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CHAPTER 7

Changes in cerebral oxygenation and cerebral blood flow during hemodialysis - a simultaneous near-infrared spectroscopy and positron emission tomography study

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ABSTRACT

Near-infrared spectroscopy (NIRS) is used to monitor cerebral tissue oxygenation (rSO$_2$) depending on cerebral blood flow (CBF), cerebral blood volume and blood oxygen content. We explored whether NIRS might be a more easy applicable proxy to [$^{15}$O]H$_2$O positron emission tomography (PET) for detecting CBF changes during hemodialysis. Furthermore, we compared potential determinants of rSO$_2$ and CBF. In 12 patients aged ≥65 years, NIRS and PET were performed simultaneously: before (T1), early after start (T2), and at the end of hemodialysis (T3). Between T1 and T3, the relative change in frontal rSO$_2$ (ΔrSO$_2$) was -8±9% ($P=0.001$) and -5±11% ($P=0.08$), whereas the relative change in frontal gray matter CBF (ΔCBF) was -11±18% ($P=0.009$) and -12±16% ($P=0.007$) for the left and right hemisphere, respectively. ΔrSO$_2$ and ΔCBF were weakly correlated for the left ($\rho=0.31$, $P=0.4$), and moderately correlated for the right ($\rho=0.69$, $P=0.03$) hemisphere. The Bland-Altman plot suggested underestimation of ΔCBF by NIRS. Divergent associations of pH, pCO$_2$ and arterial oxygen content with rSO$_2$ were found compared to corresponding associations with CBF. In conclusion, NIRS could be a proxy to PET to detect intradialytic CBF changes, although NIRS and PET capture different physiological parameters of the brain.
INTRODUCTION

Cognitive impairment is highly common in patients with advanced chronic kidney disease (CKD).\(^1\) Cognitive performance might be negatively affected by structural brain lesions that are often present in the CKD population, including lacunar infarctions,\(^2\) microbleeds,\(^3\) and loss of white matter integrity.\(^4,5\) Besides the many risk factors for cognitive decline that are present in patients with CKD, the hemodialysis procedure itself might also induce brain injury. In patients with advanced CKD, the transition to dialysis has been associated with an accelerated decline of cognitive function and an increased incidence of strokes.\(^6,7\)

Hemodialysis involves repetitive fluid removal, thereby frequently resulting in alterations in blood pressure and volume status, which might induce circulatory stress.\(^8\) The fluid removal during hemodialysis is accompanied by an increase of blood viscosity, and rapid shifts in electrolytes, acid-base balance, and uremic solutes.\(^9,10\) Furthermore, exposure of the blood to the extracorporeal circuit during hemodialysis triggers an inflammatory response with complement activation, endothelial activation, and activation of coagulation pathways.\(^11\)-\(^13\) All these processes could theoretically affect the macro- and microvascular cerebral blood flow and cerebral oxygenation.

To unravel potential mechanisms that underlie the link between cognitive impairment and the hemodialysis procedure, we previously evaluated whether hemodialysis has a direct effect on cerebral blood flow (CBF). Using \(^{15}\)O\(\text{H}_2\)O positron emission tomography (PET), we found that hemodialysis induced a 10% decline in global and regional CBF in elderly hemodialysis patients.\(^14\) This CBF decline does not automatically imply an impaired autoregulation, because the dynamic pressure-flow relationship may also be affected by alterations in cerebrovascular resistance apart from autoregulation, such as changes in pH, hematocrit and blood volume during hemodialysis. Second, we found that as hemodialysis-related factors a higher pH, higher tympanic temperature, and a larger ultrafiltration rate and volume were associated with a lower CBF. However, \(^{15}\)O\(\text{H}_2\)O PET-scanning involves radiation, requires an on-site cyclotron for nuclide generation, and is complicated to perform during a hemodialysis session. Therefore, there is a need for an alternative method that is easier to apply to monitor changes in cerebral perfusion during hemodialysis.\(^15\)

A technique that has been proposed to monitor the adequacy of cerebral perfusion is non-invasive near-infrared spectroscopy (NIRS) by measuring frontal cerebral tissue oxygenation.\(^16\) During hemodialysis, relative drops of more than 15% in frontal cerebral tissue oxygenation (rSO\(_2\)) were associated with decreased executive cognitive function one year after the start of hemodialysis.\(^17\) Changes in frontal rSO\(_2\) are commonly considered to
reflect changes in (frontal) CBF,\textsuperscript{18,19} but whether an intradialytic decline in frontal rSO\textsubscript{2} reflects a simultaneous and similar fall in frontal CBF is unknown.

In this study, we aimed to evaluate whether changes in frontal cerebral oxygenation can identify changes in frontal CBF during hemodialysis. In detail, we investigated (i) the correlation and agreement between intradialytic changes in frontal rSO\textsubscript{2} and frontal gray matter CBF, and (ii) how hemodialysis and oxygenation-related factors and markers of inflammation and endothelial activation are associated with changes in rSO\textsubscript{2}, as compared to CBF.

**MATERIALS AND METHODS**

**Ethics**

The study was performed according to the principles of the Declaration of Helsinki and was approved by the Medical Ethical Committee of the University Medical Center Groningen, and registered at clinical trials.gov (NCT02272985). All patients gave written informed consent.

**Study design and patient recruitment**

The study was performed between March and November 2015 and comprised two objectives: (i) to evaluate the effect of hemodialysis on cerebral perfusion, which was published recently,\textsuperscript{14} and (ii) to study the correlation between changes in frontal rSO\textsubscript{2} and changes in frontal and global CBF, as measured by \(\textsuperscript{15}O\textsuperscript{H}_{2}O\) PET-CT.

Hemodialysis patients aged ≥65 years from the department of Nephrology of the University Medical Center Groningen and from the Dialysis Center Groningen with an arteriovenous fistula without significant recirculation were eligible for this study. Patients with a history of dementia, hydrocephalus, cerebrovascular accident, raised intracranial pressure, end-stage liver disease, actively treated cancer, a known significant (>70%) internal carotid artery or major intracranial vessel stenosis, and patients with a contra-indication for MRI were excluded. After study-inclusion, routine Duplex evaluation was performed to exclude subjects with an asymptomatic internal carotid artery stenosis of more than 70% or major intracranial vessel stenosis, because this may interfere with the interpretation of CBF. Patient characteristics were assessed at study entry and retrieved from the patients’ medical history. Based on the highly sensitive technique of \(\textsuperscript{15}O\textsuperscript{H}_{2}O\) and based on former studies that mainly used transcranial Doppler in which the number of hemodialysis patients varied between 12 and 27,\textsuperscript{20-25} we expected that a total of 14 patients would be sufficient, and aimed to include 14 patients.
**Setting**

NIRS monitoring and three PET-CT scans were performed simultaneously during a single, regular hemodialysis session after the longest interdialytic interval (Monday or Tuesday). All hemodialysis study sessions were performed in the afternoon in the PET-CT camera room, with a constant ambient room temperature of 20°C, excluding an effect of outside temperature on cardiovascular stability during the study sessions.

First, NIRS monitoring was started. Next, the first PET-CT scan was performed (T1), after which patients started hemodialysis still being in a horizontal position in the PET-CT camera. After the second PET-CT scan (T2), which was performed at a mean of 21 minutes (range 13-29) after the start of hemodialysis, patients were transferred to a hospital bed adjacent to the PET-CT camera to continue dialysis in a 30-45 degrees supine position. Before the third PET-CT scan (T3), which was performed at the end of the hemodialysis session at mean 209 minutes (range 168-223 minutes) after the start of hemodialysis, patients were transferred back to the PET-CT camera. Prior to each PET-CT measurement, patients rested in the supine position for at least 20 minutes, thereby reducing the influence of postural change on both NIRS and PET measurements.\textsuperscript{26, 27} Blood pressure, heart rate, and tympanic temperature were measured every 30 minutes and before every PET-CT scan. Blood pressure was measured using an automated blood pressure monitor. Cerebrovascular resistance was calculated as the mean arterial pressure (MAP) divided by the CBF of the frontal gray matter. For the dialysis settings, see Supplemental file.

**NIRS monitoring and analysis**

For NIRS monitoring an In Vivo Optical Spectroscopy device (INVOS™ 5100C Cerebral/Somatic Oximeter | Covidien – Medtronic, Minneapolis, USA) was used, with sensors placed bilaterally on the patient's forehead according to the manufacturer's recommendations. The adhesive optodes were connected to the NIRS device and the sampling rate was 0.2 Hz. For the analysis of rSO\textsubscript{2}, we excluded values of zero and excluded rSO\textsubscript{2} values with a quality score <4, to increase accuracy and exclude movement artifacts. Mean rSO\textsubscript{2} values were calculated for the 5-minute time periods during which the three \[^{15}\text{O}]\text{H}_2\text{O} PET-CT scans were performed.

**PET and MRI acquisition**

For the \[^{15}\text{O}]\text{H}_2\text{O} PET-CT a Siemens Biograph 64-mCT (Siemens) Medical Systems, Tennessee, USA) was used. After performing a low-dose CT scan for attenuation and scatter correction, the dynamic PET acquisition (310 sec) was started, followed after 10 sec by an intravenous bolus injection of \[^{15}\text{O}]\text{H}_2\text{O}. The injected dose of \[^{15}\text{O}]\text{H}_2\text{O} was 500 MBq per scan, and 1500 MBq per patient for the study in total. Three of the 36 scans could not
be analyzed due to a technical problem with the automated sampling system (patient-identity 106 [T1], patient-identity 107 [T2], patient-identity 102 [T3]).

To define regional CBF, we also performed magnetic resonance imaging (MRI) in all patients using a 1.5T whole body system (Aera, Siemens, Erlangen, Germany) on a separate day. The scan protocol included T1-weighted, T2-weighted, three-dimensional fluid-attenuated inversion recovery, diffusion-weighted imaging, susceptibility weighted imaging, and two-dimensional phase contrast sequences. No intravenous contrast was used.

**Image reconstruction and processing**

Image processing and pharmacokinetic analysis were performed with PMOD 3.8 software (PMOD Technologies Ltd., Zurich, Switzerland). The average image (time-weighted) was used for rigid matching registration of the individual PET to the individual MRI. The PET list-mode data were reconstructed using the 3D OSEM algorithm (3 iterations and 24 subsets), point spread function correction and time-of-flight, and reconstructed to 28 dynamic frames (1×10 sec, 12×5 sec, 6×10 sec, and 9×20 sec). Data were corrected for attenuation, scatter and radioactivity decay. This resulted in images with a matrix of 400×400×111 of 2 mm voxels, smoothed with a 2 mm filter at full width at half maximum. The volumes of interest were transformed into the individual space, based on the Hammers atlas and limited to the gray matter tissue in the cortical regions (>30% gray matter probability based on standard probability). After spatial registration, pharmacokinetic modeling was applied to the dynamic PET images to calculate the CBF, based on the implementation of the 1-tissue compartment model developed by E. Meyer. Delay of the arterial input function and dispersion in the model were first calculated for the whole brain, and then these resulting values were fixed for the volumes of interest. For additional information on PET processing, see the Supplemental Methods.

**Laboratory measurements**

For the laboratory measurements, including hemoglobin, hematocrit, pO$_2$, pCO$_2$, SaO$_2$, and pH, arterial blood was sampled from the arterial dialysis line just before each PET-CT scan. Arterial O$_2$ content (CaO$_2$) was calculated using the following equation:

$$CaO_2 \ (mL/dL) = 1.34 \times Hb \times (SaO_2/100) + (0.0031 \times pO_2),$$

where Hb represents the hemoglobin concentration (converted to g/dL), SaO$_2$ represents the oxygen saturation (%), and pO$_2$ represents the oxygen pressure (converted to mmHg).

Markers of inflammation included high sensitive C-reactive protein (CRP), and pentraxin-3. Pentraxin-3 responds rapidly to inflammatory stimuli and is considered an appropriate marker for the intradialytic inflammatory response.
Markers of endothelial activation included angiopoietin-1, angiopoietin-2, the angiopoietin 2:1 (AP 2:1) ratio, and von Willebrand factor (vWF).\textsuperscript{32-34}

**Statistical analyses**

First, absolute changes in rSO\textsubscript{2}, CBF, and clinical and laboratory characteristics were studied using linear mixed models (LMM), which allowed for individual random effects. The likelihood ratio test was used to determine whether the LMM including a random intercept and slope statistically better fitted the data as compared to including a random intercept only.

For the primary study objective, the relative change in rSO\textsubscript{2} (\(\Delta rSO_2\)) was compared with the relative change in frontal gray matter CBF (\(\Delta CBF\)) during hemodialysis. We decided to study the correlation between relative changes instead of absolute changes, because NIRS and PET measure different physiological parameters (oxygenation vs. perfusion) and have different units. Besides, we chose the frontal gray matter CBF instead of the total (gray and white) frontal lobe CBF, because it has been estimated that approximately 85% of rSO\textsubscript{2} is derived from more superficial cortical cerebral tissue, thus not including frontal white matter.\textsuperscript{16} First, \(\Delta rSO_2\) and \(\Delta CBF\) were calculated as the mean of the individual percent change between T1 and T3 using descriptive statistics, reported as mean (\%) \(\pm\) SD. Second, Pearson or Spearman correlation tests, whether appropriate, were used to evaluate the correlation between \(\Delta rSO_2\) and \(\Delta CBF\). Finally, we created a Bland-Altman plot with the difference between \(\Delta CBF\) and \(\Delta rSO_2\) as a function of \(\Delta CBF\), since PET is considered the reference method.\textsuperscript{35, 36} 95% levels of agreement were calculated as the mean of the differences \(\pm 1.96^{*}\)SD. Subsequently, we checked for fixed and proportional bias using a T-test and linear regression model, respectively. In an additional analysis, we tested the correlation and agreement of \(\Delta rSO_2\) and \(\Delta CBF\) between T2 (as a second baseline shortly after the start of hemodialysis) and T3, and between T1 and T2. Furthermore, we additionally calculated cerebrovascular resistance (CVR) as MAP/ CBF.

For the secondary study objective, associations of hemodialysis and oxygenation-related factors and of markers of inflammation and endothelial activation with rSO\textsubscript{2} and frontal gray matter CBF were explored. For this objective, we studied absolute instead of relative rSO\textsubscript{2} and CBF values at all time points using LMM, thereby increasing power since this enabled us to use all 33 and 36 measurements of CBF and rSO\textsubscript{2}, respectively, instead of 10 and 12 measurements of \(\Delta CBF\) and \(\Delta rSO_2\), respectively. Furthermore, rSO\textsubscript{2} and CBF values of the left and right hemisphere were merged for the analyses, because rSO\textsubscript{2} and CBF changes did not differ significantly between the left and the right hemisphere. The hemodialysis-related factors were defined previously based on literature and included mean arterial pressure (MAP),\textsuperscript{17} pCO\textsubscript{2},\textsuperscript{37} pH,\textsuperscript{37} tympanic temperature,\textsuperscript{38} hematocrit,\textsuperscript{20-22, 39, 40}
and ultrafiltration (UF) volume and rate. 21,22 For this study, we additionally studied the relation of oxygenation-related factors (CaO2 and pO2) and markers of inflammation (CRP and pentraxin-3) and endothelial activation (angiopoietin-1, angiopoietin-2, AP 2:1 ratio and vWF) with both rSO2 and frontal gray matter CBF change. All the hemodialysis and oxygenation-related factors and markers of inflammation and endothelial activation were studied univariately using LMM, checking the significance of interactions with scan-order. We did not perform adjusting for multiple testing, because the hemodialysis and oxygenation-related factors and markers of inflammation and endothelial activation were selected beforehand.

In supplementary analyses we repeated the main analyses after excluding one outlier. Second, we tested the correlation between the absolute rSO2 and CBF values at all time points. Third, we tested the correlation of Δmean frontal-rSO2 (mean of left and right rSO2) with Δglobal-CBF (from a volume of interest covering the whole brain).

Two-sided P<0.05 was considered statistically significant. Statistical analyses were performed with SPSS, version 23 (SPSS Inc, IBM company, USA), Stata/Se 14.2 (StataCorp LLC, USA), and GraphPad Prism version 5.0 (GraphPad Software, USA).

RESULTS

Patient enrolment and characteristics
Of the 15 patients who gave written informed consent, 12 patients completed the study (Table 1). Three patients withdrew from the study, because of a kidney transplantation, hip fracture, and withdrawal of consent, respectively. None of the patients had to be excluded because of a significant carotid artery stenosis.

Intradialytic NIRS-rSO2 and frontal gray matter PET-CBF changes
Raw individual rSO2 levels during hemodialysis are shown in Figure 1 for the left and right hemisphere. Using LMM, left rSO2 declined from 54.8±5.7% to 51.2±7.3% (absolute difference -4.2% [95% CI, -6.6; -1.8]; P=0.001), whereas right rSO2 declined from 54.1±5.2% before hemodialysis (T1) to 50.6±7.5% (absolute difference -3.1% [95% CI, -6.4; 0.3]; P=0.08) at the end of hemodialysis (T3) (Table 2). Frontal gray matter CBF declined from 44.1±7.8 mL/100g/min at T1 to 38.3±5.4 mL/100g/min at T3 (absolute difference -5.9 mL/100g/min [95% CI, -10.4; -1.5]; P=0.009) in the left hemisphere, and from 44.7±7.7 mL/100g/min at T1 to 38.8±5.1 mL/100g/min at T3 (absolute difference -6.2 mL/100g/min [95% CI, -10.6; -1.7]; P=0.007 in the right hemisphere (Table 2).
The relative change in rSO₂ between T1 and T3 ($\Delta$rSO₂) was mean -8±9% for the left and -5±11% for the right hemisphere, respectively. The relative change in frontal gray matter CBF between T1 and T3 ($\Delta$CBF) was -11±18% for the left and -12±16% for the right hemisphere, respectively. The individual relative changes in $\Delta$MAP, $\Delta$CBF, $\Delta$rSO₂ and $\Delta$CVR per patient between T1 and T3 are shown in Table S1. $\Delta$rSO₂ and $\Delta$CBF were moderately correlated for the right hemisphere ($\rho$ 0.69, $P=0.03$), but weakly correlated for the left hemisphere ($\rho$ 0.31, $P=0.4$) (Figure 2).
Fig 2. Correlation between $\Delta rSO_2$ and $\Delta$ Frontal gray matter Cerebral Blood Flow of the left (2A) and right (2B) hemisphere, calculated between T3 and T1. Correlation coefficient for the left hemisphere: $\rho 0.31 \ (P=0.4)$, and the right hemisphere: $\rho 0.69 \ (P=0.03)$.

The Bland-Altman plot showed moderate agreement (Figure 3). The overall bias was $-3\pm16\% \ (P=0.5)$ for the left and $-5\pm12\% \ (P=0.2)$ for the right hemisphere (Figure 3). Furthermore, linear regression suggested a proportional bias, indicating underestimation of $\Delta CBF$ by NIRS with larger CBF increases or decreases.

Fig 3. Bland-Altman plot of % changes in rSO$_2$ ($\Delta$rSO$_2$) and in CBF ($\Delta$CBF) between T1 (before hemodialysis) and T3 (at the end of hemodialysis), displayed for the left (3A) and right (3B) hemisphere. The X-axis represents $\Delta$CBF (%), while the Y-axis represents the difference between $\Delta$CBF and $\Delta$rSO$_2$. The central solid line represents zero bias. The central dashed line indicates overall bias, which is $-3\%$ for the left, and $-5\%$ for the right hemisphere, calculated as the mean of the differences, while the lower and upper dashed lines represent limits of agreement: $-35\%$ and $29\%$ for the left, and $-28\%$ and $17\%$ for the right hemisphere, respectively. Linear regression suggested the presence of proportional bias (left hemisphere: $P=0.003$, regression equation: $Y= 5.1$.
regression equation: Y = 0.6 + 0.5x. This indicates that NIRS increasingly underestimated ΔCBF with large(r) changes in CBF.

One patient showed a large increase in CBF (patient-id 110) and could be regarded as an outlier, although it is unknown whether a 30-40% increase in CBF is physiologically implausible during hemodialysis. After removal of this outlier, the correlation coefficients were almost similar (correlation ΔrSO₂ and ΔCBF left hemisphere: \( \rho = 0.30 \) (\( P = 0.4 \)), and right hemisphere: \( \rho = 0.64 \) (\( P = 0.06 \)). However, removal of this outlier changed the Bland-Altman analysis yielding an (almost) significant fixed bias now, instead of a proportional bias (left hemisphere: fixed bias -7% (\( P = 0.09 \)), lower and upper limits of agreement -29% and 14%; right hemisphere: fixed bias -8% (\( P = 0.03 \)), lower and upper limits of agreement -25% and 9%).

Additionally, we studied ΔrSO₂ and ΔCBF between T2 and T3, and between T1 and T2. Between T2 and T3, the correlation between ΔrSO₂ and ΔCBF was moderate for the left (\( \rho = 0.64 \), \( P = 0.048 \)) and strong for the right hemisphere (\( \rho = 0.76 \), \( P = 0.01 \)) (Figure S1). The agreement plot was almost similar as for T1 vs. T3, and linear regression again suggested proportional bias (Figure S2). Between T1 and T2, ΔrSO₂ and ΔCBF did not correlate for both hemispheres (Figure S3), and the agreement plot again suggested proportional bias (Figure S4).

Supplementary analyses showed no significant correlation between ΔMAP and ΔrSO₂ (Figure S5), or between ΔMAP and ΔCBF (Figure S6). Second, using LMM, CVR did not change significantly during hemodialysis, and no significant correlation between ΔCVR and ΔrSO₂ was found (Figure S7). Third, no significant correlation between the absolute rSO₂ and absolute CBF values at any time point was found (Figure S8). Finally, defined as T1 versus T3, Δmean-frontal rSO₂ (mean of left and right ΔrSO₂) and Δglobal-CBF (the whole brain as region of interest) were non-significantly correlated (\( \rho = 0.51 \), \( P = 0.1 \)). Defined as T2 versus T3, Δmean-frontal rSO₂ and Δglobal-CBF showed a strong correlation (\( \rho = 0.72 \), \( P = 0.02 \)).

**Associations of hemodialysis-related factors with NIRS-rSO₂ and frontal gray matter PET-CBF**

The mean UF volume, *i.e.* the fluid volume that was removed during hemodialysis, was 1934±781 mL, and the mean UF rate, *i.e.* the rate of fluid removal, was 6.7±2.5 mL/h/kg body weight. During hemodialysis, blood pH, tympanic temperature and hematocrit increased significantly, whereas the MAP did not change significantly (Table 3).

A significant interaction of pH with scan-order was present for the associations between pH and rSO₂, and between pH and CBF. A higher blood pH was associated with a
higher rSO\textsubscript{2} at T3, as compared to T1, but with a lower CBF (Table 4; for detailed information on the interaction effects and confidence intervals, see Table S2). A higher hematocrit was significantly associated with a higher rSO\textsubscript{2} at T2 and T3 as compared to T1, but not with CBF. MAP, tympanic temperature, and UF volume were not associated with rSO\textsubscript{2}, whereas tympanic temperature and UF volume had a significant negative effect on CBF.

**Associations of oxygenation-related factors, and markers of inflammation and endothelial activation with NIRS-rSO\textsubscript{2} and frontal gray matter PET-CBF**

CaO\textsubscript{2} and pentraxin-3 increased significantly during hemodialysis. Intradialytic pCO\textsubscript{2}, pO\textsubscript{2}, CRP, and the endothelial activation markers did not change significantly (Table 3).

Of the oxygenation-related markers, a higher pCO\textsubscript{2} was significantly associated with a lower rSO\textsubscript{2} at T2, as compared to T1. Conversely, a higher pCO\textsubscript{2} was significantly associated with a higher CBF (Table 4). Higher pO\textsubscript{2} was significantly associated with higher rSO\textsubscript{2}, whereas the association between pO\textsubscript{2} and CBF yielded different effects over time (Table 4). CaO\textsubscript{2} was positively associated with rSO\textsubscript{2} whereas it was negatively associated with CBF. The inflammation markers were not associated with rSO\textsubscript{2} or CBF. Of the endothelial activation markers, a higher angiopoietin-2 and AP 2:1 ratio was associated with a lower rSO\textsubscript{2}, and a higher vWF was associated with a higher rSO\textsubscript{2}. None of the endothelial activation markers had an association with CBF.

**Adverse event**

One patient (identity 115) lost consciousness due to dialysis-induced hypotension shortly after the third NIRS/PET measurement. The mean decline in CBF of left and right frontal gray matter was 23\% (both hemispheres -23\%) and the mean frontal rSO\textsubscript{2} decline was 27\% (left -25\%; right -28\%). The patient made a full recovery without sequelae.

**DISCUSSION**

In this study, we found a moderate correlation between frontal ΔrSO\textsubscript{2} as measured with NIRS and ΔCBF of the frontal gray matter as measured with [\textsuperscript{15}O]H\textsubscript{2}O PET during hemodialysis. The agreement analysis showed moderate agreement and a trend towards predominantly a fixed bias with underestimation of ΔCBF by NIRS. Thus, NIRS could be a proxy for PET to capture intradialytic CBF changes, but some correction factor may be needed to correct for the underestimation of ΔCBF by NIRS. Furthermore, considerable differences were noted with regard to associations of hemodialysis- and oxygenation-
related factors and markers of endothelial activation with rSO$_2$ as compared to CBF. This underscores that rSO$_2$ and CBF represent different physiological parameters of the brain.

Few studies have simultaneously performed cerebral oximetry and PET scanning. One study evaluated the change in cerebral blood volume ($\Delta$CBV) as measured by NIRS, with $\Delta$CBV as measured by PET, during normoventilation and during pCO$_2$ manipulation procedures. They reported a moderate correlation ($\rho = 0.56$) between $\Delta$CBV-NIRS and $\Delta$CBV-PET, and an underestimation of $\Delta$CBV by NIRS. Villringer et al. compared changes in rSO$_2$ by NIRS with simultaneous changes in PET-CBF during rest and during cognitive activation tasks. They found a strong correlation ($\rho = 0.88$) between $\Delta$total-Hb (i.e., the sum of $\Delta$oxy-Hb and $\Delta$deoxy-Hb) and $\Delta$CBF, if a penetration depth of near-infrared light of 0.9 cm into the brain cortex was assumed. Another study from the same group showed strong correlations of $\Delta$oxy-Hb ($\rho$ range: 0.74 to 0.75), $\Delta$deoxy-Hb ($\rho$ range: -0.64 to -0.69), and $\Delta$total-Hb ($\rho$ range: 0.88 to 0.93) with $\Delta$CBF, when assuming various penetration depths $\leq$1.35 cm of near-infrared light. Several differences limit the comparison of their findings to ours. First, Villringer et al. compared NIRS to frontal CBF of a small semisphere, a so-called ‘banana-shaped’ region behind the NIRS optode. In contrary, we studied the correlation of NIRS with CBF of the total frontal gray matter. Second, Villringer et al. did not describe the correlation between $\Delta$rSO$_2$ (i.e., $\Delta$oxy-Hb/$\Delta$deoxy-Hb) and $\Delta$CBF. Besides, neither the studies from this group nor other studies investigated patients with CKD or encompassed the hemodialysis process.

The hemodialysis process is a unique physiological stimulus involving many simultaneous hemodynamic and metabolic changes, e.g. a change in pH due to bicarbonate infusion that is not necessarily accompanied by simultaneous changes in pCO$_2$ or pO$_2$. Previous studies reported either no change in rSO$_2$ during hemodialysis, or an rSO$_2$ decline only during the first 35 minutes of hemodialysis, with a subsequent increase in rSO$_2$ yielding a net non-significant change at the end of hemodialysis. Our study is new insofar that we simultaneously studied intradialytic changes in rSO$_2$ by NIRS and changes in CBF by PET.

There is increasing interest in the utilization of NIRS to monitor adequacy of brain perfusion non-invasively. The underlying assumption is that changes in rSO$_2$ reflect changes in CBF, which is theoretically correct if cerebral metabolism, CBV, and additionally CaO$_2$, blood transit time, and oxygen extraction fraction (OEF) remain constant. However, to our knowledge, it is unknown whether this is true during the hemodialysis procedure. First, absolute systemic blood volume was reported to decline by 17% during hemodialysis, but it is unknown whether CBV also declines. Second, during hemodialysis 10% of the patients of a hemodialysis cohort experienced prolonged hypoxia (arterial oxygen saturation <90% at least one third of the treatment time). Third, apart from an
intradialytic effect, it was reported that cerebral oxygen metabolism, blood transit time, and oxygen extraction were altered in hemodialysis patients compared to controls. Nevertheless, since our primary study aim was to evaluate the correlation between ΔrSO$_2$ and ΔCBF, we are not able to draw any conclusion on the other parameters such as CBV, OEF, or blood transit time.

The underestimation of CBF changes by NIRS seems to be related to predominantly a fixed bias, since with removal of one outlier no proportional bias was present anymore. The underestimation of ΔCBF by NIRS could be the result of scattering effects of extracerebral tissue on the transmission of light. Computer modeling showed that in a typical tissue volume interrogated by NIRS, approximately 30% was brain and 70% was extracerebral tissue.

Remarkably, we noted an absent correlation between ΔrSO$_2$ and ΔCBF, defined as T1 versus T2 including the first 30 minutes of hemodialysis. Previous studies reported that PaO$_2$ initially declined during the first 60 minutes of hemodialysis treatment. We speculate that early intradialytic changes in PaO$_2$ influenced ΔrSO$_2$ rather than ΔCBF between T1 and T2 thereby limiting the correlation. Of note, the divergent association of PaO$_2$ with rSO$_2$ as compared to CBF seems to underscore this hypothesis.

Another remarkable finding was the left-right asymmetry in the correlation and Bland-Altman analyses. Because removal of one outlier did not change this asymmetry, we consider this asymmetry a change finding.

We found that on average CVR did not change significantly during hemodialysis. This constant CVR could suggest that static autoregulation might have been disturbed. However, we cannot draw any definite conclusions, since this was not an autoregulation study and many factors change simultaneously during hemodialysis (e.g. pH, hematocrit), which might directly affect CVR.

Four patients experienced an rSO$_2$ drop of >20% during hemodialysis. A 20% rSO$_2$ decline has been proposed as predictor of cerebral ischemia in patients during carotid endarterectomy and cardiac surgery. A >15% drop in rSO$_2$ during hemodialysis was shown to correlate with executive function decline at 1-year follow-up. Therefore, although NIRS tended to underestimate PET-CBF with increasing CBF changes, in our opinion NIRS is still a promising technique to monitor declines in cerebral oxygenation during hemodialysis. Intradialytic changes in cerebral oxygenation might yield important information on intradialytic brain homeostasis, apart from intradialytic changes in cerebral perfusion. More studies are needed to explore whether large intradialytic rSO$_2$ drops are associated with incident cerebral ischemic injury and decline of cognitive function during follow-up.

The second aim of this study was to explore associations of several clinical and
laboratory parameters with rSO\textsubscript{2} as compared to CBF. No association of MAP with rSO\textsubscript{2} was found, similar to previous studies.\textsuperscript{45,46} Remarkably, pH was positively associated with rSO\textsubscript{2} and negatively with CBF. The positive association of pH with rSO\textsubscript{2} might be explained by a leftward shift of the oxygen-hemoglobin dissociation curve due to the increase in pH during dialysis, thereby theoretically increasing rSO\textsubscript{2}. However, others have reported a negative association of pH with rSO\textsubscript{2} in dialysis patients.\textsuperscript{57} Further examination is required on the effects of pH on CBF and rSO\textsubscript{2} during hemodialysis, especially because the intradialytic change in pH is a modifiable factor by lowering the bicarbonate concentration in the dialysate.

To our knowledge, potential associations of inflammation and endothelial activation markers with cerebral tissue oxygenation have not been reported previously. We found an association between endothelial activation markers and rSO\textsubscript{2}, since a higher angiopoietin-2 and angiopoietin 2:1 ratio was associated with lower rSO\textsubscript{2}. Angiopoietin-1 stabilizes the endothelium, whereas angiopoietin-2 functions as a vessel-destabilizing molecule.\textsuperscript{58} A higher angiopoietin 2:1 ratio seems to represent loss of endothelial barrier integrity,\textsuperscript{59} and might be an early marker of endothelial activation and dysfunction.\textsuperscript{60,61} A possible explanation for the association between angiopoietin-2, and the angiopoietin 2:1 ratio with rSO\textsubscript{2} might be found in the lungs. Recently, angiopoietin-2, which is stored in pulmonary epithelial cells, was suggested to have effects on gas exchange.\textsuperscript{62} Nevertheless, the relation between angiopoietin-2 and the angiopoietin 2:1 ratio and rSO\textsubscript{2} needs further examination, and is beyond the scope of this study.

There are a number of potential weaknesses to this study. The sample size of this study was small and one outlier might have had a relative large influence in the analyses. Furthermore, the findings on our second study aim, \textit{i.e.}, associations of hemodialysis and oxygenation-related factors, and inflammation and endothelial activation markers with rSO\textsubscript{2} and CBF, should be purely considered as hypothesis generating. Larger studies are needed to evaluate the effect size of various factors and markers dynamically by multivariate analysis, especially because various hemodynamic, metabolic, and laboratory characteristics change simultaneously during hemodialysis. Second, NIRS and CBF measurements were performed in a supine position, whereas in general patients are in a semi-upright sitting position during hemodialysis. A semi-upright sitting position might have changed the rSO\textsubscript{2} and CBF values but should not alter the correlation between both. Third, for a future study the use of a NIRS device that provides oxyHb, deoxyHb and total Hb levels is advised, because oxyHb better relates to arterial inflow than rSO\textsubscript{2}, which is a mix of arterial and venous circulation. Besides, such study might also provide more information on transit time (changes) during hemodialysis, which we were not able to take into account.
In conclusion, NIRS could be used as a proxy to PET to detect intradialytic CBF changes, but a correction factor may be needed to correct for the underestimation of CBF changes by NIRS. The different associations of hemodialysis- and oxygenation-related factors and markers of endothelial activation with rSO₂ as compared to CBF underscore that NIRS and PET capture different physiological parameters of the brain.

ACKNOWLEDGEMENTS

We want to thank the positron emission tomography technicians Yvonne van der Knaap, Eelco Severs, Paul van Snick, Johan Wiegers, and Aafke Zeilstra of the Medical Imaging Center, Department of Nuclear Medicine and Molecular Imaging at University Medical Center Groningen, The Netherlands for their technical support during the study sessions. Furthermore, we want to thank medical students Brandt Dijksterhuis, Thom Eshuis, Rozemarijn Ettema, Marleen Huberts, and Renske Wiersema for their help with the study sessions.
**Table 1** Patient characteristics (*N*=12)

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (yr)</td>
<td>75.4 ± 5.2</td>
</tr>
<tr>
<td>Men (%)</td>
<td>7 (58%)</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>26.6 ± 3.5</td>
</tr>
<tr>
<td><strong>Primary kidney disease:</strong> (%)</td>
<td></td>
</tr>
<tr>
<td>Glomerulonephritis</td>
<td>4 (33%)</td>
</tr>
<tr>
<td>Diabetes</td>
<td>1 (8%)</td>
</tr>
<tr>
<td>Vascular</td>
<td>3 (25%)</td>
</tr>
<tr>
<td>Other diagnosis</td>
<td>3 (25%)</td>
</tr>
<tr>
<td>Unknown</td>
<td>1 (8%)</td>
</tr>
<tr>
<td><strong>Current smoker</strong></td>
<td>4 (33%)</td>
</tr>
<tr>
<td>Hypertension</td>
<td>11 (73%)</td>
</tr>
<tr>
<td>Myocardial infarction</td>
<td>2 (17%)</td>
</tr>
<tr>
<td>Heart failure</td>
<td>1 (8%)</td>
</tr>
<tr>
<td>Peripheral artery disease</td>
<td>1 (8%)</td>
</tr>
<tr>
<td>COPD</td>
<td>1 (8%)</td>
</tr>
<tr>
<td><strong>Medication:</strong></td>
<td></td>
</tr>
<tr>
<td>CCB</td>
<td>4 (33%)</td>
</tr>
<tr>
<td>Nitrate</td>
<td>3 (25%)</td>
</tr>
<tr>
<td>ACE inhibitor</td>
<td>1 (8%)</td>
</tr>
<tr>
<td>Angiotensin receptor blocker</td>
<td>1 (8%)</td>
</tr>
<tr>
<td>Beta-blocker</td>
<td>9 (75%)</td>
</tr>
</tbody>
</table>

Data are presented as means ± SD or (range), or number and percentages (%). ACE: angiotensin-converting enzyme; BMI: body mass index; CBF: cerebral blood flow; CCB: calcium channel blocker.
### Table 2

Intradialytic changes in NIRS -\( rSO_2 \) and in PET -\( CBF \)

<table>
<thead>
<tr>
<th>Region</th>
<th>Before Start HD</th>
<th>At the End of HD</th>
<th>Dialysis Treatment Effect</th>
</tr>
</thead>
<tbody>
<tr>
<td>Frontal left (%)</td>
<td>54.8 ± 5.7</td>
<td>51.6 ± 7.7</td>
<td>-3.2 ± 6.5 ( -6.6; -1.8)</td>
</tr>
<tr>
<td>Frontal right (%)</td>
<td>54.1 ± 5.2</td>
<td>51.6 ± 6.5</td>
<td>-2.5 ± 7.1 ( -5.7; 1.3)</td>
</tr>
<tr>
<td>Cerebral blood flow (mL/100g/min)</td>
<td>44.1 ± 6.2</td>
<td>42.7 ± 5.4</td>
<td>-4.9 ± 5.1 ( -10.4; 1.0)</td>
</tr>
<tr>
<td>GM left (%)</td>
<td>44.8 ± 7.7</td>
<td>43.5 ± 6.7</td>
<td>-2.8 ± 5.1 ( -5.3; 2.0)</td>
</tr>
<tr>
<td>GM right (%)</td>
<td>44.9 ± 7.8</td>
<td>43.5 ± 6.9</td>
<td>-2.7 ± 5.1 ( -5.3; 2.1)</td>
</tr>
</tbody>
</table>

Data are presented as unadjusted means ± SD. Dialysis treatment effects are obtained from linear mixed effects models and are presented as estimated difference (95% CI). Abbreviations: GM: gray matter; HD: hemodialysis.

*\( P < 0.05 \), **\( P < 0.01 \).
### Table 3: Intradialytic Changes in Hemodialysis and Oxygenation-related Factors, and in Inflammation and Endothelial Activation Markers

<table>
<thead>
<tr>
<th>Region</th>
<th>Before Start HD</th>
<th>At the End of HD</th>
<th>Dialysis Treatment Effect</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Hemodialysis</strong>&lt;br&gt; MAP (mmHg)</td>
<td>101 ± 11</td>
<td>105 ± 15</td>
<td>-6 (-14; 1)</td>
</tr>
<tr>
<td>Tympanic temperature (°C)</td>
<td>36.3 ± 0.5</td>
<td>36.2 ± 0.5</td>
<td>-0.3 (-0.6; 0.0)</td>
</tr>
<tr>
<td>pH</td>
<td>7.38 ± 0.04</td>
<td>7.40 ± 0.03</td>
<td>0.10 (0.08; 0.12)**&lt;br&gt; <strong>p&lt;0.01</strong>&lt;br&gt; <strong>p&lt;0.001</strong>&lt;br&gt; <strong>p&lt;0.001</strong></td>
</tr>
<tr>
<td>Hematocrit (v/v)</td>
<td>0.33 ± 0.04</td>
<td>0.31 ± 0.04</td>
<td>0.02 (0.006; 0.03)**&lt;br&gt; <strong>p&lt;0.01</strong>&lt;br&gt; <strong>p&lt;0.01</strong>&lt;br&gt; <strong>p&lt;0.01</strong></td>
</tr>
<tr>
<td><strong>Oxygenation</strong>&lt;br&gt; pO₂ (kPa)</td>
<td>12.2 ± 2.1</td>
<td>11.5 ± 1.8</td>
<td>0.4 (-0.7; 1.5)</td>
</tr>
<tr>
<td>pCO₂ (kPa)</td>
<td>5.0 ± 0.5</td>
<td>5.2 ± 0.5</td>
<td>0.1 (-0.1; 0.3)**&lt;br&gt; <strong>p&lt;0.05</strong>&lt;br&gt; <strong>p&lt;0.05</strong>&lt;br&gt; <strong>p&lt;0.05</strong>&lt;br&gt; <strong>p&lt;0.05</strong></td>
</tr>
<tr>
<td>CaO₂ (mL/dL)</td>
<td>14.0 ± 1.5</td>
<td>13.2 ± 1.7</td>
<td>0.8 (0.4; 1.2)**&lt;br&gt; <strong>p&lt;0.05</strong>&lt;br&gt; <strong>p&lt;0.05</strong>&lt;br&gt; <strong>p&lt;0.05</strong>&lt;br&gt; <strong>p&lt;0.05</strong></td>
</tr>
<tr>
<td><strong>Inflammation markers</strong>&lt;br&gt; C-reactive protein (mg/L)</td>
<td>7.9 ± 6.3</td>
<td>7.5 ± 6.1</td>
<td>0.4 (0.0; 0.8)**&lt;br&gt; <strong>p&lt;0.05</strong>&lt;br&gt; <strong>p&lt;0.05</strong>&lt;br&gt; <strong>p&lt;0.05</strong>&lt;br&gt; <strong>p&lt;0.05</strong></td>
</tr>
<tr>
<td>Pentraxin-3 (ng/mL)</td>
<td>1.89 ± 0.83</td>
<td>2.08 ± 1.36</td>
<td>1.5 (-0.2; 3.1)**&lt;br&gt; <strong>p&lt;0.05</strong>&lt;br&gt; <strong>p&lt;0.05</strong>&lt;br&gt; <strong>p&lt;0.05</strong>&lt;br&gt; <strong>p&lt;0.05</strong></td>
</tr>
<tr>
<td><strong>Endothelial activation markers</strong>&lt;br&gt; Angiopoietin-1 (ng/mL)</td>
<td>14.1 ± 6.8</td>
<td>14.4 ± 5.7</td>
<td>0.7 (-5.1; 8.2)**&lt;br&gt; <strong>p&lt;0.05</strong>&lt;br&gt; <strong>p&lt;0.05</strong>&lt;br&gt; <strong>p&lt;0.05</strong>&lt;br&gt; <strong>p&lt;0.05</strong></td>
</tr>
<tr>
<td>Angiopoietin-2 (ng/mL)</td>
<td>10.4 ± 3.5</td>
<td>10.4 ± 3.5</td>
<td>-0.1 (-0.5; 0.3)**&lt;br&gt; <strong>p&lt;0.05</strong>&lt;br&gt; <strong>p&lt;0.05</strong>&lt;br&gt; <strong>p&lt;0.05</strong>&lt;br&gt; <strong>p&lt;0.05</strong></td>
</tr>
<tr>
<td>Angiopoietin-2:1 ratio</td>
<td>1.0 ± 0.6</td>
<td>1.0 ± 0.6</td>
<td>0.4 (-0.07; 0.85)**&lt;br&gt; <strong>p&lt;0.05</strong>&lt;br&gt; <strong>p&lt;0.05</strong>&lt;br&gt; <strong>p&lt;0.05</strong>&lt;br&gt; <strong>p&lt;0.05</strong></td>
</tr>
<tr>
<td>vWF (%):</td>
<td>158 ± 43</td>
<td>141 ± 45</td>
<td>2 (-12; 16)**&lt;br&gt; <strong>p&lt;0.05</strong>&lt;br&gt; <strong>p&lt;0.05</strong>&lt;br&gt; <strong>p&lt;0.05</strong>&lt;br&gt; <strong>p&lt;0.05</strong></td>
</tr>
</tbody>
</table>

Data are presented as unadjusted means ± SD. **Dialysis treatment effects are obtained from linear mixed effects models including a random intercept, or a random intercept and slope (hematocrit, CRP, Pentraxin-3, and Angiopoietin-2:1 ratio), and presented as estimated mean difference (95% CI).**

*Abbreviations: CaO₂ arterial oxygen content; MWP, mean arterial pressure; vWF, von Willebrand factor.*
Table 4

Associations of Hemodialysis- and Oxygenation-related factors, and of Inflammation and Endothelial activation markers with NIRS-\(rSO_2\) (left panel), and frontal gray matter PET-CBF (right panel)

<table>
<thead>
<tr>
<th>Estimated effect on frontal (rSO_2) (%)</th>
<th>Estimated effect on frontal gray matter CBF (mL/100g/min)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Interaction with time present ‡</td>
<td>Effect at T2, as compared to T1</td>
</tr>
<tr>
<td></td>
<td>Effect at T3, as compared to T1</td>
</tr>
<tr>
<td>Interaction with time present ‡</td>
<td>Effect at T2, as compared to T1</td>
</tr>
<tr>
<td></td>
<td>Effect at T3, as compared to T1</td>
</tr>
<tr>
<td>Effect at T1, as compared to T2, as</td>
<td>Effect at T2, as compared to T1</td>
</tr>
<tr>
<td></td>
<td>Effect at T3, as compared to T1</td>
</tr>
</tbody>
</table>

Hemodialysis-related factors:

- MAP (mmHg)  
- Tympanic temperature (°C)  
- pH  
- Ht (per 0.1 mmol/L)  
- UF volume (L)  
- UF rate (mL/h/kg)

Oxygenation-related factors:

- \(pO_2\) (kPa)  
- \(pCO_2\) (kPa)  
- \(CaO_2\) (mL/dL)  
- Angiopoietin 2 (ng/mL)  
- Angiopoietin 2:1 ratio  
- vWF (%)

Endothelial activation markers:

- Markers of inflammation (pentraxin-3, and CRP) were not associated with \(rSO_2\) or CBF.
- The analysis of mean arterial pressure and CBF was inconclusive due to patient variation and missing values.

The associations of individual characteristics on \(rSO_2\) and frontal gray matter CBF that were significant are presented.

- \(* P < 0.05\)  
- \(** P < 0.01\)  
- \(*** P < 0.001\)

† For these characteristics, no interaction was present, meaning that the association of the characteristic with \(rSO_2\) or CBF is similar for all time points. For example, a 1 point higher Angiopoietin 2:1 ratio is associated with a 5% lower \(rSO_2\), independent of T1, T2 or T3.

‡ An interaction with time means that the association of the characteristic with \(rSO_2\) or CBF is different per time point, as compared to T1. For example, a 0.1 higher pH is associated with a 5% lower \(rSO_2\) at T2, and 7% higher at T3, as compared to T1, which means that the association of the characteristic with \(rSO_2\) is different per time point, as compared to T1. For example, a 0.1 higher pH is associated with a 5% lower \(rSO_2\) at T2, and 7% higher at T3, as compared to T1.

§ The analysis of mean arterial pressure and CBF was inconclusive due to patient variation and missing values.

The associations were studied using linear mixed effects models including a random intercept, or a random intercept and slope, whether appropriate according to the likelihood-ratio test.

The estimated effects (95% CI) of the individual characteristics on \(rSO_2\) and frontal gray matter CBF that were significant are presented.

- \(* P < 0.05\)  
- \(** P < 0.01\)  
- \(*** P < 0.001\)

Markers of inflammation (pentraxin-3, and CRP) were not associated with \(rSO_2\) or CBF.

Markers of hemodialysis and oxygenation-related factors, and of inflammation and endothelial activation markers with NIRS.

Table 4

Associations of Hemodialysis- and Oxygenation-related factors, and of Inflammation and Endothelial activation markers with NIRS-\(rSO_2\) (left panel), and frontal gray matter PET-CBF (right panel)
Abbreviations: CaO, arterial oxygen content; Ht, hematocrit; MAP, mean arterial pressure; NA, not available; n.s., not significant; rSO, regional oxygen saturation; UF, ultrafiltration; vWF, von Willebrand factor.
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Supplementary METHODS

Additional information on Dialysis Settings
All patients were on bicarbonate dialysis with a low-flux polysulfone hollow-fiber dialyzer (F8; Fresenius Medical Care, Bad Homburg, Germany). Blood flow and dialysate flow rates were 200-300 and 500 mL/min, respectively. Dialysate temperature was 36.5°C in all patients. We used constant UF rate and dialysate conductivity. Dialysate composition was sodium 139 mmol/L, potassium 1.0 or 2.0 mmol/L depending on the prevailing plasma potassium, calcium 1.5 mmol/L, magnesium 0.5 mmol/L, chloride 108 mmol/L, bicarbonate 34 mmol/L, acetate 3.0 mmol/L, and glucose 1.0 g/L. The water for hemodialysis complied with the requirements of the European Pharmacopoeia (<100 colony-forming units/mL; <0.25 endotoxin units/mL).

Additional information on Image Reconstruction and Preprocessing
We used the 3D T2-FLAIR images for the registration process, because the 3D acquisition of the T1-weighted sequence was not available. Furthermore, several patients had marked brain atrophy and white matter lesions. Therefore, we used the population-based gray matter/white matter (GM/WM) maps to segment the cortical tissue, instead of using the subject probability maps. This means that the cortical volumes of interest (VOIs) are slightly larger than when the individual maps for the subject are used. Since we did the modeling in the subject brain space (no deformations to adjust to the atlas) and the VOIs were based on the population-based GM/WM probabilities, the effect of the atrophy and lesions is expected to be minimal.
Supplementary Figure S1. Scatter plots of Δ Cerebral Blood Flow (X-axis) and Δ Regional Oxygen Saturation (Y-axis) calculated between T2 and T3 displayed per left (S1A) and right (S1B) hemisphere. Correlation coefficient for the left hemisphere: \( \rho = 0.64 \) (\( P = 0.048 \)), and the right hemisphere: \( \rho = 0.76 \) (\( P = 0.01 \)).
Fig S2. Bland–Altman plot of changes in \( \Delta rSO_2 \) (\( \Delta rSO_2 \)) and in CBF (\( \Delta CBF \)) between T2 (early after start of hemodialysis) and T3 (at the end of hemodialysis), displayed for the left (S2A) and right (S2B) hemisphere. The X-axis represents \( \Delta CBF \) (%), while the Y-axis represents the difference between \( \Delta CBF \) and \( \Delta rSO_2 \). The central solid line represents zero bias. The central dashed line indicates overall bias, which was not significant (left hemisphere: \(-6\%\), \( P = 0.2 \); right hemisphere: \(-8\%\), \( P = 0.2 \)). The upper and lower dashed lines represent limits of agreement: \(-35\%\) and \(22\%\) for the left and \(-33\%\) and \(17\%\) for the right hemisphere. The diagonal solid line represents proportional bias. Linear regression modeling suggested the presence of proportional bias (left hemisphere: \( P = 0.001 \), regression equation: \( Y = -0.5 + 0.7x \); right hemisphere: \( P = 0.01 \), regression equation: \( Y = -0.01 + 0.07x \)). This suggests that NIRS increasingly underestimated \( \Delta CBF \) with larger changes.
Supplemental Figure S3: Scatter plots of Δ Cerebral Blood Flow (X-axis) and Δ Regional Oxygen Saturation (Y-axis) calculated between T2 and T1, displayed per left (S3A) and right (S3B) hemisphere. Correlation for the left hemisphere: ρ = 0.09 (P = 0.4) and the right hemisphere: ρ = 0.21 (P = 0.6).
Supplemental Figure S4. Bland-Altman plot of changes in rSO\textsubscript{2} (ΔrSO\textsubscript{2}) and in CBF (ΔCBF) between T1 (before start of hemodialysis) and T2 (early after start of hemodialysis), displayed for the left (S4A) and right (S4B) hemisphere. The X-axis represents ΔCBF (%), while the Y-axis represents the difference between ΔCBF and ΔrSO\textsubscript{2}. The upper and lower dashed lines represent limits of agreement: 1.8% and 2.7% for the left and -2.3% and 3.1% for the right hemisphere, respectively. Linear regression suggested the presence of proportional bias (left hemisphere: P=0.004, regression equation: Y=0.004x+7.0; right hemisphere: P=0.002, regression equation: Y=0.002x+4.0). The central solid line represents zero bias. The central dashed line indicates overall bias, which was not significant (left hemisphere: 4%, P=0.2; right hemisphere: 3%, P=0.4). The upper and lower dashed lines represent limits of agreement: 1.8% and 2.7% for the left and -2.3% and 3.1% for the right hemisphere, respectively. Linear regression suggested the presence of proportional bias (left hemisphere: P=0.004, regression equation: Y=7.0+1.3x; right hemisphere: P=0.002, regression equation: Y=4.0+1.3x), which was not significant (left hemisphere: 4%, P=0.2; right hemisphere: 3%, P=0.4). The upper and lower dashed lines represent limits of agreement: 1.8% and 2.7% for the left and -2.3% and 3.1% for the right hemisphere, respectively. Linear regression suggested the presence of proportional bias (left hemisphere: P=0.004, regression equation: Y=7.0+1.3x; right hemisphere: P=0.002, regression equation: Y=4.0+1.3x), which was not significant (left hemisphere: 4%, P=0.2; right hemisphere: 3%, P=0.4).
Supplemental Figure S5. Scatter plots of frontal gray matter CBF (x-axis) and regional oxygen saturation (y-axis) at T1, T2, and T3, and displayed per left and right hemisphere. No significant correlations were found.
**Supplemental Table S1**

Associations of Hemodialysis- and Oxygenation-related factors, and of Inflammation and Endothelial activation markers with NIRS-rSO$_2$ (left panel), and frontal gray matter PET-CBF (right panel).

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>T2</th>
<th>Characteristic</th>
<th>T3</th>
</tr>
</thead>
<tbody>
<tr>
<td>Estimated effect on frontal rSO$_2$ (%)</td>
<td>Overall effect</td>
<td>Characteristic</td>
<td>T2</td>
</tr>
<tr>
<td>Mean arterial pressure (mmHg)</td>
<td>None</td>
<td>None</td>
<td>None</td>
</tr>
<tr>
<td>Tympanic temperature</td>
<td>None</td>
<td>None</td>
<td>-$2.0\ (0.0; 1.0^*)$</td>
</tr>
<tr>
<td>pH (per 0.1 change)</td>
<td>7.4 (6.7; 1.4)</td>
<td>7.4 (6.7; 1.4)</td>
<td>7.4 (6.7; 1.4)</td>
</tr>
<tr>
<td>Hematocrit (per 0.1 mmol/L)</td>
<td>2.0 (1.0; 3.0)</td>
<td>2.0 (1.0; 3.0)</td>
<td>2.0 (1.0; 3.0)</td>
</tr>
</tbody>
</table>

Estimated effect on frontal gray matter CBF (mL/100g/min)

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>T2</th>
<th>Characteristic</th>
<th>T3</th>
</tr>
</thead>
<tbody>
<tr>
<td>Estimated effect on frontal gray matter CBF (mL/100g/min)</td>
<td>Overall effect</td>
<td>Characteristic</td>
<td>T2</td>
</tr>
<tr>
<td>Mean arterial pressure (mmHg)</td>
<td>None</td>
<td>None</td>
<td>None</td>
</tr>
<tr>
<td>Tympanic temperature</td>
<td>None</td>
<td>None</td>
<td>-$2.0\ (0.0; 1.0^*)$</td>
</tr>
<tr>
<td>pH (per 0.1 change)</td>
<td>7.4 (6.7; 1.4)</td>
<td>7.4 (6.7; 1.4)</td>
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</tr>
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<td>Hematocrit (per 0.1 mmol/L)</td>
<td>2.0 (1.0; 3.0)</td>
<td>2.0 (1.0; 3.0)</td>
<td>2.0 (1.0; 3.0)</td>
</tr>
</tbody>
</table>

**Notes:**
- *P < 0.05, **P < 0.01, ***P < 0.001.
- The estimated effect (95%CI) of the individual characteristics on frontal rSO$_2$ and frontal gray matter is presented.
- The models with an interaction could be interpreted by adding the overall effect to the interaction effect, e.g. a 0.1 pH increase is associated with a ($-1.2 + 6.2 = 5\%$) rSO$_2$ increase at T2, and a ($-1.2 + 8.4 = 7.2\%$) rSO$_2$ increase at T3.
- Associations were studied using linear mixed effect models including a random intercept, or a random intercept and slope, whether appropriate according the likelihood-ratio test.
- Hemoglobin concentrations (ng/mL) of the individual characteristics on frontal rSO$_2$ and frontal gray matter were presented.
- p-values for the interaction model could not be computed due to the small number of participants.

**Legend:**
- N/A, not available; rSO$_2$, regional oxygen saturation.