Intensities fluctuations between an ON-state and an OFF-state, also called blinking, are common to all luminous objects when studied at the level of individuals. We studied blinking of three dyes from a homologous series (Cy3, Cy5, Cy7). The underlying radial anion states were induced by removing oxidants (i.e., oxygen) and by adding the reductant ascorbic acid. We find that for different conditions with distinct levels of oxidants in solution the OFF-state lifetime always increases in the order \( \text{Cy3} < \text{Cy5} < \text{Cy7}. \) Longer OFF-times are related to higher reduction potentials of the fluorophores, which increase with the size of the chromophore. Interestingly, we find reaction rates of the radical anion that are unexpectedly low at the assumed oxygen concentration. On the other hand, reaction rates meet the expectations of similar Rehm–Weller plots when methylviologen is used as oxidant, confirming the model of photoinduced reduction and oxidation reactions. The relation of OFF-state lifetimes to redox potentials might enable predictions about the nature of dark states, depending on the fluorophores’ nano-environment in super-resolution microscopy.

1. Introduction

On the single-molecule level, essentially all fluorophores exhibit intensity fluctuations between an ON-state and an OFF-state that is referred to as blinking. Blinking is often caused by transitions into the triplet state or radical states.\(^1,2\) Triplet states result from intersystem crossing and have a typical lifetime in the microsecond to lower millisecond range. Recently, it has been shown that radical ions formed as a result of one-electron transfer reactions can have a much longer lifetime of up to minutes (redox blinking).\(^3,4\)

For a long time unpredictable blinking had been plaguing single-molecule experiments. Only in recent years, has blinking become more controllable and even valuable for optical super-resolution microscopy.\(^5–12\) For microscopy techniques based on subsequent localization of single molecules, such as PALM, iPALM, STORM, GSDIM, dSTORM or Blink Microscopy, the majority of molecules has to be shelved to a transient dark state and the stochastically active molecules successively localized.\(^6–11\) By repeating this procedure, all molecule positions are determined and super-resolved fluorescence images are reconstructed. For this method to work efficiently, the lifetimes of the dark states of the molecules have to be long enough so that a sufficient number of molecules can be co-localized within one diffraction-limited spot.\(^13\) Presumed radical ion states have proven to be well-suited for super-resolution microscopy because they are generic dark states of all fluorophores.\(^10,13\) On the other hand, single-molecule studies showed that the lifetimes of these dark states vary significantly between different dyes at otherwise identical conditions, indicating that some dyes are better suited for super-resolution microscopy than others.\(^10,11,14\)

It has been argued that a higher reduction potential of the fluorophores, that is, a better-stabilized radical ion, should be accompanied by a longer OFF-state lifetime.\(^16\) Accordingly, the reduction potential might be the parameter to evaluate dyes for localization-based super-resolution microscopy. This is why certain oxazine dyes with high reduction potentials have been chosen for super-resolution microscopy.\(^4\) On the other hand, very recently, it has been shown that these oxazine dyes can undergo a second reduction, leading to a long-lived leuko form.\(^15\) Then there is the general question about the nature of the underlying dark states in localization based super-resolution microscopy using the blinking of dyes.\(^16,11,16\) To advance blinking-based super-resolution imaging, the understanding of the underlying dark states is crucial. In addition, the ability to predict the properties of dyes at the single-molecule level will foster further developments. A systematic study of chromo-
phore properties and blinking is required to test the hypothesis of the correlation of blinking kinetics and reduction potentials.

Herein, we study the dependence of redox blinking on the electronic properties of dyes with a focus on the absorption wavelength and the reduction potential. To minimize other influences such as chromophore structure, for example due to chemical substituents, differing hydrophobicity or steric constraints, we used dyes from the homologous series of cyanine fluorophores. Cy3, Cy5, and Cy7 exhibit a systematic increase of their chromophore system and the energetic properties can be described well by the simple particle-in-the-box model (i.e. absorption wavelength and chromophore size are quantitative-ly connected by the number of delocalized \( \pi \) electrons, see Figure 1, \( n = 1,2,3 \)). According to the Kuhn model\(^{17}\), the redu-

![Figure 1](https://example.com/figure1.png)

**Figure 1.** A) Overview of the chemical structures of the cyanine dyes used \( [n = 1 (\text{Cy3}), n = 2 (\text{Cy5}), n = 3 (\text{Cy7})] \). Residues are \( R_1 = R_2 = \text{CH}_3 \) and \( R_1 = (\text{CH}_3)_2\text{CO}_2\text{NH}(\text{CH}_2)_3\text{PO}_4^-\cdot\text{DNA} \) for Cy3/Cy5; \( R_1 = (\text{CH}_3)_2\text{SO}_4^- \), and \( R_2 = (\text{CH}_3)_2\text{CO}_2\text{DNA} \) for Cy7. The basic chromophore is highlighted in grey. B) Jablonski diagram of the fluorophores from one homologous series. One single molecule is excited from its singlet ground state \( S_0 \) into the first excited state \( S_1 \); accordingly, the intersystem crossing occurs with the rate constant \( k_{\text{int}} \). Intersystem crossing occurs with the rate constant \( k_{\text{int}} \). The triplet is built up within the homologous series. The absorption properties of cyanines can be described well by Kuhn’s free-electron gas model\(^ {17} \). The model also allows estimating redox potentials assuming that certain energy contributions such as the solvation energy do not change within the homologous series. In this case the LUMO energy is related to the redox potential by Equation (1):

\[
-\Delta E_{\text{red}} = \frac{A}{F} + k \approx \frac{E_{\text{LUMO}}}{F} + k
\]

where \( A \) is the electron affinity, \( F \) is Faraday’s constant and \( k \) is a constant that includes the free energy change for solvation\(^ {21} \). Since the LUMO energy decreases with increasing size of the box, the reduction potential increases. That is, larger fluorophores within a homologous series are reduced more easily. Since the reaction rates of non-diffusion-limited processes are linked to the Gibb’s free energy of the reaction, we expect that the re-oxidation of the radical anion will be faster in the order Cy7 \( \geq \) Cy5 \( \geq \) Cy3 under otherwise identical experimental conditions\(^ {22} \). This means that the OFF-state lifetime of the radical anion should increase in the order Cy3 \( \leq \) Cy5 \( \leq \) Cy7 in a reducing buffer and low concentrations of oxidant. We used molecular oxygen and 1,1’-dimethyl-4,4’-bipyridinium dichloride hydrate (methylviologen, MV) as oxidants.

To study the OFF-state lifetime of the homologous series of cyanines, fluorophores bound to biotinylated DNA were immobilized with BSA-biotin/neutravidin on glass cover slips. For excitation of the fluorophores from different spectral regions, we used a multi-colour single-molecule setup equipped with a supercontinuum laser and acousto-optical components for free wavelength selection. The detection is carried out by avalanche photodiodes and the spectral windows are selected using dichroic mirrors and appropriate filters. For details of the setup see the Experimental Section and refs.\(^ {23,24} \). After recording adding 75 \( \mu \text{M} \) of the reductant ascorbic acid (AA)\(^ {10} \). Single molecules were placed in the laser focus of a confocal microscope and fluorescence transients of redox blinking molecules were recorded. From these transients we extracted the OFF-state lifetime for the three cyanine derivatives. A strong correlation of chromophore size and OFF-state lifetime corroborates the idea that the simple model is able to predict qualitatively the OFF-state lifetime of cyanines, substantiating that the redox potentials are a crucial parameter for dye selection in super-resolution microscopy. In the future, deviations from such predictions might point towards a different nature of observed OFF-states.

### 2. Results and Discussion

We used three dyes from the homologous series of cyanine fluorophores, Cy3 (\( \lambda_{\text{max}} = 550 \text{ nm} \)), Cy5 (\( \lambda_{\text{max}} = 650 \text{ nm} \)), and Cy7 (\( \lambda_{\text{max}} = 750 \text{ nm} \)). These fluorophores are commonly used to label proteins and nucleic acids and also in demanding applications such as single-molecule FRET and super-resolution microscopy\(^ {16-20} \).

As a basis for our experiment we make a simple qualitative prediction on the blinking properties of fluorophores from one homologous series. The absorption properties of cyanines can be described well by Kuhn’s free-electron gas model\(^ {17} \). The model also allows estimating redox potentials assuming that certain energy contributions such as the solvation energy do not change within the homologous series. In this case the LUMO energy is related to the redox potential by Equation (1):

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fluorescence images (Figure 2A), fluorescence intensity transients were recorded and analysed using custom-made LabVIEW code and OFF-times were determined by autocorrelation analysis as described in ref. [4]. For the analysis only transients that showed a distinct single bleaching step at the end were selected, which allows for individual background correction. A section of an exemplary transient is depicted in Figure 2B together with the corresponding autocorrelation function (Figure 2C).

Besides the component for redox blinking \( t_{\text{OFF}} \) = 41 ms and \( t_{\text{ON}} \) = 3.8 ms, a short bunching term \( t_{\text{OFF}} \) = 180 \( \mu \)s and \( t_{\text{ON}} \) = 20 \( \mu \)s is visible that is assigned to \( \text{cis} \rightarrow \text{trans} \) isomerisation common to cyanine dyes.[25] At the conditions used, the average ON- and OFF-times of \( \text{cis} \rightarrow \text{trans} \) isomerisation are \( t_{\text{OFF}} \) (Cy3) = 37 \( \mu \)s and \( t_{\text{ON}} \) (Cy3) = 47 \( \mu \)s, \( t_{\text{OFF}} \) (Cy5) = 165 \( \mu \)s and \( t_{\text{ON}} \) (Cy5) = 22 \( \mu \)s, and \( t_{\text{OFF}} \) (Cy7) = 227 \( \mu \)s and \( t_{\text{ON}} \) (Cy7) = 34 \( \mu \)s for the different dyes, respectively.

### 3. Influence of Oxygen on Single-Molecule Blinking

The concentration of the reductant ascorbic acid is chosen to maximize the OFF-state lifetime and is much higher than the oxidant concentration.[10] In the presence of oxygen at atmospheric pressure corresponding to ~500 \( \mu \)M dissolved oxygen, no blinking of Cy5 is observed because triplet as well as radical anionic states are rapidly depleted by reactions with oxygen. Additionally, the molecules bleach quickly due to the formation of singlet oxygen. To also exclude initial photoinduced oxidation by the oxidant we used a significantly lower oxidant than reductant concentration. We can thus safely assume that all observed dark states that cannot be assigned to \( \text{cis} \rightarrow \text{trans} \) isomerisation but represent radical anionic states (with respect to an assumed neutral ground state chromophore). The assumption is justified for two reasons. First, due the concentration difference, reduction from an excited state is more likely than the oxidation. Second, the oxidized state would be too short-lived to be detected at the AA concentration used.

When using oxygen as oxidant, its concentration has to be controlled in a reproducible fashion. To this end, we developed a protocol for oxygen removal by the GOC system that we calibrated with the aid of an oxygen meter. The GOC system allows the removal of dissolved oxygen down to an equilibrium concentration of ~10 \( \mu \)M.[46] To check the oxygen concentration in the single-molecule experiments and to ensure that equilibrium is achieved, we studied the oxygen concentration as a function of time for different glucose oxidase concentrations by monitoring the blinking kinetics of the cyanines. Figures 3A,C show the progression of the OFF-times of Cy5 for two concentrations of glucose oxidase starting directly after addition of GOC and sealing of the chamber (1% v/v vs 10% v/v, see the Experimental Section for details). Each data point represents the average OFF-time obtained for the transient of a single cyanine molecule. The data shown in Figure 3A represent ~400 molecules and Figure 3C ~200 molecules. Both graphs exhibit an initial increase in the OFF-times. Saturation, however, occurs much faster for the higher glucose oxidase concentration after about 10 min whereas saturation is reached after more than 90 min for 1% v/v. In addition, at a lower concentration of the enzyme (1% v/v), a higher oxygen concentration is indicated by lower mean OFF-times for 1% v/v (12 ± 10 ms) compared to 10% v/v (41 ± 20 ms). Figures 3B,D show the corresponding OFF-time histograms derived after saturation was reached. These results show the importance of a sufficient concentration of glucose oxidase to reach the highest possible OFF-time and also the need to wait for...
the system to reach equilibrium. As a consequence we used 10% v/v glucose oxidase for reaching the steady state quickly and reproducibly in all subsequent experiments.

A more subtle detail of the plots in Figures 3 A,C is the asymmetric shape of the distribution. Initially, the OFF-times are quite homogenous and short. With decreasing oxygen concentration the distribution of OFF-times broadens with a remarkably sharp edge for shorter OFF-times. This distribution might reflect sample inhomogeneity with some fluorophores being more exposed to the solution (short OFF-times) and other being better shielded from oxygen by the BSA coating of the surface. A small fraction of individual fluorophore time transients showed switching between distinct OFF-times, which could be an indication of a discrete rearrangement of one fluorophore’s specific nano-environment. Moreover, measurements of dsDNA structures with Cy5 at the 5’-end yielded ~50% longer OFF-times for comparable experimental conditions (data not shown, see ref. [10]), indicating the environmental sensitivity of OFF-state lifetimes.


The dyes of the homologous series Cy3/Cy5/Cy7 exhibit an increase in chromophore size and number of π electrons that goes along with systematic changes of their spectroscopic and electrochemical properties. Some of these properties are listed in Table 1.

Investigating more than 100 single-molecule transients for each dye reveals a mean OFF-time of 18 ± 8 ms for Cy3 (Figure 4A), 41 ± 20 ms for Cy5 (Figure 4B) and 71 ± 23 ms for Cy7 (Figure 4C). We ascribe the broad distributions to the mentioned variation of the molecules nano-environment, causing a shoulder in the histograms displayed in Figure 4. Nevertheless, the average values of the OFF-time distributions are reproducible within several milliseconds when repeating the experiment, including new surface preparation, immobilization of dye molecules and oxygen removal. Table 1 shows the average OFF-time for multiple repeats (n ≥ 3) consisting of at least 100 evaluated transients per experiment. We also studied the excitation intensity dependence and found that OFF-times slightly decrease for higher excitation powers.[35] For the given conditions, Cy5, for example, exhibits ~25% shorter OFF-times at 5-fold higher excitation intensity. To reduce the influence of sample heterogeneity in addition to calculating the mean values of the distributions we investigated the behaviour of the mentioned sharp edge for shorter OFF-times for the different cyanine dyes. As a quantitative measure for this lower bound we calculated the lowest decile for the distributions and obtained \( \tau_{\text{off}} \) (Cy3) = 11 ms, \( \tau_{\text{off}} \) (Cy5) = 23 ms, and \( \tau_{\text{off}} \) (Cy7) = 52 ms, which additionally confirms the expected trend of OFF-times.

The data allow an estimation of the rate constants of these electron transfer reactions using \( \tau_{\text{off}} = k_{\text{on}} = k_{\text{ox}} [\text{Ox}] \), that is,
The rate constant for switching the molecule back ON by the electron transfer to the oxidant $k_{on}$. Assuming an oxygen concentration of 10 $\mu$M, we obtain $k_{on}$ in the range of $(10^{6} \text{ to } 10^{7})$ s$^{-1}$ M$^{-1}$ for the three dyes. The driving force for the reaction is estimated by the reduction potentials of the fluorophores and the reduction potential of oxygen.$^{[29]}$ The Gibbs energy is $\Delta G = -0.60$ eV for Cy3, $\Delta G = -0.44$ eV for Cy5, and $\Delta G = -0.32$ eV for Cy7, assuming a redox potential of $E_{red}$ (O$_2$) = -0.40 V vs SCE for 1 mol L$^{-1}$ oxygen concentration.$^{[29]}$ Comparing to typical Rehm–Weller plots,$^{[22, 31]}$ the expected order of magnitude for the rate constant of these Gibbs energies is between $(10^{6} \text{ to } 10^{7})$ s$^{-1}$ M$^{-1}$, that is, several orders of magnitude larger than the observed values. While our data qualitatively meet expectation, we cannot explain the large deviation with respect to the absolute expected values. Possibly, the oxygen concentration is much lower than the measured −10 $\mu$M because this concentration is close to the detection limit of the employed oxygen meter. Additionally, the large increase in OFF-times for many fluorophores from < 1 $\mu$s in the presence of oxygen to > 30 ms after enzymatic oxygen removal, that is, by a factor of $3 \times 10^{4}$ can hardly be explained by an oxygen reduction of only 50-fold from 500 $\mu$M to 10 $\mu$M (see for example refs. [4, 29]).

To test whether an overestimation of the oxygen concentration can be the reason for the deviations or whether there might be a more fundamental problem with our model, we used methylviologen (MV) as an alternative oxidant whose concentration is better controlled. We measured the OFF-times of the three dyes at various concentration of MV keeping the AA concentration constant at 75 $\mu$M. Oxygen was removed as in previous experiments. For all dyes, the OFF-time already decreases at nanomolar concentrations although MV (E$_{red}$ = 0.69 V vs SCE) has less oxidative potential than oxygen (Figure 5). At concentrations greater than 100 nM the OFF-times become so short that they cannot clearly be distinguished from blinking due to cis–trans isomerisation. To determine the rate constants for this system, we fitted the data in Figure 5 by Equation (2):

$$\frac{1}{t_{off}} = k_{on}[MV] + k_{on}[O_2] = k_{on}[MV] + \text{constant} \quad (2)$$

The offset in the equation accounts for the fact that the remaining oxygen concentration also contributes to the shortening of the OFF-state lifetime, that is, the OFF-state lifetime is not infinite in the absence of MV. The fits yield the reaction rate constants $k_{on}$ (Cy7) = $5.68 \times 10^{7}$ s$^{-1}$ M$^{-1}$, $k_{on}$ (Cy5) = $7.08 \times 10^{5}$ s$^{-1}$ M$^{-1}$, and $k_{on}$ (Cy3) = $7.20 \times 10^{8}$ s$^{-1}$ M$^{-1}$. Taking into account a Gibbs energy for the reaction with MV (E$_{red}$ = 0.69 V vs SCE) of $\Delta G = -0.31$ eV for Cy3, $\Delta G = -0.15$ eV for Cy5, and $\Delta G = 0.03$ eV for Cy7, these values match expectations from Rehm–Weller plots very well.$^{[22]}$ The stronger driving forces for Cy5 and Cy3 yield similar values for their quenching constant since the values approach the diffusional plateau. Cy7 only has a small driving force explaining the larger difference between Cy7 and Cy5 compared to Cy3 and Cy5. Considering the small driving force for Cy7 the observed value for $k_{on}$ is still relatively high. In summary, the kinetics of the oxidation of radical anions by MV corroborates the proposed model and suggests the tight connection between OFF-states of single-molecule redox blinking and chromophores’ electronic properties. In addition variations by different local environments of the fluorophores are indicated.

5. Conclusions

We have studied redox blinking of a homologous series of cyanine dyes. The underlying radical anionic states were induced...
by removing oxidants (i.e. oxygen) and by adding the reduc-
tant ascorbic acid. We found that for different conditions the
OFF-state lifetime was increasing in the order Cy3 < Cy5 < Cy7.
The behavior was related to the reduction potential of the flu-
orophores which increases with the size of the chromophore.
Thus, we could qualitatively predict blinking properties of flu-
orophores on the simple basis of the free-electron gas model
that a better stabilized radical anion yields longer OFF-state
lifetimes. Interestingly, we found reaction rates of the radical
anion that were unexpectedly low at the assumed oxygen con-
centration. On the other hand, reaction rates met the expecta-
tions of similar Rehm–Weller plots when methylviologen (MV)
was used as oxidant confirming the model of photoinduced re-
duction and oxidation reactions.[29] The deviation between the
results obtained with oxygen and MV as oxidants might be ex-
plained by overestimated oxygen concentrations. On the other
hand, assuming similar quenching constants for MV and oxygen,
oxgen exhibits an activity as if it was present at an ef-
fective concentration between 1–10 nM as estimated from the
fits in Figure 5. Our data also shed new light on the nature of
the underlying dark states in super-resolution microscopy that
relies on the successive localization of single molecules. Tetra-
methylrhodamine (TMR), for example, was used in live-cell
super-resolution imaging and exhibited pronounced blinking
with long OFF-states.[32,33] TMR has an estimated reduction po-
tential of $E_{\text{red}}$ (TMR) = −0.95 V vs SCE,[16] yielding a driving force
for the back electron transfer between that of Cy3 and Cy5.
From these values, OFF-states long enough for imaging at
50 Hz in living cells are not expected. This either indicates that
the nature of the underlying OFF-state might be different. Al-
ternatively, the oxygen activity might be reduced or oxygen is
heavily consumed due to the irradiation in the presence of
thiols.[15]

### Experimental Section

#### Sample Preparation

Samples for single-molecule experiments were prepared based on
dNA oligonucleotides. A 40-mer oligonucleotide labelled with a
single fluorophore at position 24 (TAC GAT TCG ATT CCT TAC ACT
Tris[2-carboxyethyl]phosphine hydrochloride (TCEP), 40
solution was prepared according to the following protocol: 40
mM or 1% (v/v) of a prior prepared glucose oxidase solution. This
pH 7.4, containing 10% (wt/v) glucose was mixed with either 10%
conditions. Standard phosphate-buffered saline (PBS) with a pH of
ried out at room temperature (22
counter-strand (also from IBA, Germany). All experiments were car-
performed in a chambered cover slide (LabTek, NUNC)
with a volume of ~750 µL sealed with microseal adhesive seals
(MSB 1001, Bio-Rad, Germany).

#### Calibration of Oxygen Removal

To calibrate the oxygen removal of the GOC system described
above we used a dissolved-oxygen meter (OxyScan light, equipped
with a micro oxygen sensor for low sample volumes, UMS GmbH,
Germany). Due to the size and characteristics of the sensor, higher
sample volumes of around 8 mL had to be used and a magnetic
stir bar was employed to ensure a constant flow. Sealing of the
small beaker glass was also performed with microseal adhesive
seals. For measurements with 10% GOC an equilibrium concentra-
tion of −10 µM oxygen was reached after approximately 10 min,
which is in good agreement with prior published data under simi-
lar conditions.[20]

#### Single-Molecule Spectroscopy

Double-stranded DNA (dsDNA) was immobilized on a glass sub-
strate coated with BSA/biotin-streptavidin according to published
procedures. By homogenizing the environment through immobili-
ization of fluorophores in solution the static heterogeneity of the
molecules is greatly reduced and thus allows the extraction of de-
tailed kinetic information through the analysis of single-molecule
fluorescence transients. To study fluorescence on the level of single
molecules, a custom-built confocal microscope was used as de-
scribed in refs. [1, 23] (detection of Cy3/Cy5: ref. [1], detection of
Cy7: ref. [23]). The laser beam of a pulsed supercontinuum source
(SuperK Extreme, Koheras, Denmark) was coupled into a single-
mode fiber and appropriate excitation light was chosen by an
acoustic-optical tunable filter (AOTFInc-VIS, AA optoelectronic).
The following excitation wavelengths were chosen: 533 nm (excitation
of the Cy3), 640 nm (excitation of Cy5) and 725 nm (excitation
of Cy7). The spatially filtered beam entered an inverse microscope
and was coupled into an oil immersion objective (60
m, NA 1.35,
UPLSAPO 60XO, or 100
m, NA 1.45, PLAPO100XO/TIRFM-SP, both
from Olympus) by a dual-band dichroic beam splitter (Dualband
z532/633 rfc for Cy5/Cy7 and a 740 DCXXR for Cy7, AHF Analysentechnik, Germany). Fluorescence transients and confocal images
(size of 10 µm × 10 µm, integration time 2 ms per pixel, 50 nm per
pixel) were recorded under the following conditions: 1.5 kW cm
2 at 533 nm, 3 kW cm
2 at 640 nm, and 8 kW cm
2 at 725 nm. The re-
sulting fluorescence was collected by the same objective, focused
onto a 50 µm pinhole, and split spectrally at 640 nm onto two
APDs for Cy3/Cy5 by a dichroic beam splitter (640DCXR, AHF Ana-
lysentechnik, Germany) or onto a 120 µm pinhole and directly
imaged onto only one APD for Cy7. Fluorescence was detected by
avalanche photodiodes (SPCM-AQR-14, PerkinElmer) with appropi-
ate spectral filtering (Brightline HCS82/75 for Cy3, ET-Bandpass
700/75 M and Razoredge longpass for Cy5, Brightline HC 785/62
for Cy7, AHF Analysentechnik, Germany).

#### Data Evaluation and Analysis

The detector signal was registered and evaluated using custom
made LabVIEW software. Single-molecule traces and autocorrela-
tion analysis was performed according to published procedures.[6]

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