Monchy, J. G., & Vellenga, E. (2003). Sensitivity of IL-5 production to the cAMP-dependent pathway in human T-cells is reduced by exogenous IL-2 in a phosphoinositide 3-kinase-dependent way. *Eur J Immunol* 33, 2206–2215.
- Heijink, I. H., van den Berge, M., Vellenga, E., de Monchy, J. G., Postma, D. S., & Kauffman, H. F. (2004). Altered Beta2-adrenergic regulation of T-cell activity after allergen challenge in asthma. *Clin Exp Allergy* 34, 1356–1363.
- Herrick, C. A., & Bottomly, K. (2003). To respond or not to respond: T-cells in allergic asthma. *Nat Rev Immunol* 3, 405–412.
- Hessel, E. M., Chu, M., Lizcano, J. O., Chang, B., Herman, N., Kell, S. A., et al. (2005). Immunostimulatory oligonucleotides block allergic airway inflammation by inhibiting Th2 cell activation and IgE-mediated cytokine induction. *J Exp Med* 202, 1563–1573.
- Hirai, H., Tanaka, K., Yoshie, O., Ogawa, K., Kenmotsu, K., Takamori, Y., et al. (2001). Prostaglandin D2 selectively induces chemotaxis in T helper type 2 cells, eosinophils, and basophils via seven-transmembrane receptor CRTH2. *J Exp Med* 193, 255–261.
- Hori, S., Nomura, T., & Sakaguchi, S. (2003). Control of regulatory T-cell development by the transcription factor FOXP3. *Science* 9, 9.
- Humbles, A. A., Lu, B., Friend, D. S., Okinaga, S., Lora, J., Al Garawi, A., et al. (2002). The murine CCR3 receptor regulates both the role of eosinophils and mast cells in allergen-induced airway inflammation and hyperresponsiveness. *Proc Natl Acad Sci U S A* 99, 1479–1484.
- Ikeda, R. K., Nayar, J., Cho, J. Y., Miller, M., Rodriguez, M., Raz, E., et al. (2003). Resolution of airway inflammation following ovalbumin inhalation: comparison of ISS DNA and corticosteroids. *Am J Respir Cell Mol Biol* 28, 655–663.
- Ito, K., Lim, S., Caramori, G., Cosio, B., Chung, K. F., Adcock, I. M., et al. (2002). A molecular mechanism of action of theophylline: induction of histone deacetylase activity to decrease inflammatory gene expression. *Proc Natl Acad Sci U S A* 99, 8921–8926.
- Jaffar, Z. H., Stanciu, L., Pandit, A., Lordan, J., Holgate, S. T., & Roberts, K. (1999). Essential role for both CD80 and CD86 costimulation, but not CD40 interactions, in allergen-induced Th2 cytokine production from asthmatic bronchial tissue: role for alphabeta, but not gammadelta, T-cells. *J Immunol* 163, 6283–6291.
- Jain, V. V., Kitagaki, K., Businga, T., Hussain, I., George, C., O'Shaughnessy, P., et al. (2002). CpG-oligodeoxynucleotides inhibit airway remodeling in a murine model of chronic asthma. *J Allergy Clin Immunol* 110, 867–872.
- Jee, Y. K., Gilmour, J., Kelly, A., Bowen, H., Richards, D., Soh, C., et al. (2005). Repression of interleukin-5 transcription by the glucocorticoid receptor targets GATA3 signaling and involves histone deacetylase recruitment. *J Biol Chem* 280, 23243–23250.
- Kapsenberg, M. L. (2003). Dendritic-cell control of pathogen-driven T-cell polarization. *Nat Rev Immunol* 3, 984–993.
- Kapsenberg, M. L., Jansen, H. M., Bos, J. D., & Wierenga, E. A. (1992). Role of type 1 and type 2 T helper cells in allergic diseases. *C O Immunology* 4, 788–793.
- Karagiannidis, C., Akdis, M., Holopainen, P., Woolley, N. J., Hense, G., Ruckert, B., et al. (2004). Glucocorticoids upregulate FOXP3 expression and regulatory T-cells in asthma. *J Allergy Clin Immunol* 114, 1425–1433.
- Kawasaki, S., Takizawa, H., Yoneyama, H., Nakayama, T., Fujisawa, R., Izumizaki, M., et al. (2001). Intervention of thymus and activation-regulated chemokine attenuates the development of allergic airway inflammation and hyperresponsiveness in mice. *J Immunol* 166, 2055–2062.
- Keaney-Myers, A. M., Gause, W. C., Finkelman, F. D., Xhou, X., & Wills-Karp, M. (1998). Development of murine asthma is dependent upon B7-2 costimulation. *J Immunol* 160, 1036–1043.
- Khademi, M., Illes, Z., Gielen, A. W., Marta, M., Takazawa, N., Baecher-Allan, C., et al. (2004). T-cell Ig- and mucin-domain-containing molecule-3 (TIM-3) and TIM-1 molecules are differentially expressed on human Th1 and Th2 cells and in cerebrospinal fluid-derived mononuclear cells in multiple sclerosis. *J Immunol* 172, 7169–7176.
- Kim, C. H., Rott, L., Kunkel, E. J., Genovese, M. C., Andrew, D. P., Wu, L., et al. (2001). Rules of chemokine receptor association with T-cell polarization in vivo. *J Clin Invest* 108, 1331–1339.
- Kline, J. N., Kitagaki, K., Businga, T. R., & Jain, V. V. (2002). Treatment of established asthma in a murine model using CpG oligodeoxynucleotides. *Am J Physiol Lung Cell Mol Physiol* 283, 170–179.
- Kremer, J. M., Dougados, M., Emery, P., Durez, P., Sibilia, J., Shergy, W., et al. (2005). Treatment of rheumatoid arthritis with the selective costimulation modulator abatacept: twelve-month results of a phase IIB, double-blind, randomized, placebo-controlled trial. *Arthritis Rheum* 52, 2263–2271.
- Kubo, M., Ransom, J., Webb, D., Hashimoto, Y., Tada, T., & Nakayama, T. (1997). T-cell subset-specific expression of the IL-4 gene is regulated by a silencer element and STAT6. *EMBO J* 16, 4007–4020.
- Kubsch, S., Graulich, E., Knop, J., & Steinbrink, K. (2003). Suppressor activity of anergic T-cells induced by IL-10-treated human dendritic cells: association with IL-2- and CTLA-4-dependent G1 arrest of the cell cycle regulated by P27Kip1. *Eur J Immunol* 33, 1988–1997.
- Kuchroo, V. K., Umetsu, D. T., Dekruyff, R. H., & Freeman, G. J. (2003). The TIM gene family: emerging roles in immunity and disease. *Nat Rev Immunol* 3, 454–462.
- Kwak, Y. G., Song, C. H., Yi, H. K., Hwang, P. H., Kim, J. S., Lee, K. S., et al. (2003). Involvement of PTEN in airway hyperresponsiveness and inflammation in bronchial asthma. *J Clin Invest* 111, 1083–1092.
- Larche, M., Till, S. J., Haselden, B. M., North, J., Barkans, J., Corrigan, C. J., et al. (1998). Costimulation through CD86 is involved in airway antigen-presenting cell and T-cell responses to allergen in atopic asthmatics. *J Immunol* 161, 6375–6382.
- Larsen, C. P., Pearson, T. C., Adams, A. B., Tso, P., Shirasugi, N., Strobert, E., et al. (2005). Rational development of LEA29Y (Belatacept), a high-affinity variant of CTLA4-Ig with potent immunosuppressive properties. *Am J Transplant* 5, 443–453.
- Lim, Y. C., Wakelin, M. W., Henault, L., Goetz, D. J., Yednock, T., Cabanas, C., et al. (2000). Alpha4beta1-integrin activation is necessary for high-efficiency T-cell subset interactions with VCAM-1 under flow. *Microcirculation* 7, 201–214.
- Lin, K., Ateeg, H. S., Hsiung, S. H., Chong, L. T., Zimmerman, C. N., Castro, A., et al. (1999). Selective, tight-binding inhibitors of integrin Alpha4beta1 that inhibit allergic airway responses. *J Med Chem* 42, 920–934.

- Lipworth, B. J. (2005). Phosphodiesterase-4 inhibitors for asthma and chronic obstructive pulmonary disease. *Lancet* 365, 167–175.
- Lordan, J. L., Davies, D. E., Wilson, S. J., Dent, G., Corkhill, A., Jaffar, Z., et al. (2001). The role of CD28-B7 costimulation in allergen-induced cytokine release by bronchial mucosa from patients with moderately severe asthma. *J Allergy Clin Immunol* 108, 976–981.
- Loza, M. J., Foster, S., Peters, S. P., & Penn, R. B. (2005). Beta-agonists modulate T-cell functions via direct actions on type 1 and type 2 cells. *Blood*.
- Luster, A. D., Alon, R., & von Andrian, U. H. (2005). Immune cell migration in inflammation: present and future therapeutic targets. *Nat Immunol* 6, 1182–1190.
- Lutz, M. B., & Schuler, G. (2002). Immature, semi-mature and fully mature dendritic cells: which signals induce tolerance or immunity? *Trends Immunol* 23, 445–449.
- Ma, W., Bryce, P. J., Humbles, A. A., Laouini, D., Yalcindag, A., Alenius, H., et al. (2002). CCR3 is essential for skin eosinophilia and airway hyperresponsiveness in a murine model of allergic skin inflammation. *J Clin Invest* 109, 621–628.
- Mangan, P. R., O'Quinn, D., Harrington, L., Bonder, C. S., Kubes, P., Kucik, D. F., et al. (2005). Both Th1 and Th2 cells require P-selectin glycoprotein ligand-1 for optimal rolling on inflamed endothelium. *Am J Pathol* 167, 1661–1675.
- Mathew, A., Medoff, B. D., Carafone, A. D., & Luster, A. D. (2002). Cutting edge: Th2 cell trafficking into the allergic lung is dependent on chemoattractant receptor signaling. *J Immunol* 169, 651–655.
- Matricardi, P. M., Bjorksten, B., Bonini, S., Bousquet, J., Djukanovic, R., Dreborg, S., et al. (2003). Microbial products in allergy prevention and therapy. *Allergy* 58, 461–471.
- McIntire, J. J., Umetsu, S. E., Akbari, O., Potter, M., Kuchroo, V. K., Barsh, G. S., et al. (2001). Identification of Tapr (an airway hyperreactivity regulatory locus) and the linked Tim gene family. *Nat Immunol* 2, 1109–1116.
- McIntire, J. J., Umetsu, D. T., & Dekruff, R. H. (2004). TIM-1, a novel allergy and asthma susceptibility gene. *Springer Semin Immunopathol* 25, 335–348.
- Meurs, H., Koeter, G. H., de Vries, K., & Kauffman, H. F. (1982). The beta-adrenergic system and allergic bronchial asthma: changes in lymphocyte beta-adrenergic receptor number and adenylate cyclase activity after an allergen-induced asthmatic attack. *J Allergy Clin Immunol* 70, 272–280.
- Meyers, J. H., Chakravarti, S., Schlesinger, D., Illes, Z., Waldner, H., Umetsu, S. E., et al. (2005). TIM-4 is the ligand for TIM-1, and the TIM-1-TIM-4 interaction regulates T-cell proliferation. *Nat Immunol*.
- Mills, K. H. (2004). Regulatory T-cells: friend or foe in immunity to infection? *Nat Rev Immunol* 4, 841–855.
- Morgan, A. J., Symon, F. A., Berry, M. A., Pavord, I. D., Corrigan, C. J., & Wardlaw, A. J. (2005). IL-4-expressing bronchoalveolar T-cells from asthmatic and healthy subjects preferentially express CCR 3 and CCR 4. *J Allergy Clin Immunol* 116, 594–600.
- Mori, A., Suko, M., Nishizaki, Y., Kaminuma, O., Kobayashi, S., Matsuzaki, G., et al. (1995). IL-5 production by CD4+ T-cells of asthmatic patients is suppressed by glucocorticoids and the immunosuppressants FK506 and cyclosporin A. *Int Immunol* 7, 449–457.
- Mosmann, T. R., & Coffman, R. L. (1989). TH1 and TH2 cells: different patterns of lymphokine secretion lead to different functional properties. *Ann Rev Immunol* 7, 145–173.
- Mueller, C., Weaver, V., Vanden Heuvel, J. P., August, A., & Cantorna, M. T. (2003). Peroxisome proliferator-activated receptor gamma ligands attenuate immunological symptoms of experimental allergic asthma. *Arch Biochem Biophys* 418, 186–196.
- Nakamura, Y., Ghaffar, O., Olivenstein, R., Taha, R. A., Soussi-Gounni, A., Zhang, D. H., et al. (1999). Gene expression of the GATA-3 transcription factor is increased in atopic asthma. *J Allergy Clin Immunol* 103, 215–222.
- Nel, A. E. (2002). T-cell activation through the antigen receptor: Part 1. Signaling components, signaling pathways, and signal integration at the T-cell antigen receptor synapse. *J Allergy Clin Immunol* 109, 758–770.
- Nie, M., Corbett, L., Knox, A. J., & Pang, L. (2005). Differential regulation of chemokine expression by peroxisome proliferator-activated receptor gamma agonists: interactions with glucocorticoids and Beta2-agonists. *J Biol Chem* 280, 2550–2561.
- Norris, V., Choong, L., Tran, D., Corden, Z., Boyce, M., Arshad, H., et al. (2005). Effect of IVL745, a VLA-4 antagonist, on allergen-induced bronchoconstriction in patients with asthma. *J Allergy Clin Immunol* 116, 761–767.
- Panina-Bordignon, P., Papi, A., Mariani, M., Di Lucia, P., Casoni, G., Bellettato, C., et al. (2001). The C-C chemokine receptors CCR4 and CCR8 identify airway T-cells of allergen-challenged atopic asthmatics. *J Clin Invest* 107, 1357–1364.
- Pedersen, A. E., Gad, M., Walter, M. R., & Claesson, M. H. (2004). Induction of regulatory dendritic cells by dexamethasone and 1alpha,25-Dihydroxyvitamin D(3). *Immunol Lett* 91, 63–69.
- Peek, E. J., Richards, D. F., Faith, A., Lavender, P., Lee, T. H., Corrigan, C. J., et al. (2005). Interleukin 10 secreting 'regulatory' T-cells induced by glucocorticoids and Beta2-agonists. *Am J Respir Cell Mol Biol*.
- Pitcher, L. A., & van Oers, N. S. (2003). T-cell receptor signal transmission: who gives an ITAM? *Trends Immunol* 24, 554–560.
- Polanczyk, M. J., Carson, B. D., Subramanian, S., Afentoulis, M., Vandenberg, A. A., Ziegler, S. F., et al. (2004). Cutting edge: estrogen drives expansion of the CD4+CD25+ regulatory T-cell compartment. *J Immunol* 173, 2227–2230.
- Popescu, F. D. (2003). New asthma drugs acting on gene expression. *J Cell Mol Med* 7, 475–486.
- Powell, N., Till, S., Bungre, J., & Corrigan, C. (2001). The immunomodulatory drugs cyclosporin A, mycophenolate mofetil, and sirolimus (rapamycin) inhibit allergen-induced proliferation and IL-5 production by PBMCs from atopic asthmatic patients. *J Allergy Clin Immunol* 108, 915–917.
- Renz, H. (2004). Usefulness of mycobacteria in redirecting the immune response in atopic disease. *Clin Exp Allergy* 34, 167–169.
- Riley, J. L., & June, C. H. (2005). The CD28 family: a T-cell rheostat for therapeutic control of T-cell activation. *Blood* 105, 13–21.
- Robinson, D. S., Hamid, Q., Ying, S., Tscipulos, A., Barkans, J., Bentley, A. M., et al. (1992). Predominant Th2-Like Bronchoalveolar T lymphocyte population in atopic asthma. *N Engl J Med* 326, 298–304.
- Romagnani, S. (1992). Human TH1 and TH2 subsets: regulation of differentiation and role in protection and immunopathology. *Int Arch Allergy Immunol* 98, 279–285.
- Rook, G. A. W., & Stanford, J. L. (1998). Give us this day our daily germs. *Immunol Today* 19, 113–116.
- Sakaguchi, S. (2005). Naturally arising Foxp3-expressing CD25(+)CD4(+) regulatory T-cells in immunological tolerance to self and non-self. *Nat Immunol* 6, 345–352.
- Schafer, P. H., Wadsworth, S. A., Wang, L., & Siekierka, J. J. (1999). P38 alpha mitogen-activated protein kinase is activated by CD28-mediated signaling and is required for IL-4 production by human CD4+CD45RO+ T-cells and Th2 effector cells. *J Immunol* 162, 7110–7119.
- Schuh, J. M., Power, C. A., Proudfoot, A. E., Kunkel, S. L., Lukacs, N. W., & Hogaboam, C. M. (2002). Airway hyperresponsiveness, but not airway remodeling, is attenuated during chronic pulmonary allergic responses to aspergillus in CCR4-/- mice. *FASEB J* 16, 1313–1315.
- Sedy, J. R., Gavrieli, M., Potter, K. G., Hurchla, M. A., Lindsley, R. C., Hildner, K., et al. (2005). B and T lymphocyte attenuator regulates T-cell activation through interaction with herpesvirus entry mediator. *Nat Immunol* 6, 90–98.
- Sekiya, T., Miyamasu, M., Imanishi, M., Yamada, H., Nakajima, T., Yamaguchi, M., et al. (2000). Inducible expression of a Th2-Type CC chemokine thymus- and activation-regulated chemokine by human bronchial epithelial cells. *J Immunol* 165, 2205–2213.
- Sekiya, T., Yamada, H., Yamaguchi, M., Yamamoto, K., Ishii, A., Yoshie, O., et al. (2002). Increased levels of a TH2-Type CC chemokine thymus and activation-regulated chemokine (TARC) in serum and induced sputum of asthmatics. *Allergy* 57, 173–177.
- Shirakawa, T., Enomoto, T., Shimazu, S., & Hopkin, J. M. (1997). The inverse association between tuberculin responses and atopic disorder. *Science* 275, 77–79.
- Singh, J., Adams, S., Carter, M. B., Cuervo, H., Lee, W. C., Lobb, R. R., et al. (2004). Rational design of potent and selective VLA-4 inhibitors and their utility in the treatment of asthma. *Curr Top Med Chem* 4, 1497–1507.
- Staples, K. J., Bergmann, M., Tomita, K., Houslay, M. D., McPhee, I., Barnes, P. J., et al. (2001). Adenosine 3',5'-cyclic monophosphate (cAMP)-

- dependent inhibition of IL-5 from human T lymphocytes is not mediated by the cAMP-dependent protein kinase A. *J Immunol* 167, 2074–2080.
- Stenton, G. R., Ulanova, M., Dery, R. E., Merani, S., Kim, M. K., Gilchrist, M., et al. (2002). Inhibition of allergic inflammation in the airways using aerosolized antisense to Syk kinase. *J Immunol* 169, 1028–1036.
- Sugimoto, H., Shichijo, M., Iino, T., Manabe, Y., Watanabe, A., Shimazaki, M., et al. (2003). An orally bioavailable small molecule antagonist of CRTH2, ramatroban (BAY U3405), inhibits prostaglandin D2-induced eosinophil migration in vitro. *J Pharmacol Exp Ther* 305, 347–352.
- Szabo, S. J., Kim, S. T., Costa, G. L., Zhang, X., Fathman, C. G., & Glimcher, L. H. (2000). A novel transcription factor, T-Bet, directs Th1 lineage commitment. *Cell* 100, 655–669.
- Tang, M. L., & Fiscus, L. C. (2001). Important roles for L-selectin and ICAM-1 in the development of allergic airway inflammation in asthma. *Pulm Pharmacol Ther* 14, 203–210.
- Tsuyuki, S., Tsuyuki, J., Einsle, K., Kopf, M., & Coyle, A. J. (1997). Costimulation through B7-2 (CD86) is required for the induction of a lung mucosal T helper cell 2 (TH2) immune response and altered airway responsiveness. *J Exp Med* 185, 1671–1679.
- Tulic, M. K., Fiset, P. O., Christodoulouopoulos, P., Vaillancourt, P., Desrosiers, M., Lavigne, F., et al. (2004). Amb a 1-immunostimulatory oligodeoxynucleotide conjugate immunotherapy decreases the nasal inflammatory response. *J Allergy Clin Immunol* 113, 235–241.
- Ulanova, M., Duta, F., Puttagunta, L., Schreiber, A. D., & Befus, A. D. (2005). Spleen tyrosine kinase (Syk) as a novel target for allergic asthma and rhinitis. *Expert Opin Ther Targets* 9, 901–921.
- Umetsu, S. E., Lee, W. L., McIntire, J. J., Downey, L., Sanjanwala, B., Akbari, O., et al. (2005). TIM-1 induces T-cell activation and inhibits the development of peripheral tolerance. *Nat Immunol*.
- Underwood, S. L., Haddad, E., Birrell, M. A., McCluskie, K., Pecoraro, M., Dabrowski, D., et al. (2002). Functional characterization and biomarker identification in the Brown Norway model of allergic airway inflammation. *Br J Pharmacol* 137, 263–275.
- van Neerven, R. J., Van de Pol, M. M., Van der Zee, J. S., Stiekema, F. E., De Boer, M., & Kapsenberg, M. L. (1998). Requirement of CD28-CD86 costimulation for allergen-specific T-cell proliferation and cytokine expression. *Clin Exp Allergy* 28, 808–816.
- van Oosterhout, A. J., & Motta, A. C. (2005). Th1/Th2 paradigm: not seeing the forest for the trees? *Eur Respir J* 25, 591–593.
- van Oosterhout, A. J. M., & Bloksma, N. (2005). Regulatory T lymphocytes in asthma. *Eur Respir J* 26, 918–932.
- van Oosterhout, A. J., Hofstra, C. L., Shields, R., Chan, B., van Ark, I., Jardieu, P. M., et al. (1997). Murine CTLA4-IgG treatment inhibits airway eosinophilia and hyperresponsiveness and attenuates IgE upregulation in a murine model of allergic asthma. *Am J Respir Cell Mol Biol* 17, 386–392.
- van Schayck, C. P., & Smit, H. A. (2005). The prevalence of asthma in children: a reversing trend. *Eur Respir J* 26, 647–650.
- Walker, M. R., Kasprovicz, D. J., Gersuk, V. H., Benard, A., Van Landeghen, M., Buckner, J. H., et al. (2003). Induction of FoxP3 and acquisition of T regulatory activity by stimulated human CD4+CD25- T-cells. *J Clin Invest* 112, 1437–1443.
- Wills-Karp, M., Santeliz, J., & Karp, C. L. (2001). The germless theory of allergic disease: revisiting the hygiene hypothesis. *Nat Rev Immunol* 1, 69–75.
- Woerly, G., Honda, K., Loyens, M., Papin, J. P., Auwerx, J., Staels, B., et al. (2003). Peroxisome proliferator-activated receptors alpha and gamma down-regulate allergic inflammation and eosinophil activation. *J Exp Med* 198, 411–421.
- Xystrakis, E., Kusumakar, S., Boswell, S., Peek, E., Urry, Z., Richards, D. F., et al. (2005). Reversing the defective induction of IL-10-secreting regulatory T-cells in glucocorticoid-resistant asthma patients. *J Clin Invest*.
- Yagi, H., Nomura, T., Nakamura, K., Yamazaki, S., Kitawaki, T., Hori, S., et al. (2004). Crucial role of FOXP3 in the development and function of human CD25+CD4+ regulatory T-cells. *Int Immunol* 16, 1643–1656.
- Yamamoto, N., Takeshita, K., Shichijo, M., Kokubo, T., Sato, M., Nakashima, K., et al. (2003). The orally available spleen tyrosine kinase inhibitor 2-[7-(3,4-Dimethoxyphenyl)-Imidazo[1,2-c]Pyrimidin-5-Ylamino]Nicotinamide Dihydrochloride (BAY 61-3606) blocks antigen-induced airway inflammation in rodents. *J Pharmacol Exp Ther* 306, 1174–1181.
- Zhang, D. H., Cohn, L., Ray, P., Bottomly, K., & Ray, A. (1997). Transcription factor GATA-3 is differentially expressed in murine Th1 and Th2 cells and controls Th2-specific expression of the interleukin-5 gene. *J Biol Chem* 272, 21597–21603.
- Zhang, J., Salojin, K. V., & Delovitch, T. L. (2001). CD28 co-stimulation restores T-cell responsiveness in NOD mice by overcoming deficiencies in Rac-1/P38 mitogen-activated protein kinase signaling and IL-2 and IL-4 gene transcription. *Int Immunol* 13, 377–384.
- Zhu, C., Anderson, A. C., Schubart, A., Xiong, H., Imitola, J., Khoury, S. J., et al. (2005). The Tim-3 ligand galectin-9 negatively regulates T helper type 1 immunity. *Nat Immunol*.
- Zuany-Amorim, C., Sawicka, E., Manlius, C., Le Moine, A., Brunet, L. R., Kemeny, D. M., et al. (2002). Suppression of airway eosinophilia by killed *Mycobacterium vaccae*-induced allergen-specific regulatory T-cells. *Nat Med* 8, 625–629.