Prevalence of hypoglycaemia in a random population after Roux-en-Y gastric bypass after a meal test

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Abstract

Objective: Roux-en-Y gastric bypass (RYGB) is an effective way to induce sustainable weight loss and can be complicated by postprandial hyperinsulinaemic hypoglycaemia (PHH). To study the prevalence and the mechanisms behind the occurrence of hypoglycaemia after a mixed meal tolerance test (MMTT) in patients with primary RYGB.

Design: This is a cross-sectional study of patients 4 years after primary RYGB.

Methods: From a total population of 550 patients, a random sample of 44 patients completed the total test procedures. A standardized mixed meal was used as stimulus. Venous blood samples were collected at baseline, every 10 min during the first half hour and every 30 min until 210 min after the start. Symptoms were assessed by questionnaires. Hypoglycaemia is defined as a blood glucose level below 3.3 mmol/L.

Results: The prevalence of postprandial hypoglycaemia was 48% and was asymptomatic in all patients. Development of hypoglycaemia was more frequent in patients with lower weight at surgery ($P = 0.045$), with higher weight loss after surgery ($P = 0.011$), and with higher insulin sensitivity calculated by the homeostasis model assessment indexes (HOMA2-IR, $P = 0.014$) and enhanced beta cell function (insulinogenic index at 20 min, $P = 0.001$).

Conclusion: In a randomly selected population 4 years after primary RYGB surgery, 48% of patients developed a hypoglycaemic event during an MMTT without symptoms, suggesting the presence of hypoglycaemia unawareness in these patients. The findings in this study suggest that the pathophysiology of PHH is multifactorial.

Introduction

The laparoscopic Roux-en-Y gastric bypass (RYGB) is often used to achieve sustained weight loss, resolve comorbidity and improve survival in patients with obesity (1).

It is an effective procedure, but unfortunately, it can be accompanied by long-term complications such as postprandial hyperinsulinaemic hypoglycaemia (PHH) (2, 3).

PHH typically occurs 1–3 h after a meal and can lead to autonomic and neuroglycopenic symptoms such as sweating, palpitations, hunger and drowsiness, which may even result in coma (4, 5, 6). Studies have observed prevalence rates after primary gastric bypass surgery of 12% when assessed by means of questionnaires and up to 75% when assessed by continuous glucose monitoring.
systems (CGMSs) (7). Glucagon-like peptide-1 (GLP-1) is assumed to play a pivotal role in the pathophysiology of PHH, most convincingly because the GLP-1 antagonist exendin (9-39) prevents postprandial hyperinsulinaemia and the subsequent hypoglycaemia in people with severe symptoms of late dumping (8, 9). Other gut hormones may also play a role after gastric bypass surgery such as peptide tyrosine tyrosine (PYY) in reducing appetite and suppressing pancreatic secretion and the vasoactive intestinal peptide (VIP) as a local neuromodulator released by intrinsic enteric neurons and responsible for the induction of intestinal gluconeogenesis through a VPAC1 receptor-dependent mechanism in enterocytes (10, 11, 12). The role of these hormones in PHH is not clear (12, 13).

Although these studies have provided more insight into PHH, several (methodological) shortcomings should be mentioned. First of all, the questionnaires and symptom scales that were used for the assessment of the prevalence of PHH have not been validated. Further, these subjective methods do not take hypoglycaemic unawareness into account, a phenomenon that is described in patients with diabetes who have frequent episodes of hypoglycaemia (14). Diagnostic tools like CGMS have the advantage of monitoring glucose concentrations during daily activities, but the accuracy of CGMS in the hypoglycaemic range is poor (15, 16). Also, this method cannot be utilized to investigate the pathophysiology of PHH. Dynamic (or provocative) testing, however, can provide more insight into the pathophysiology, for instance, by providing a standardized stimulus or meal in a laboratory while performing extensive hormone analyses. Older studies have used the oral glucose challenge as standardized stimulus, but nowadays this is considered not physiological accurate; therefore, the mixed meal tolerance test (MMTT) is preferred (17). Smaller studies including up to ±20 patients using the MMTT after gastric bypass have been performed comparing patients with spontaneous severe hypoglycaemic symptoms to those without them (18, 19). These extremes of the spectrum of PHH have been informative, but do not allow a proper evaluation of the real prevalence of PHH and its main driving mechanisms in those with milder or no symptoms. For this purpose, a random selection of sufficient numbers of patients is necessary.

Therefore, we investigated the prevalence of PHH by venous sampling and by questionnaires during an MMTT in a random sample of patients with primary RYGB. Furthermore, to gain insight in the mechanisms provoking hypoglycaemia, we performed gut hormone analyses and assessed the beta-cell function and insulin resistance with several indices.

Patients and methods

Study population

Details on the study design were described previously (20). In brief, non-diabetic patients aged 18–75 years who underwent primary RYGB between 2008 and 2011 in our centre were eligible to participate in the study. A power analysis was performed and it revealed that a sample size of 50 patients was able to show a prevalence of at least 11% (based on an earlier study with self-reported symptoms of late dumping). From the entire cohort (n=550), a random sample of 140 patients was drawn. All patients underwent an antecolic–antegastric Roux-en-Y reconstruction with a gastric pouch sized 30–60 mL, a biliopancreatic limb length of 80 cm and an alimentary limb length of 150 cm (21). In total, 51 patients agreed to participate, but due to exclusion and withdrawal of consent, only 44 patients were analysed. The selection process, as well as reasons for exclusion, is shown in Fig. 1. One patient was registered as having a primary gastric bypass, but during the test, we found out she had gastric banding before, and so she was excluded from the analysis. The study protocol was approved by the Regionale Toetsingscommissie Patiëntengebonden Onderzoek Leeuwarden (local Medical Ethical Review Board) and all patients provided written informed consent. This study was conducted between February 2014 and March 2015.

Study protocol

All scheduled patients underwent the MMTT in the morning after an overnight fast of at least 8h. Anthropometric measures were obtained before the test meal and throughout the test. Via a peripheral intravenous cannula, blood samples were obtained before the meal (t=0) and at 10, 20 and 30min followed by every half hour until 210min after the meal. Before the beginning of the test and at every 30min until 210min after the meal, patients filled in symptom-related questionnaires.

The mixed meal consisted of a 200 mL liquid nutrition supplement (Abbott Ensure S Plus) containing 300 kcal, 12.5 g protein, 40.4 g carbohydrate (of which 13.8 g is sugars), 9.84 g fat and 154.9 g water. The patients were asked to finish the meal within 10 min.

Blood analyses

Venous blood for glucose and insulin analysis was collected in lithium–heparin tubes with a gel separator.
For the measurement of Peptide YY (PYY), VIP, active GLP-1 and inactive GLP-1, blood was collected in pre-cooled tubes containing 15% aprotonin; to this, 50μL dipeptidyl peptidase 4 (DPP-4) inhibitor (Merck Millipore DPP4-010) was added by injection through the cap directly prior to blood withdrawal without influencing the integrity of the pre-existing vacuum in the tube (22). All samples were centrifuged at 4°C and analysed immediately (glucose and insulin) or stored at –80°C until analysed (GLP-1, VIP and PYY).

The glucose (hexokinase reaction) and insulin analyses (sandwich principle assay) were performed on a Roche Diagnostics Cobas analytical unit (glucose with 701 module; insulin with 801 module).

The concentrations of the active GLP-1, VIP, inactive GLP-1 and PYY were determined using commercial ELISA kits on a two-plate ELISA processing system (DS2, DYNEX Technologies, Chantilly, VA, USA), as per the manufacturer’s instructions. The following commercial ELISA kits were used for the active form and inactive form of GLP-1: IBL International, Hamburg, Germany, code JP27784 and code JP27788. For human VIP: RayBiotech, Norcross, GA, USA, cat. # EIA-VIP. For human PYY: Millipore Corporation, cat. # EZHPYYT66K. For the active and inactive forms of GLP-1 and VIP, low- and high-level internal quality controls (QCs) were prepared by selecting and pooling plasma containing 15% aprotonin and DPP-4 inhibitor (Merck Millipore DPP4-010). Low and high QCs for human PYY were provided by the manufacturer. The inter-assay variations of the low-level internal QC and the high-level internal QC were 57.0 and 43.7% for active GLP-1; 7.4 and 15.7% for inactive GLP-1; 62.0 and 54.1% for human VIP and 12.1 and 17.2% for human PYY, respectively. Considering these variations, only within-subject changes from baseline were used for analysis. The maximum storage was 30 months. We did not analyse stability of the incretins, but we assume that with the precautions made (immediate storage), stability was warranted (22).
Questionnaire

Symptoms were assessed during the MMTT with the dumping severity score (DSS) developed by Arts et al. This questionnaire is based on symptom pattern descriptions in the literature, using a four-point Likert scale (23). The patient is asked to grade the intensity (0 = absent; 1 = mild; 2 = moderate and 3 = severe, interfering with daily activities) of eight early-dumping symptoms (within 1h after food ingestion) and six symptoms associated/attributed with/to hypoglycaemia (more than 1h after food ingestion). These six hypoglycaemic symptoms are either adrenergic (sweating, palpitations, hunger, tremor) or neuroglycopenic (irritability, drowsiness) (23).

If patients rated three or more symptoms at an intensity 2 or 3 (i.e. moderate or severe, interfering with daily activities) on the DSS of which at least one was a neuroglycopenic symptom, this was considered a clinically relevant sign of postprandial hypoglycaemia based on previous research (21). To assess the psychological impact of these complaints, we also asked if this provoked anxiety or insecurity. In addition to the DSS, all patients were asked to report any other accompanying symptoms.

During the test, all patients were also observed for symptoms of hypoglycaemia by the researchers with open and specific questions and observation of physical evidence of symptoms (sweating, trembling, drowsiness, etc.).

Definition of PHH

Patients were divided into two groups, a group with hypoglycaemia defined as a postprandial glucose concentration <3.3 mmol/L and a group without hypoglycaemia defined by a postprandial glucose concentration ≥3.3 mmol/L in accordance with the guidelines (17). Symptomatic PHH was defined as hypoglycaemia in combination with a positive score on the DSS. Asymptomatic PHH was defined as hypoglycaemia without or with mild symptoms of neuroglycopenic or adrenergic symptoms.

Calculations of beta-cell function and insulin sensitivity

In the fasting state

The HOMA-IR was calculated as the product of fasting insulin and glucose divided by 22.5 (Basal\text{gluc} × Basal\text{ins} / 22.5) (24). The quantitative insulin sensitivity check index (QUICKI) was calculated by 1/(log(fasting insulin in ng/ml)+ (log(fasting glucose in mg/dl)) (25). The HOMA-β was calculated with fasting glucose and insulin concentrations using the HOMA2 calculator available at http://www.dtu.ox.ac.uk/homacalculator/index.php.

In the postprandial state

Insulin resistance was calculated using the formula of the product of the area under the curves (AUCs) of the circulating glucose and insulin responses (AUC\text{gluc} × AUC\text{ins}) as used during an OGTT (26).

Insulin sensitivity calculations were based upon the Matsuda Index (MISI) and was calculated as 10,000/ (square root of (fasting glucose × fasting insulin) × (mean glucose × mean insulin)) (27). The insulin secretion index (ISI) was calculated by dividing the total AUC for insulin at 60 min by the total AUC for glucose at 60 min.

The β-cell sensitivity was calculated by three different methods. First, by the ratio of the AUCs of the circulating insulin and glucose responses (AUC\text{ins} / AUC\text{gluc}) (7, 9). Secondly, by the liquid mixed meal tolerance disposition index (LMTT-DI), which was calculated according the following formula: MISI × ISI (10). Lastly, by the insulinogenic index calculated as the change in the insulin concentration at 10, 20 or 30 min versus baseline divided by the change in glucose in the corresponding time period.

Statistical analysis

Data are presented as mean (± standard deviation), median (interquartile ranges (IQR)), frequencies or percentages, where appropriate.

In order to study the representativeness of the included study population, it was compared with the entire population and the random selection for age, sex, preoperative weight and preoperative comorbidities with the independent Student’s t tests and Fisher’s exact tests.

Differences between the group with hypoglycaemia and the group without hypoglycaemia were assessed with t tests (for continuous variables) or chi-square tests (for categorical variables). An alpha level of 0.05 was used for determining statistical significance. For graphical representations, the mean with the standard error of the mean are shown. All statistical analyses were performed using the Statistical Package for the Social Sciences (SPSS, Inc.), version 23.
Results

Demographic characteristics

Data of 44 patients were available for analysis; 32 women and 12 men with a median age of 47 years (39–56); all demographic characteristics are presented in Table 1. The study population was representative of the entire surgical cohort (n = 550) in terms of age, sex, comorbidities before surgery, preoperative weight and postoperative weight at the time of the study (Table 2).

Patients who developed hypoglycaemia during the MMTT were more often female, had a shorter follow-up after surgery, had less often a history of type 2 diabetes, had a lower BMI and weight at the MMTT and had lost more (excess) weight after gastric bypass surgery (Table 1).

Glucose and insulin

Hypoglycaemia was present in 21 patients (48%) occurring between 30 and 150 min, but mostly between 60 and 120 min (Fig. 2). One patient started the MMTT with a fasting glucose concentration of 2.9 mmol/L; this increased to 9.5 mmol/L at 20 min after the meal and after 60 min, the patient had a hypoglycaemia with a nadir glucose concentration of 1.7 mmol/L. Thirteen patients (30%) developed hypoglycaemia with a nadir glucose concentration below 2.8 mmol/L. Patients who developed a hypoglycaemic episode had significantly lower glucose values at baseline and on all time points after ingestion of the meal except for 10, 180 and 210 min. Insulin concentrations were significantly higher at 10 and 30 min and significantly lower in the group with hypoglycaemia at baseline and in the late postprandial phase from 90 to 210 min.

Hypoglycaemia-related symptoms

No differences in moderate or severe hypoglycaemic symptoms were seen between the group with hypoglycaemia versus the group without hypoglycaemia (Table 3).

In accordance with this, no symptoms of neuroglycopenia or adrenergic stimulation were observed by the investigators.

Insulin sensitivity and beta-cell function

All indices of insulin resistance showed that insulin sensitivity was significantly higher in the group with hypoglycaemia compared to the group without hypoglycaemia (Table 4).

In contrast, the HOMA2-β was not different between the groups; the estimates of the postprandial beta-cell function showed an increased insulin secretion as calculated by the LMTT-DI and the insulinogenic index in the group with hypoglycaemia (Table 4).

Table 1  Comparison of patient characteristics between patients with and without hypoglycaemia.

<table>
<thead>
<tr>
<th></th>
<th>Patients without hypoglycaemia (n = 23)</th>
<th>Patients with hypoglycaemia (n = 21)</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (years)</td>
<td>48 (42–56)</td>
<td>42 (39–56)</td>
<td>0.449</td>
</tr>
<tr>
<td>Female (%)</td>
<td>14 (61)</td>
<td>18 (86)</td>
<td>0.094</td>
</tr>
<tr>
<td>Time between surgery and study (months)</td>
<td>54 (44–58)</td>
<td>43 (38–53)</td>
<td>0.010</td>
</tr>
<tr>
<td>Weight and weight loss</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Weight at surgery (kg)</td>
<td>143 (128–157)</td>
<td>130 (120–147)</td>
<td>0.045</td>
</tr>
<tr>
<td>BMI at surgery</td>
<td>45 (43–52)</td>
<td>45 (41–48)</td>
<td>0.287</td>
</tr>
<tr>
<td>Weight at MMTT (kg)</td>
<td>99 (88–118)</td>
<td>85 (79–94)</td>
<td>0.001</td>
</tr>
<tr>
<td>BMI at MMTT</td>
<td>32 (30–39)</td>
<td>29 (26–32)</td>
<td>0.014</td>
</tr>
<tr>
<td>EWL at MMTT (%)</td>
<td>62 (48–76)</td>
<td>78 (64–92)</td>
<td>0.011</td>
</tr>
<tr>
<td>TWL at MMTT (%)</td>
<td>28 (22–34)</td>
<td>33 (27–41)</td>
<td>0.036</td>
</tr>
<tr>
<td>Comorbidities preoperative</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Type 2 diabetes</td>
<td>13 (57)</td>
<td>1 (5)</td>
<td>0.000</td>
</tr>
<tr>
<td>Hypertension</td>
<td>15 (61)</td>
<td>6 (29)</td>
<td>0.040</td>
</tr>
<tr>
<td>Dyslipidaemia&lt;sup&gt;a&lt;/sup&gt;</td>
<td>6 (26)</td>
<td>3 (14)</td>
<td>0.462</td>
</tr>
<tr>
<td>Comorbidities postoperative</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Type 2 diabetes&lt;sup&gt;b&lt;/sup&gt;</td>
<td>0</td>
<td>0</td>
<td>0.902</td>
</tr>
<tr>
<td>Hypertension</td>
<td>6 (26)</td>
<td>4 (19)</td>
<td>0.609</td>
</tr>
<tr>
<td>Dyslipidaemia</td>
<td>3 (13)</td>
<td>1 (5)</td>
<td></td>
</tr>
</tbody>
</table>

Data are median and interquartile ranges (IQR), or numbers and frequencies (percentages). Bold indicates statistical significance.

<sup>a</sup>Defined as patients using lipid-lowering medication.<sub>b</sub>Exclusion criteria.

EWL, excess weight loss; MMTT, mixed meal tolerance test; TWL, total weight loss.
Other intestinal hormones

The concentrations of PYY, total GLP-1 and VIP measured as a percentage of change were not different between both groups (Fig. 3).

Discussion

In a random population 4 years after primary gastric bypass surgery, 48% of the patients developed a hypoglycaemic event (<3.3 mmol/L) without concurrent symptoms after a test meal. Patients who developed hypoglycaemia after a test meal were more often female and had lost more weight after their operation. In addition, they showed a higher insulin sensitivity (lower HOMA-IR and no prior history of type 2 diabetes) and an enhanced beta-cell function in the postprandial phase.

This is the first study consisting of a randomly selected, sufficiently large number of patients with a mid-term follow-up after primary gastric bypass surgery investigated with a dynamic test of sufficient duration. Previous studies have shown that the prevalence of PHH assessed with an oral glucose tolerance test varied between 10.4 and 80% depending on the glucose load (75 or 100 g) and cut-off value (2.8 or 3.3 mmol/L) and selection of the study population (28, 29, 30, 31, 32).

The study by Raverdy et al. on the prevalence of PHH contains data 60 months after RYGB and found a prevalence of 7.9% of PHH (33). However, they used an OGTT and measured only at 30 and 120 min after oral ingestion with a definition of PHH as having a glucose value lower than 2.8 mmol/L. We chose the MMTT as a provocation test as the composition of this liquid meal with a combination of fat, protein and 40 g of carbohydrate (of which 14 g is sugar) resembles a more normal food intake. To study the prevalence of PHH in a daily life setting, the MMTT is therefore a better test than the OGTT (17).

The first intriguing observation of our study is the high prevalence of PHH in combination with a total lack of related symptoms as reported by patients and as observed by the researchers. This phenomenon is known as hypoglycaemia unawareness or hypoglycaemia-associated autonomic failure (HAAF).

Table 2  Comparison between the sample and entire surgical cohort of patients who had undergone primary gastric bypass.

<table>
<thead>
<tr>
<th></th>
<th>Sample (N = 44)</th>
<th>Entire cohort (N = 506)</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (years)</td>
<td>45.3 (38.1–53.2)</td>
<td>44.4 (37.5–51.5)</td>
<td>0.859</td>
</tr>
<tr>
<td>Female (%)</td>
<td>32 (72.7)</td>
<td>403 (79.6)</td>
<td>0.332</td>
</tr>
<tr>
<td>Comorbidities preoperative</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Type 2 diabetes</td>
<td>13 (29.5)</td>
<td>112 (22.1)</td>
<td>0.265</td>
</tr>
<tr>
<td>Hypertension</td>
<td>19 (43.2)</td>
<td>195 (38.5)</td>
<td>0.629</td>
</tr>
<tr>
<td>Dyslipidaemia</td>
<td>9 (20.5)</td>
<td>84 (16.6)</td>
<td>0.535</td>
</tr>
<tr>
<td>Weight and weight loss</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Weight at surgery (kg)</td>
<td>137 (125–150)</td>
<td>130 (116–146)</td>
<td>0.820</td>
</tr>
<tr>
<td>BMI at surgery</td>
<td>45 (42–48)</td>
<td>44 (41–48)</td>
<td>0.097</td>
</tr>
<tr>
<td>Last weight at outpatient clinic (kg)</td>
<td>95.0 (84.3–106.8)</td>
<td>93.1 (82.0–110.0)</td>
<td>0.336</td>
</tr>
<tr>
<td>Last BMI at outpatient clinic</td>
<td>30 (26–33)</td>
<td>31 (27–35)</td>
<td>0.746</td>
</tr>
<tr>
<td>EWL (%)</td>
<td>75 (61–92)</td>
<td>69 (53–88)</td>
<td>0.263</td>
</tr>
<tr>
<td>TWL (%)</td>
<td>32 (28–39)</td>
<td>30 (23–37)</td>
<td>0.625</td>
</tr>
</tbody>
</table>

Data are median and interquartile ranges (IQR), or numbers and frequencies (percentages).
It is frequently observed in people with type 1 diabetes on intensive insulin treatment who experience recurrent hypoglycaemia, but it has also been suggested to occur during PHH in the bariatric population (14). Our results are in line with Abrahamsson et al., who performed a stepwise hypoglycaemic clamp study in patients before and 23 weeks after gastric bypass surgery (34). They found that not only the hypoglycaemia symptom score, but also the levels of counter-regulatory hormone such as glucagon, cortisol and catecholamine as well as the sympathetic nerve responses were much lower after surgery compared to those before. One study investigated 15 RYGB patients without known hypoglycaemia by means of the CGMS and found that the patients had glucose concentrations below 3.3 mmol/L 2.9% of the time and 1.5% of the time, these concentrations dropped below 2.8 mmol/L (35). The patients were unaware of their hypoglycaemia four out of five (80%) times. Thus, it is plausible that the high prevalence of hypoglycaemia as seen in our study translates in daily life as recurrent hypoglycaemia, leading to hypoglycaemia-induced autonomic failure and hypoglycaemia unawareness. This could also explain the discrepancy of the prevalence of PHH measured during dynamic tests and by means of questionnaires.

Not only after gastric bypass surgery, but also in patients who underwent a vertical banded gastroplasty with 30% weight loss, a decrease in counter-regulation to hypoglycaemia has been observed (36). This suggests that the development of hypoglycaemia unawareness cannot be attributed to a specific type of bariatric surgery. The discrepancy between the asymptomatic low blood sugars in our study and the development of neuroglycopenia for which patients must be hospitalized in daily life is interesting and further research must clear this discrepancy of hypo-unawareness.

The second main finding in the current study is the observation of differences in insulin sensitivity and insulin secretion between the groups with and without hypoglycaemia. The group with hypoglycaemia had in comparison with the group without hypoglycaemia a lower preoperative weight, more postoperative weight loss and were less likely to have a history of type 2 diabetes. All these are factors that are known to contribute to high insulin sensitivity. In our study, this was supported by lower fasting glucose and insulin concentrations and a lower HOMA2-IR.
Also contributing is the increased insulin secretion in the first half hour of the test and the disposition index, which was indicative of an enhanced postprandial beta-cell function. Finally, the patients with a hypoglycaemic event returned to lower insulin levels at later time points. This could reflect higher insulin sensitivity. Our findings of higher insulin sensitivity and increased insulin secretion in patients with hypoglycaemia are in line with the prior studies (14, 28). One of these studies was by Guarino et al. They performed an OGTT in patients who were all diabetic preoperatively, but the majority (74%) were at the time of testing postoperatively in remission (28). Also, Vauers et al. found that the patients after RYGB with hypoglycaemia and during the OGTT had an increased insulin secretion rate in the first 15 min. Just as in the current study, these patients had lost more weight postoperatively and were less frequently diagnosed with diabetes before surgery. However, they did not observe a difference in insulin sensitivity between patients with and without hypoglycaemia (14). This discrepancy with the current study could be explained by the difference in the definition of hypoglycaemia, which was a glucose concentration <2.8 mmol/L in Vauers et al. and <3.3 mmol/L in ours. However, it could also be caused by the difference in inclusion criteria, as they specifically included patients who experienced hypoglycaemic symptoms during daily life, while, in our current study, a random sample was drawn and none of the patients with hypoglycaemia during the MMTT had clinically relevant symptoms. A more rapid glucose peak and a higher GLP-1 response were found in patients with post-gastric bypass hypoglycaemia, suggesting a role for GLP-1 in the pathophysiology (28, 37). Blocking the GLP-1-receptor by the GLP-1-receptor antagonist exendin (9-39) abolishes the occurrence of hypoglycaemia during an MMTT, providing further evidence for a role of GLP-1 in the pathogenesis of PHH (38). Interestingly, we did not find differences in the GLP-1, PYY and VIP concentrations between the group with hypoglycaemia versus the group without hypoglycaemia. Vauers et al. also did not observe differences in GLP-1 (14). Difference in the stimulus, glucose versus a mixed meal, and the selection of patients could be the explanation for this discrepancy. Moreover, the lower glucose concentration, in the group with hypoglycaemia in our study, was already present in the fasting state, which suggests that the postprandial GLP-1 hypersecretion is not the only mechanism contributing to PHH. This may be a consequence of a higher glucose sensitivity, but differences in hepatic glucose production by changes in glucagon concentrations or by changes in autonomic innervation have to be also considered. These mechanisms need to be studied further in detail.

The strength of our study is the random selection of patients with the cohort being comparable to the entire surgical cohort in terms of age, sex, comorbidities before surgery, preoperative weight and postoperative weight at the time of the study. Furthermore, we used an MMTT, which is more close to normal food intake compared to an OGTT.

Some limitations of the study must be mentioned. First, for various reasons, most unrelated to dumping, only 44 patients of the 51 patients in the random sample were tested, However, because most of these exclusions were due to problematic blood withdrawal, the risk of bias...
is low. The demographics of the tested patients were not different from the total group of operated patients, but there is still a possibility of selection bias.

Secondly, the test meal for an MMTT is not standardized, which is a general shortcoming of meal tests. Standardization should be recommended in order to compare studies in future research. Thirdly, patients were not tested before surgery and therefore other predictors beside the demographics cannot be extrapolated from our study. Pigeyre et al. found in nondiabetic patients after RYGB that a normal glucose tolerance test and a lower HbA1c before surgery were predictors of the hypoglycaemia during a 75g OGTT 12 months after surgery (39). Finally, for GLP-1, PYY and VIP, only relative changes compared to the baseline could be presented due to the (unexpected) large inter-assay variation. Another limitation is the fact that no validated questionnaire for early and late dumping is available. The best-known questionnaire is the Sigstad score, which was initially developed as a clinical score for early dumping to be administered by a doctor. Later, some have used a modified Sigstad score as a patient questionnaire and had employed this in post-gastrectomy patients in case of ulcer disease (32, 40). It is not widely used for post-gastric bypass hypoglycaemia. As indicated earlier, the Edinburgh hypoglycaemia questionnaire is also not validated for late dumping. A potentially more useful and newer patient questionnaire is the DSS, developed by Arts and colleagues (23). This questionnaire is used for the evaluation of treatment responses in patients with post-gastric bypass hypoglycaemia. Validated cut-off levels are not available.

From clinical practice, we know that PHH can be provoked by physical activity. Physical activity can influence insulin sensitivity. In this study, we did not ask patients about their physical activity in the days before the test. Also, patients were tested without performing any physical activity. Future research also must focus on this aspect.

Conclusion

In a randomly selected population mid-term after primary RYGB surgery, 48% of patients developed a hypoglycaemic event during an MMTT. No hypoglycaemic symptoms accompanied these events suggesting the presence of hypoglycaemia unawareness in these patients. The findings in this study suggest that the pathophysiology of PHH is multifactorial.

Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

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