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Published in:
American journal of physiology-Regulatory integrative and comparative physiology

DOI:
10.1152/ajpregu.00251.2013

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Document Version
Publisher's PDF, also known as Version of record

Publication date:
2013

Link to publication in University of Groningen/UMCG research database

Citation for published version (APA):

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High-saturated fat-sucrose feeding affects lactation energetics in control mice and mice selectively bred for high wheel-running behavior


1Center for Behavior and Neurosciences (CBN), Unit Neuroendocrinology, University of Groningen, The Netherlands; 2Center for Isotope Research, Energy and Sustainability Research Institute Groningen, University of Groningen, The Netherlands; 3CBN, Behavioral Biology Group, University of Groningen, Groningen, The Netherlands; and 4Department of Biology, University of California, Riverside, California

Submitted 17 May 2013; accepted in final form 28 September 2013

Guidotti S, Jónás I, Schubert KA, TG, Meijer HA, Scheurink AJ, van Dijk G. High-saturated fat-sucrose feeding affects lactation energetics in control mice and mice selectively bred for high wheel-running behavior. Am J Physiol Regul Integr Comp Physiol 305: R1433–R1440, 2013. First published October 2, 2013; doi:10.1152/ajpregu.00251.2013.—Feeding a diet high in fat and sucrose (HFS) during pregnancy and lactation is known to increase susceptibility to develop metabolic derangements later in life. A trait for increased behavioral activity may oppose these effects, since this would drain energy from milk produced to be made available to the offspring. To investigate these interactions, we assessed several components of behavioral energetics during lactation in control mice (C) and in mice of two lines selectively bred for high wheel-running activity (S1, S2) subjected to a HFS diet or a low-fat (LF) diet. Energy intake, litter growth, and milk energy output at peak lactation (MEO; assessed by subtracting maternal metabolic rate from energy intake) were elevated in HFS-feeding dams across all lines compared with the LF condition, an effect that was particularly evident in the S dams. This effect was not preceded by improved lactation behaviors assessed between postnatal days 1 and 7 (PND 1–7). In fact, S1 dams had less high-quality nursing, and S2 dams showed poorer pup retrieval than C dams during PND 1–7, and S dams had generally higher levels of physical activity at peak lactation. These data demonstrate that HFS feeding increases MEO underlying increased litter and pup growth, particularly in mice with a trait for increased behavioral physical activity.

lactation; maternal behavior; maternal diet; metabolism; milk energy output; physical activity

PHYSICAL ACTIVITY LEVELS CAN VARY greatly between, as well as within animal species (11). Clearly, differences in physical activity can have major repercussions for energy balance regulation and fuel homeostasis. One approach to study this is to compare these regulations in control mice with those in mice from lines that have been selectively bred over multiple generations for increased running-wheel behavior. Activity-selected mice, and, in particular, females, have relatively low body masses, reduced body fat, increased mass-adjusted food consumption, increased daily energy expenditure, and a high maximal rate of oxygen consumption during forced treadmill exercise. These traits are expressed even if the mice are not allowed access to running wheels (12, 34, 35, 46). On a high-fat diet (with or without sucrose), the “physical activity” trait protects these mice against diet-induced obesity (DIO), although they are markedly hyperphagic compared with control mice that do become obese (46). The DIO resistance of highly active mice is, in part, attributable to a diet-induced augmentation of spontaneous locomotor activity and metabolic/endocrine changes that stimulate fuel metabolism (46). These effects are most pronounced in activity-selected females, pointing toward a sex-specific penetration of the physical hyperactivity trait (12).

While exercise training per se does not seem to negatively impact offspring growth and development (6, 21), a hyperactivity trait and associated increased metabolic rate (MR) when subjected to a high fat-sucrose (HFS) diet could be potential constraints to reproduction. Reproduction requires energy allocation to developing offspring, and this is particularly evident in lactating rodents (28). For this reason, we have studied behavioral energetics of lactation in control mice and in mice selectively bred for high running-wheel behavior (43) subjected to a standard low-fat (LF) control diet, or a HFS diet. Energetic costs of lactation in rodents can increase approximately four-fold above normal energy requirements (20). The maximum energy consumption and expenditure during this period—termed sustainable energy intake and sustainable metabolic rate, respectively—are probably limited intrinsically by aspects of physiology, and this concept has been outlined by Król and colleagues (26, 28, 42). When subjected to HFS feeding, mice during lactation may be channeling more energy in milk toward their offspring, thus leading to larger weight gain of pups compared with LF-feeding dams (47), although this has never been directly tested to our knowledge. We hypothesized that this effect would be limited in activity-selected mice, as HFS feeding in these mice may cause behavioral and metabolic effects that would drain energy away from offspring growth and development. This would be considered a “peripheral” limiting factor to offspring development (25).

MATERIALS AND METHODS

Animals and housing. Mice from lines selectively bred for voluntary wheel-running behavior (the base population was the Hsd:ICR strain) were obtained from T. Garland Jr., Riverside, CA. Originally, eight mouse lines were created (four selected and four control) (43).
Ten pairs from each of two selectively bred lines—here referred to as S1 and S2, but originally designation lines 7 and 8, respectively (17)—and from one randomly bred control line—here referred to as C, but originally designated line 2—were shipped to Groningen. These mice were in their 45th generation of selection upon arrival. We did not study selection lines 3 and 6, both of which exhibit the “minimuscle” phenotype, caused by a Mendelian recessive allele with many pleiotropic effects (10, 18). Some of the differences between S1 and S2 line were previously described by Hannon et al. (17). The lines were maintained at the animal facility of the Center for Behavior and Neurosciences in Groningen with further selection for wheel-running activity (until generation 52), with males separated from females after weaning and typically housed with three or four mice per cage (Macrolon Type II UNO Roestvaststaal BV, Zevenaar, NL) with ad libitum access to water and a fibered low-fat (LF) standard laboratory mouse chow (15.9 kJ/g; 62% carbohydrate, 13% fat, 24% protein; RMH-B 2181, HopeFarms BV, Woerden, NL) at 22 ± 1°C, and maintained on a 12:12-h light-dark cycle with lights on at 8 AM. Pine shavings and EnviroDry were used as bedding material. For the entire study, mice had no access to running wheels. All methods were approved by the Institutional Animal Use and Care Committee of the University of Groningen.

Virgin C, S1, and S2 female mice of 4.5 mo old weighing 35.2 ± 1.24 g, 27.1 ± 0.5 g, and 32.9 ± 0.9 g, respectively, were paired with males of corresponding lines. One week before pairing, half of each group was switched from the LF diet to a home-made HFS diet (19.4 kJ/g; 30% carbohydrate, 45% fat, and 24% protein). After 2 wk of pairing, males were removed, and females were left to deliver and raise their litters until PND 21. The presence of new-born litters was checked daily at onset and at the end of the light phase and designated as PND 0. Litters were not disturbed before PND 3 to allow formation of maternal olfactory bonding (45). From PND 3 onward, dams and litters were weighed, and litter sizes were counted.

Maternal behavior/pup-retrieving test. During PND 1–7, maternal care was assessed daily by two observers performing direct observations, as well as by observations from videotapes made from the dams and litters. The videotapes of all dams and litters were available and checked regularly to ensure consistency among observers. Maternal care was assessed at circadian time (CT) 3, CT9, and CT13, each period consisting of 100 min. Within each period, behaviors of dams were scored every 4 min, according to Champagné et al. (3), and adapted to our study. It discriminated the following categories: 1) Dam is not in contact with any pup. Contact with the tail alone is not considered to be “in contact”; 2) dam is in contact with three or more pups. She is with them but not in any defined nursing posture. If only the dam’s tail is touching the pups, then she is considered to be in contact; 3) dam is retrieving one or more pups in her mouth and depositing it/them somewhere else; 4) dam is licking/grooming one or more pups; 5) one or more pups are away from the rest of the litter (split litter). The pups may or may not be clustered together in piles; 6) dam is licking/grooming one or two isolated pups; 7) dam is engaged in passive nursing (i.e., lying on her side or back) with at least 3 pups attached to nipples; and 8) dam is engaged in arched back nursing (ABN) 1–4; this is subdivided into four different positions with an increasing degree of active nursing behavior. This can range from ABN1 (the dam is lying flat on top of the pups) to ABN4 (back of the dam is arched maximally to give space allowing suckling).

Above-mentioned behavioral categories were also analyzed after merging them into “in contact” (total scores from 2, 4, 7, 8) and “not in contact” (total scores from 1, 3, 5, 6).

To test maternal behavioral responsiveness to standardized conditions, dams and litters were subjected to a pup-retrieving trial at PND 3 (4, 14). The test was performed in the middle of the light phase. Specifically, at the start of each trial, the dam was separated from her pups by placing her into another clean cage. The pups were then taken gently from the nest, while using odorless gloves and placed in the opposite corner of the cage. The distance between nest and new location of the pups was kept standard at 21 cm. After reintroducing the dam back to the empty nest, the latency to retrieve each pup was recorded, as well as the number of times the dam approached and then moved away from the displaced litter without retrieving a pup. A “move-away” was scored each time the dam was within 1 cm of a displaced pup and then moved away by a distance greater than her body length (excluding tail). The trial had a cut-off time of 10 min. It was noted when the dam started building a new nest at the location of the displaced pups. Pups not returned to the original nest at the end were returned by the observer.

Mother and offspring characteristics from parturition to weaning. Maternal body mass was assessed on the day prior to breeding and weekly from pairing to parturition. Right after parturition [postnatal day 0 (PND 0)], litter size was determined by outside inspection (no handling) when the dam was not in the nest. Litter characteristics are presented in Fig. 1. Following parturition, litter size and the mass of the litters were assessed, and mean pup mass was calculated. Measurements were done at PND 3, 8, 13–15. Because maternal stress may play an important role in perinatal programming (33), particular care was taken to disturb the dams and litters as little as possible, and, when it was necessary, in a standardized manner.

Food and water intake was assessed daily throughout lactation. Bedding was also checked for spilled or crumbled food (23). At peak lactation (between postnatal day 13 and 15), feces produced by the female and her offspring were collected. The energy content in dried, homogenized feces and food was determined using a bomb calorimeter CBB 330, standard benzoic acid 26.44 kJ/g, BCS-CRM No.90N (Boom BV, Meppel, The Netherlands).

Absorbed energy intake (AEI) of mothers at peak lactation was calculated from the difference between energy intake and fecal energy content. During PND 13–15, MR of dams in their home cages was estimated using the doubly labeled water (DLW) method (41). Milk energy output (MEO) was calculated by the difference between AEI and MR (27). 2H2O (>99.9%) water was provided by Sigma-Aldrich (Zwijndrecht, The Netherlands) and 18O (≥98%, specified to 0.1%) by RÖTENDEUROM (Rehovoth, Israel).

Mice were weighed to the nearest 0.1 g and injected intraperitoneally with a −0.12 g of enriched water (66.6% 18O, 33.3% 2H). The precise injectant weight was weighed to the nearest 0.0001 g before and after injection. After 1 h of equilibration, animals were bled at the tail tip, and two initial blood samples (2 μl) were collected in duplicate glass capillary tubes, which were immediately flame-sealed with an isobutane torch. A final blood sample was taken 48 h after the collection of the initial.

At PND 15, mice were subjected to indirect calorimetry over 3 h in the middle of the light phase to assess respiratory quotient (RQ); the latter was needed for the DLW analysis. Each animal was placed in a 2-liter Plexiglas metabolic chamber for 3 h. Bedding taken from their home cage was provided to reduce stress of novelty. During this period, females were separated from their litters, which remained in their home cage. Specifically, measurements included O2 consumption (VO2, ml/h) and CO2 production (VCO2, ml/h) with an open-flow system (32) with inlet airflow set at 20 l/h (Brooks Type 5850 mass flow controller; Rijswijk, The Netherlands). We calculated RQ as VCO2/VO2. During this time, voluntary activity of dams was assessed by a passive infrared (PIR) detection system (46). For clarity, the chronological order of the procedures mentioned above is presented in a time line shown in Fig. 2.

Analysis of DLW samples. Determinations of 3H/H2O and 18O/H2O ratios in blood samples were performed at our Center for Isotope Research. A detailed description of the analytical procedures followed in our laboratory has been described previously (15). Briefly, samples were prepared by microdistillation (cryodistillation) at vacuum, first heating broken capillaries and cryogenically trapping the emerging water vapor with liquid nitrogen. Water samples were stored in glass vials and automatically injected into a high-temperature pyrolysis unit (Hekatech, Wegberg, Germany) (13), in which the injected water reacted with glassy carbon at 1420°C. The resultant H2 and CO gases, emerging into a continuous helium flow through the system,
were then led through a gas chromatography column to separate the two gases in time and finally fed into an isotope ratio mass spectrometer (GV Instruments IsoPrime, Manchester, UK) for analysis of $\delta^18$O and $\delta^2$H. Measurements were corrected for memory effects using an algorithm described by Guidotti et al. (15).

Body water pools were estimated by the intercept (ic) method (5) from the $\delta^{18}$O(NO) dilution space:

$$E_{\text{ini-ic}} = (E_{\text{ini}} - E_{\text{bkg}})e^{k_i(t_{\text{ini}}-t_{\text{inj}})} + E_{\text{bkg}}$$

$$N_i = (E_{\text{ini-ic}} - E_{\text{inj}})$$

where $E_{\text{ini}}$ and $E_{\text{bkg}}$ are the enrichments at equilibrium and at background in the body, and $t_{\text{ini}}-t_{\text{inj}}$ is the time elapsed between injection of the DLW and sampling of the initial enrichment. $E_{\text{inj}}$ is the enrichment of the injected water. $E_{\text{ini-ic}}$ is the initial value extrapolated from the $E_{\text{ini}}$ back to the time of the injection (i.e., using the ic method) and $k_i$ is the $\delta^{18}$O turnover rate given in h$^{-1}$. $E$ is expressed in parts per million. Final $^2$H and $\delta^{18}$O dilution spaces were calculated from the final body mass, assuming the same percentage of body mass as measured for the initial dilution spaces, and $k_i$ is the $^2$H or $\delta^{18}$O turnover rate is given in h$^{-1}$.

Isotope turnover rates were calculated as follows:

$$k = \frac{\log_e(E_{\text{ini}} - E_{\text{bkg}}) - \log_e(E_{\text{fin}} - E_{\text{bkg}})}{t}$$

where $E_{\text{fin}}$ represents the enrichment level of the final sample. Rate of CO$_2$ production for each animal was calculated according to Speakman (41) using a single-pool model equation (Eq. 7.17). CO$_2$ produc-

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**Fig. 1.** Dam weight (A), litter weight (B), and litter size (C) until end of peak lactation for control [C; low-fat (LF), $n = 8$; high-fat and high sucrose (HFS), $n = 7$] and highly active lines (S1: LF, $n = 6$; HFS, $n = 7$; S2, LF $n = 8$; HFS, $n = 8$) of mice. Values are expressed as means ± SE. Open bars represent LF, and closed bars represent HFS. *LF vs. HF, $P < 0.05$.

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**Fig. 2.** Timeline of procedures from start of the experiment until end of peak lactation. PND, postnatal day; DLW, doubly labeled water.
tion was converted to energy expenditure using an equation derived from Weir et al. (48).

Statistical analysis. Repeated-measures ANOVA (RM-ANOVA) was used to analyze differences in body mass and food intake of females throughout lactation. General linear model (GLM) univariate analysis was used to analyze energetics at peak lactation, maternal behavior, and litter characteristics followed by the Tukey post hoc test when appropriate. GLM analysis of covariance was used to assess contribution of maternal body mass (or fat-free mass) when appropriate. To assess litter size effects, we used this as a covariate, and we performed regression analysis with litter size, diet, and line as independent factors. A value of $P < 0.05$ (for two-tailed tests) was considered statistically significant for all tests.

RESULTS

Dams characteristics. At PND 3, S1 dams weighed less than C dams (C vs. S1 $P < 0.001$) with S2 having an intermediate body weight (C vs. S2 $P < 0.05$). Over the period from PND 3 to PND 15, dams on the HFS diet gained less mass than the ones on the LF diet but started at PND 3 with a higher body mass [time × diet: $F(5,190) = 2.81, P < 0.05$], and S lines gained less weight than C dams, again with S2 being intermediate [line: $F(2,38) = 12.88, P < 0.0001$, see Fig. 1A]. From PND 1 to PND 15, daily food intake (daily data not shown) of lactating females was increasing over time [time: $F(14,532) = 74.29, P < 0.0001$] with S1 and S2 dams overall eating less food than C dams [line: $F(2,38) = 23.65, P < 0.0001$]. Assessment of cumulative energy intake over PND 1–7 (Fig. 3A) revealed that HFS feeding increased energy intake in the C line but not in the S1 and S2 lines during PND 1–7 [line × diet: $F(2,38) = 4.33, P < 0.05$]. During PND 8–15 (Fig. 3B), increased cumulative energy intake was observed in all lines when given HFS diet [time: $F(1,38) = 37.74; P = 0.0001$], with S lines having lower energy intake [line: $F(2,38) = 17.47; P < 0.0001$], but without an interaction between line and diet. Those effects were present even if dam body mass was included as a covariate.

Litter characteristics. Dams included in the study had litters varying between 6 and 15 pups (one S1 dam with a litter size of two was excluded to avoid stretching and skewing). Although litter size of S1 dams was slightly lower than in C and S2 dams, this effect did not reach significance [line: $F(2,38) = 2.47, P = 0.098$]. Over PND 1–15, pups were found missing in all lines [time: $F(14,532) = 8.28, P < 0.0001$], but S1 dams tended to lose fewer pups than C and S2 dams during the first days of lactation [time × line: $F(28,532) = 1.42; P = 0.076$] (Fig. 1C).

At PND 3, HFS-feeding dams had heavier litters than LF-feeding dams [diet: $F(1,38) = 7.38, P < 0.01$] and S1 and S2 dams had lighter litters than C dams [line: $F(2,38) = 10.00, P < 0.001$]; however, no interaction between line and diet was observed. Further testing showed that HFS feeding increased litter weight only in the C line ($P < 0.05$). During PND 3–15, LF litters grew less in S1 and S2 lines than in the C line (S2 being intermediate), an effect that was attenuated in the HFS condition, but particularly so in S lines [time × line × diet: $F(8,152) = 2.62, P < 0.01$] (Fig. 1B). Average pup mass per litter was only affected by a time × diet interaction [$F(4,152) = 2.703, P < 0.0001$] (data not shown).

Maternal behavior. S1 dams showed a higher percentage of side nursing [$F(2,38) = 3.29, P < 0.05$] and ABN1 [$F(2,38) = 4.80, P < 0.05$] than S2 and C dams, and a lower percentage of ABN3 [$F(2,38) = 10.42, P < 0.01$] with S2 dams being intermediate (summary data can be found in Table 1). All lines declined “in contact with the litter” from PND 1 to PND 7 [$F(6,228) = 57.95, P < 0.0001$]. However, this decline in litter contact was less pronounced in S1 dams from PND 4 to PND 7 [time × line: $F(12,228) = 2.29, P < 0.01$], with the HFS feeding dams contributing most to this effect (Fig. 4, A and B).

At PND 3, litters were subjected to a pup-retrieving test. S2 dams showed an impaired performance to retrieve the pups back to their nest with an overall higher average percentage of incomplete trials [line: $F(2,38) = 7.59, P < 0.01$] (Fig. 4C). Among eight S2 dams, only two completed the pup retrieval trial. The inability of S2 dams to successfully retrieve the litter was characterized by an increased number of “move-aways” [line: $F(2,38) = 4.28, P < 0.05$] (Fig. 4D). There were no differences between diet groups, and between lines S1 and C.

Peak lactation. At peak lactation, S1 dams had overall lower body masses than C dams, with S2 dams being intermediate [line: $F(2,38) = 10.73, P < 0.001$]. Body composition of dams was assessed by isotope dilution and by assuming a constant hydration factor of fat-free mass of 0.73 (38). S dams showed a strong trend for a higher body fat content (fat%) and lower percentage fat-free mass (%FFM) than C dams (see Table 2). HFS-feeding dams had overall heavier litters [$F(1,38) = 60.50, P < 0.0001$], and S dams raised lighter litters at peak lactation than C dams, with S2 dams having litters of intermediate mass [$F(2,38) = 27.00, P < 0.0001$]. Average pup mass was increased in litters of HFS-feeding dams [$F(1,38) = 19.01, P < 0.01$]. Average pup mass was not affected by line or line × diet interaction.
Table 1. Maternal behavior

<table>
<thead>
<tr>
<th></th>
<th>LF</th>
<th>HFS</th>
<th>LF</th>
<th>HFS</th>
<th>LF</th>
<th>HFS</th>
</tr>
</thead>
<tbody>
<tr>
<td>In contact</td>
<td>3.71 ± 0.38</td>
<td>4.02 ± 0.65</td>
<td>4.48 ± 0.46</td>
<td>4.88 ± 0.90</td>
<td>5.97 ± 0.68</td>
<td>5.93 ± 1.19</td>
</tr>
<tr>
<td>Not in contact</td>
<td>34.99 ± 2.83</td>
<td>38.09 ± 2.08</td>
<td>37.18 ± 1.16</td>
<td>28.86 ± 3.02</td>
<td>34.64 ± 2.50</td>
<td>33.51 ± 2.15</td>
</tr>
<tr>
<td>Licking/grooming</td>
<td>2.73 ± 0.54</td>
<td>2.35 ± 0.39</td>
<td>2.60 ± 0.49</td>
<td>4.36 ± 0.96</td>
<td>3.49 ± 0.70</td>
<td>3.97 ± 0.67</td>
</tr>
<tr>
<td>Licking 1 pup</td>
<td>0.07 ± 0.04</td>
<td>0.08 ± 0.08</td>
<td>0.00 ± 0.00</td>
<td>0.19 ± 0.10</td>
<td>0.00 ± 0.00</td>
<td>0.09 ± 0.09</td>
</tr>
<tr>
<td>Split litter</td>
<td>0.47 ± 0.21</td>
<td>0.89 ± 0.64</td>
<td>0.25 ± 0.15</td>
<td>0.09 ± 0.07</td>
<td>0.34 ± 0.22</td>
<td>1.03 ± 0.53</td>
</tr>
<tr>
<td>Retrieving</td>
<td>0.17 ± 0.07</td>
<td>0.09 ± 0.04</td>
<td>0.00 ± 0.00</td>
<td>0.15 ± 0.06</td>
<td>0.02 ± 0.02</td>
<td>0.09 ± 0.09</td>
</tr>
<tr>
<td>Side Nursing</td>
<td>2.24 ± 1.19</td>
<td>0.16 ± 0.11</td>
<td>5.84 ± 5.50</td>
<td>12.67 ± 5.90</td>
<td>4.60 ± 2.25</td>
<td>1.30 ± 1.17</td>
</tr>
<tr>
<td>ABN1</td>
<td>1.41 ± 0.31</td>
<td>0.74 ± 0.18</td>
<td>2.79 ± 0.85</td>
<td>2.08 ± 0.49</td>
<td>1.47 ± 0.31</td>
<td>1.47 ± 0.38</td>
</tr>
<tr>
<td>ABN2</td>
<td>24.63 ± 4.02</td>
<td>22.28 ± 1.78</td>
<td>30.84 ± 5.30</td>
<td>24.49 ± 5.73</td>
<td>24.15 ± 4.37</td>
<td>27.77 ± 3.90</td>
</tr>
<tr>
<td>ABN3</td>
<td>22.75 ± 2.77</td>
<td>26.24 ± 1.72</td>
<td>13.55 ± 2.28</td>
<td>16.83 ± 1.56</td>
<td>18.97 ± 1.55</td>
<td>19.65 ± 1.90</td>
</tr>
<tr>
<td>ABN4</td>
<td>6.83 ± 1.81</td>
<td>5.05 ± 1.21</td>
<td>2.48 ± 0.62</td>
<td>5.39 ± 1.82</td>
<td>6.34 ± 1.63</td>
<td>5.18 ± 1.32</td>
</tr>
</tbody>
</table>

Values are expressed as means ± SE. Average percentage of maternal behaviors in control mice (C) and selected lines (S1, S2) feeding low-fat (LF) or high-fat sucrose (HFS) diet during first week after parturition (PND 1–7). ABN1–4, arched back nursing, day 1–4. We assessed maternal care during three periods (each 100 min) of observations per day at CT 3, 9, 13. The total number of observation per day was 75 (every 4 min for 100 min, three times a day). Within each observation period, the behavior of each dam was scored every 4 min for a total of 25 observations per period and a total of 75 observations per dam per day.

Fecal mass, fecal energy content, and absorption efficiency were assessed during peak lactation in a subcohort of dam/litter \( (n = 4) \) groups. Fecal mass, energy absorption (in kJ), and % absorbed energy were not statistically different in C and S lines. Fecal energy content of S2 animals was higher than of C and S1 animals irrespective of diet. Fecal energy content \( F(2,18) = 4.02, P < 0.05 \) and HFS-feeding mice had a higher fecal energy content than LF-feeding mice \( F(1,18) = 8.32, P < 0.01 \), irrespective of line.

Energetic traits during peak lactation (PND 13–16) are shown in Table 2. Total food intake (in grams) was lower in S1 and S2 dams compared with C dams \( F(2,38) = 10.47, P < 0.001 \) irrespective of diet. Energy intake (in kilojoules) was higher in the HFS-feeding dams \( F(1,38) = 30.73, P < 0.0001 \) than LF-feeding dams, and S1 and S2 dams consumed overall less energy than C dams \( F(2,38) = 7.67, P < 0.01 \). MR, measured by the DLW method, was lower in S dams than in C dams \( F(2,38) = 5.09, P < 0.05 \), irrespective of dietary condition. This line difference was not observed when body mass or FFM was used as a covariate. With respect to home cage activity, a line, but not a diet effect was observed when body mass or FFM was used as a covariate. With LF feeding, MEO was markedly lower in S1 and S2 dams than in C dams \( F(2,38) = 25.52, P < 0.01 \).

An overall increased MEO by HFS-feeding \( F(1,38) = 9.08, P < 0.01 \) was observed. Post hoc analysis revealed that S1 offspring had higher PIR activity than C, with S2 having an intermediate phenotype (Table 2).

With LF feeding, MEO was markedly lower in S1 and S2 dams than in C dams \( F(2,38) = 5.18, P < 0.01 \). An overall increased MEO by HFS-feeding \( F(1,38) = 25.52, P < 0.0001 \) was observed above the ones observed in the LF, which was particularly evident in the S lines (Fig. 5). These stronger increases in the S lines (S1: 83%, S2: 62%) than in the C line (24%) caused MEO to be indifferent between the lines.
in the HFS condition. When corrected for body mass, this percent difference in MEO in S1 (90%) and S2 (70%), but not in C dams (18%), became even more pronounced. Statistical outcomes were not affected when using litter size as a covariate. Neither did litter size remain a significant factor in a backward regression analysis, with line and diet as other independent factors. Finally, by post hoc removing cases with relatively high (>14) litters size (2 in C/HFS group; 2 in S2/HFS group) and with relatively low (<8) litter sizes (2 in S1/HFS group), average (10.5–11.7), and median (11–12) litter sizes were exactly similar in all groups, and this did not yield a different pattern of MEO outcomes and neither did it lead to changes in statistical outcomes. This is a strong indication that subtle differences in litter size were not a relevant factor in differently limiting MEO.

**DISCUSSION**

Previous rodent studies have shown that HFS feeding during pregnancy and lactation causes advanced pup growth and development relative to LF feeding (19, 40, 44). This can be associated with increased body length (8); however, increases in lean mass are less obvious (24). The HFS-induced pup growth and development may be dampened by intense behavioral and metabolic activities (37). To study the energetic bases of these effects, we 1) quantified MEO from dams to pups under HFS and LF diet conditions and 2) assessed whether a trait of increased physical activity would impact negatively on MEO.

In the assessment of potential differences in MEO during lactation, it is important to consider nursing behavior of lactating dams. This is particularly important during the first week after parturition, because early maternal behavior is crucial for mother–pup bonding, pup development, and thrill (45). In our analysis, merging several maternal behavioral categories into percentage contact time showed very high (i.e., ~80% of time scored) maternal care during the early postnatal days, and then slowly decreased to levels around 40–50% being “in contact” on postnatal day 7. This decline of dams being “in contact” with their litter has been observed before (2, 39) and probably reflects a reciprocal daily increase in feeding behavior to increase MEO (39). To our surprise, we did not observe differences between HFS-feeding and LF-feeding dams on feeding or related maternal behaviors. Although increased nursing behavior in HFS-feeding dams was anticipated on the basis of data from Purcell et al. (33) in lactating rats, this was not observed in our mice. We did observe, however, that S1 dams performed more low-care nursing behavior than C dams. The reduced level of high-quality nursing appeared to be compensated with overall more time spent with the litter, a situation that was particularly increased when S1 dams were subjected to the HFS diet. Contrary to S1 dams, S2 dams showed similar maternal care as C dams, but performed less well in the pup retrieval test relative to C and S1 dams. Girard et al. (14) did not observe differences in maternal behavior on PND 9 and PND 16 between four C and four S lines, which may indicate that maternal care differences between lines are lost at a later stage of lactation. In conclusion, S1 dams showed subtle impairments in maternal care over the first postpartum week relative to control dams, and S2 dams showed poorer pup retrieval. These effects in the S lines could have been relevant for energy availability to the offspring, pup growth and development, and pup survival at any point of the preweaning period.

Because MEO is maximized under conditions of peak lactation (27) and dams were weight-stable during this period, this allowed us to calculate MEO by subtracting MR from AEI (27). We were able to confirm that MEO at peak lactation was

### Table 2. Peak lactation energetics

<table>
<thead>
<tr>
<th></th>
<th>LF</th>
<th>HFS</th>
<th>LF</th>
<th>HFS</th>
<th>LF</th>
<th>HFS</th>
</tr>
</thead>
<tbody>
<tr>
<td>Food intake, g/day</td>
<td>20.7 ± 1</td>
<td>19 ± 0.8</td>
<td>14.7 ± 1.1#</td>
<td>16.5 ± 1.4</td>
<td>16.1 ± 0.8#</td>
<td>16.7 ± 1.2</td>
</tr>
<tr>
<td>Energy intake, kJ/day</td>
<td>329 ± 15</td>
<td>374 ± 16*</td>
<td>233 ± 18#</td>
<td>325 ± 27*</td>
<td>256 ± 14#</td>
<td>328 ± 24*</td>
</tr>
<tr>
<td>AEI, kJ/day</td>
<td>301 ± 14</td>
<td>346 ± 15*</td>
<td>213 ± 16#</td>
<td>302 ± 25*</td>
<td>229.8 ± 12#</td>
<td>300 ± 22#</td>
</tr>
<tr>
<td>Body weight dam, g</td>
<td>47.1 ± 1.5</td>
<td>49.6 ± 2.8</td>
<td>41.6 ± 0.4#</td>
<td>39.9 ± 1.2</td>
<td>45.9 ± 1.1#</td>
<td>44.1 ± 1.2</td>
</tr>
<tr>
<td>MR*, kJ/day</td>
<td>120.2 ± 6.1</td>
<td>120 ± 3.7</td>
<td>92.7 ± 8.6#</td>
<td>106.3 ± 8.9</td>
<td>106.3 ± 3.4#</td>
<td>99.7 ± 3.2</td>
</tr>
<tr>
<td>RQ, VO₂/CO₂</td>
<td>1.04 ± 0.04</td>
<td>1.02 ± 0.05</td>
<td>1.1 ± 0.05</td>
<td>1 ± 0.04</td>
<td>1.01 ± 0.03</td>
<td>1.02 ± 0.04</td>
</tr>
<tr>
<td>PIR</td>
<td>5.46 ± 0.76</td>
<td>4.72 ± 0.26</td>
<td>9.94 ± 1.5#</td>
<td>8.16 ± 0.92</td>
<td>6.74 ± 1.04</td>
<td>7.67 ± 0.73</td>
</tr>
<tr>
<td>Fat%</td>
<td>8.27 ± 0.59</td>
<td>7.3 ± 0.55</td>
<td>8.82 ± 1.2</td>
<td>9.77 ± 1.55</td>
<td>9.5 ± 1.22</td>
<td>10.79 ± 1.31</td>
</tr>
<tr>
<td>Feces (n = 4)</td>
<td>Total dry weight: 5.852 ± 0.55</td>
<td>5.316 ± 0.3</td>
<td>3.618 ± 0.414</td>
<td>4.744 ± 1.43</td>
<td>4.98 ± 0.67242</td>
<td>4.869 ± 0.758</td>
</tr>
<tr>
<td>Energy density, kJ/g</td>
<td>15.96 ± 0.48</td>
<td>17.63 ± 0.77</td>
<td>16.12 ± 0.82</td>
<td>16.52 ± 0.73</td>
<td>16.9 ± 0.29#</td>
<td>19.1 ± 0.06#</td>
</tr>
<tr>
<td>Energy absorption, %</td>
<td>91.69 ± 1.39</td>
<td>92.42 ± 0.5</td>
<td>91.27 ± 0.567</td>
<td>93.13 ± 1.54</td>
<td>89.95 ± 0.58139</td>
<td>91.59 ± 0.683</td>
</tr>
</tbody>
</table>

Values are expressed as means ± SE. Energetics during peak lactation (postnatal days 13–15) dams from the control line (C) and two selected lines (S1, S2) fed either an LF or an HFS diet. AEI, absorbed energy intake; MR, metabolic rate; RQ, respiratory quotient; PIR, passive infrared. Tukey post hoc test *P < 0.05, diet; #P < 0.05, S1 or S2 vs. C, $P < 0.05, S1 vs. S2. aMeasured by doubly labeled water.
greater in HFS-feeding mice than in LF-feeding mice. To our knowledge, this is the first demonstration using the DLW method to show that HFS-feeding causes an elevation of milk energy availability at peak lactation, which then increases litter mass (1, 9, 16). At least three mechanisms could underlie this effect. As opposed to LF feeding, HFS feeding increases fat and “fast carbohydrate” absorption, which can immediately be converted to milk ingredients without first having to be processed by energy-demanding mechanisms, such as de novo lipogenesis (31, 36). Second, the HFS diet is more energetically dense and, thus, allows dams to derive more energy from it. Third, because dietary fat has a lower postingestive thermic effect than complex carbohydrates (7) and because energy intake may be constrained by heat loss under conditions of peak lactation (28), sustainable energy intake could have been elevated more on the HFS diet than on the LF diet without risk of overheating. The explanation that subtle differences in litter size played a role in affecting MEO is not likely because inclusion of litter size as a cofactor did not contribute to differences in MEO.

When viewing the mouse lines separately, S1 and S2 dams had markedly lower levels of MEO than C dams under LF feeding conditions. Contrary to our expectation, however, this was not caused by a higher MR in S1 and S2 dams, analogous to what we previously observed under nonlactating conditions in S females (46), but rather by a lower level of food intake by S dams compared with C dams. This effect appeared to be body (lean) mass-related. As a result of the lower MEO, litter masses of S dams were lower than those of C dams (Fig. 1B). However, MEO in S dams showed an increase upon HFS-feeding relative to LF-feeding (S1 +83%, S2 +62%), which was much stronger than in C dams subjected to HFS diet (+24%). When expressed per unit body mass, this effect was further amplified in S relative to C dams. A HFS-induced increase in food intake that was stronger in S dams than in C dams combined with an unresponsively low MR contributed to this effect. A potential mechanism may be related to thermoregulatory effects mentioned above. Thus, although we did not directly assess thermoregulation in the present study, it may be speculated that S dams were able to transmit more heat to the environment than C dams in the HFS condition, potentially because of a higher surface-to-volume ratio (22). In turn, this would cause sustainable energy intake to be limited at a higher level in the S dams (26–28). Increased food intake in HFS-feeding S dams, however, was only observed during PND 8–PND 15, while HFS-induced increases in food intake in C dams were observed during PND 1–PND 7, as well as PND 8–PND 15. Thus, although MEO in C and S dams were indistinguishable under the HFS feeding condition at peak lactation (PND 13–PND 15), body masses of S litters did not reach those of C litters, presumably because of the hyperphagia unresponsiveness of the S dams to the HFS diet during PND 1–7. Indeed, the HFS diet increased litter weight in the C line from PND 3 onward, whereas this effect could be observed in the S lines only from postnatal day PND 8 onward. The mechanism underlying this unresponsiveness is not known, but it seems to fit the overall lower level of quality nursing and reduced pup retrieval by S1 and S2 dams, respectively, as shown in our behavioral analysis over PND 1–7.

In conclusion, HFS feeding during lactation clearly causes an increase in MEO from dams to the offspring. Combined with findings of Wahlig et al. (47), showing that milk of HFS-feeding mice is richer in fat than that of LF-feeding dams, this could collectively underlie increased litter growth. The increased MEO in such a critical window not only allows increased growth and development in the offspring, but, at the same time, could increase the susceptibility in the offspring to develop metabolic derangement later in life, such as increased adiposity and insulin resistance (1, 9, 16, 29, 47). In fact, in a follow-up study, we observed that the preweaning HFS diet increased the adult proneness to diet-induced metabolic derangements irrespective of line (Guidotti S, Meyer N, Scheurink A, Garland Jr T, and van Dijk G, unpublished data).

Mice selectively bred for increased running-wheel behavior showed impaired MEO under LF condition, but this was surprisingly increased to the same level as controls when subjected to HFS feeding. This contrasts sharply with their behavioral energetics under nonreproductive conditions, during which HFS feeding augments wheel-running activity (30) and MR (46) even further. Whereas nonreproducing S females are diet-induced obesity-resistant (i.e., as opposed to C females), there was even a strong trend toward an increase in body fat content in lactating S dams compared with lactating C dams feeding HFS diet. Although the S dams were still more physically active (assessed by PIR activity) than the C dams, the data suggest that the hypermetabolic state of the S dams is offset by lactation, and apparently does not prevent HFS feeding-induced weight gain in the offspring. While we are currently investigating the underlying neurobiological causes and consequences of these prioritizing mechanisms, such an effect may be relevant for obesogenic programming in human offspring too (29), irrespective of the physical activity phenotype of the mother.

ACKNOWLEDGMENTS

Jan Bruggink, Gerard Overkamp, Berthe Versstappen-Dumoulin are thanked for technical assistance. Tobias-Navarro Schroeder and Jantien Zandvoort are thanked for their help in performing the experiment. TG is supported by NSF grant IOS-1121273, and GvD is supported by the Dutch Diabetes Research Foundation.

DISCLOSURES

No conflicts of interest, financial or otherwise, are declared by the authors.

AUTHOR CONTRIBUTIONS


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