

Arabidopsis Mutants Deleted in the Light-Harvesting Protein Lhcb4 Have a Disrupted Photosystem II Macrostructure and Are Defective in Photoprotection

Silvia de Bianchi,a,n 1 Nico Betterle,a,n 1 Roman Kouril,b Stefano Cazzaniga,a Egbert Boekema,b Roberto Bassi,a,n,2 and Luca Dall’Ostoa

a Dipartimento di Biotecnologie, Università di Verona, 37134 Verona, Italy
b Department of Biophysical Chemistry, Groningen Biomolecular Sciences and Biotechnology Institute, University of Groningen, 9747 AG Groningen, The Netherlands
c Institut für Pflanzenwissenschaften-2, Pflanzenwissenschaften, Forschungszentrum Jülich, D-52425 Juelich, Germany

The role of the light-harvesting complex Lhcb4 (CP29) in photosynthesis was investigated in Arabidopsis thaliana by characterizing knockout lines for each of the three Lhcb4 isoforms (Lhcb4.1/4.2/4.3). Plants lacking all isoforms (koLhcb4) showed a compensatory increase of Lhcb1 and a slightly reduced photosystem II/I ratio with respect to the wild type. The absence of Lhcb4 did not result in alteration in electron transport rates. However, the kinetic of state transition was faster in the mutant, and nonphotochemical quenching activity was lower in koLhcb4 plants with respect to either wild type or mutants retaining a single Lhcb4 isoform. KoLhcb4 plants were more sensitive to photoinhibition, while this effect was not observed in knockout lines for any other photosystem II antenna subunit. Ultrastructural analysis of thylakoid grana membranes showed a lower density of photosystem II complexes in koLhcb4. Moreover, analysis of isolated supercomplexes showed a different overall shape of the C2S2 particles due to a different binding mode of the S-trimer to the core complex. An empty space was observed within the photosystem II supercomplex at the Lhcb4 position, implying that the missing Lhcb4 was not replaced by other Lhc subunits. This suggests that Lhcb4 is unique among photosystem II antenna proteins and determinant for photosystem II macro-organization and photoprotection.

INTRODUCTION

Oxygenic photosynthesis is performed in the chloroplast by a series of reactions that exploits light as an energy source to fuel ATP and NADPH production for CO2 fixation and synthesis of organic compounds. Light harvesting is the primary process in photosynthesis and consists of absorption of photons by an array of hundreds of chlorophylls (Chls) organized into photosystems. Excitons are transferred among Chls and to the reaction center (RC), where charge separation occurs. In photosystem II (PSII), electrons are transferred to the quinonic acceptor plastoquinone (PQ), leading to charge accumulation in the oxygen evolving complex and water splitting (Nelson and Benson, 2004). RC of PSII consists of the D1/D2/cytochrome b559 complex carrying the cofactors for electron transport, which forms, together with the nearest subunits CP43 and CP47, a core complex (Ferreira et al., 2004). Core complexes form dimers (C2), which bind an extended system of nuclear-encoded light-harvesting proteins (Lhc), each binding Chl a, Chl b, and xanthophylls. Lhcb4 (CP29) and Lhcb5 (CP26) are located near the core and mediate the binding of a trimeric LHCCI antenna complex called LHCI-I (strongly bound). These components form the basic PSII supercomplex structure, called C2S2 (Boekema et al., 1999), which is the major PSII form found in plants grown under high light (HL) (Morosinotto et al., 2006; Frigerio et al., 2007). Under low/moderate light, larger supercomplexes, called C2S3M2, are formed: one additional monomeric subunit Lhcb6 (CP24) and two LHCCI trimers (LHCCI-M and LHCCI-L), moderately and loosely bound, respectively) are accumulated to extend the light-harvesting capacity (Melis 1991; Balottari et al., 2007). In Arabidopsis thaliana, single genes encode Lhcb5 (CP26) and Lhcb6 (CP24) subunits, while Lhcb4 (CP29) is encoded by three highly conserved genes. Lhcb4.1 and Lhcb4.2 are similarly expressed, while the level of the Lhcb4.3 messenger is 20 times lower under control conditions (Jansson, 1999). The polypeptide encoded by Lhcb4.3 is predicted to lack a large part of the C-terminal domain, a peculiar feature of Lhcb4.1 and Lhcb4.2 isoforms, leading to the suggestion of renaming it as Lhcb8 (Klimmek et al., 2006).

A remarkable property of the antenna system is the ability to actively regulate PSII quantum efficiency to avoid the damaging effects of excess light. Indeed, under constant moderate light conditions, the efficiency of energy conversion is high, due to photochemical reactions. On the other hand, fluctuations of light intensity/temperature/water availability may yield into the

1 These authors contributed equally to this work.
2 Address correspondence to roberto.bassi@univr.it.
The author responsible for distribution of materials integral to the findings presented in this article in accordance with the policy described in the Instructions for Authors (www.plantcell.org) is: Roberto Bassi (roberto.bassi@univr.it).
Some figures in this article are displayed in color online but in black and white in the print edition.
Online version contains Web-only data.
www.plantcell.org/cgi/doi/10.1105/tpc.111.087320
excitation of PSII over the capacity for photochemical quenching of Chl singlet excited states (\(^{3}\text{Chl}^*\)). The consequent lifetime increase enhances the probability of Chl triplet (\(^{3}\text{Chl}^*) formation by intersystem crossing and yields into single oxygen (\(^{1}\text{O}_2\)) production (Melis, 1999). Since formation of \(^{3}\text{Chl}^*\) is a constitutive property of Chls (Mozzo et al., 2008), photoprotection mechanisms are activated to prevent damage and improve fitness in the ever-changing environment experienced by plants. Safety systems have evolved to either detoxify the reactive oxygen species (ROS) (Asada, 1999) or to prevent their formation (Niyogi 2000) by (1) downregulating \(^{1}\text{Chl}^*\) lifetime through the process of nonphotochemical quenching (NPQ) that dissipates excess excited states into heat (Horton 1996); (2) by quenching \(^{3}\text{Chl}^*\); and (3) by scavenging ROS. Sustained overexcitation is counteracted by the long-term reduction of PSII antenna size (Anderson, 1986). Additional regulation is activated in limiting light conditions by transferring LHCII complexes between photosystems, which balances excitation delivery to photosystem I (PSI) and PSII (Allen, 1992; Haldrup et al., 2001).

The conservation of the different Lhcb gene products through evolution suggests each has a specific functional role, which, however, is not yet fully clarified. Two major approaches have been used to this aim: namely, the analysis of individual gene products either purified from thylakoids (Bass et al., 1987; Caffarri et al., 2001) or recombinant (Caffarri et al., 2004; Formaggio et al., 2001) and reverse genetics, by which plants lacking one or more Lhcb proteins have been produced and characterized (Andersson et al., 2001; Kovács et al., 2006; de Bianchi et al., 2008; Damkjaer et al., 2009). These studies have provided evidence for differential roles of LHCII components (Lhcb1-3) with respect to monomeric subunits (Lhcb4-6). Antisense lines of Arabidopsis devoid of Lhcb1+2 (Ruban et al., 2003) showed a reduced fitness for plants grown in the field, while only minor differences were observed for growth rate, PSIII quantum yield, photosynthetic rate, and capacity for NPQ in a controlled environment (Andersson et al., 2003). Similar considerations apply to Lhc5, the major effect of its depletion consisting of a faster kinetic of state I to state II transitions (Damkjaer et al., 2009). More specific effects have been reported for monomeric Lhcb4 and Lhc6, which have been shown to (1) carry protonatable (DCCD binding) sites (Pesaresi et al., 1997; Walters et al., 1996); and (2) undergo conformation changes upon exchange of violaxanthin (Viola) to zeaxanthin (Zea) in their L2 xanthophyll binding site (Morosinotto et al., 2002), an important feature since Zea is synthesized in excess light conditions and correlates with NPQ (Demmig-Adams et al., 1989). The Zea-dependent allosteric change was shown to affect the properties of these pigment proteins, including a decrease in fluorescence lifetime (Crivi et al., 2001; Moya et al., 2001) and formation of carotenoid radical cations (Ahn et al., 2008; Avenson et al., 2008), suggesting a major role in NPQ. Consistently, koLhcb6 plants exhibited decreased qE (Kovács et al., 2006; de Bianchi et al., 2008), while koLhcb5 plants were affected in the slower relaxing component of NPQ called qI. Other properties of Lhcb proteins, which are essential for photoprotection, appear to be redundant. In fact, while the total depletion of Lhcb proteins induces photosensitivity in chl mutants (Kim et al., 2009; Dall’Osto et al., 2010), no individual knockout (KO) lines have shown major impairment in their capacity to resist HL treatment so far (Ruban et al., 2003; de Bianchi et al., 2008). Lhcb4 antisense lines showed NPO kinetics and amplitude, photosynthetic electron transport rate, and light sensitivity similar to that of the wild type (Andersson et al., 2001). However, since this work only employed growth in mild conditions, the question is open for the role of Lhcb4 in photoprotection. Indeed, the maintenance of Lhcb4, in HL stress conditions that leads to the reduction of Lhcb6 and LHCII (Ballottari et al., 2007), suggests that Lhcb4 might be a key factor in light harvesting and photoprotection. Lhcb4 phosphorylation in both monocots (Testi et al., 1996) and dicots (Hansson and Vener, 2003; Tikkanen et al., 2006) affects the spectral properties of the protein (Croce et al., 1996) and correlates with higher resistance to HL and cold (Bergantino et al., 1995; Mauro et al., 1997) and water stress (Liu et al., 2009). Although the mechanism of this protective effect is unknown, there is evidence for a link between Lhcb4 and NPQ, since the protein has been identified as an interaction partner of PsbS (Teardo et al., 2007), the pH-dependent trigger of qE (Li et al., 2000), and it is part of a pentameric complex whose dissociation is indispensable for the establishment of NPQ (Betterle et al., 2009). Finally, two Chl ligands in Lhcb4 have been identified as components of the quenching site, which catalyzes the formation of a xanthophyll radical cation (Holt et al., 2005; Avenson et al., 2008).

In this work, we constructed KO mutants for Lhcb4 isoforms in Arabidopsis and analyzed their performance in photosynthesis and photoprotection. PSIII quantum efficiency and capacity for NPQ were affected by lack of Lhcb4, and, unlike that of any other Lhcb subunit, the capacity of resisting excess light conditions was affected. Although depletion of Lhcb subunits is usually complemented by the overaccumulation of other members of the subfamily (Ruban et al., 2003; de Bianchi et al., 2008), this is not the case for Lhcb4 and PSIII supercomplexes isolated from koLhcb4 that although they retained their C2S2 organization, they lacked Lhcb4. This caused a different mode of binding of LHClI-S trimer within the PSI supercomplex and changed the overall shape of the C2S2 particle. Deletion of either Lhcb4.1 or Lhcb4.2 yielded into a compensatory accumulation of the remaining subunit; instead, the double mutant koLhcb4.1.4.2 was unable to accumulate Lhcb4.3. We conclude that Lhcb4 is a fundamental component of PSII, which is essential for maintenance of both the function and structural organization of this photosystem.

RESULTS

The construction of a plant without a CP29 complex (hereafter named koLhcb4) requires the isolation of KO mutants at three distinct loci, namely, Lhcb4.1, Lhcb4.2, and Lhcb4.3 (Jansson, 1999). We identified koLhcb4.1, koLhcb4.2, and koLhcb4.3 homozygous plants in T-DNA FS seed pools, obtained from the European Arabidopsis Stock Centre (NASC), by PCR analysis of genomic DNA using specific primers (Figure 1A). The triple KO mutant koLhcb4 was obtained by selection of the progeny from crossing single mutants. PCR analysis confirmed that all Lhcb4 coding regions carried a T-DNA insertion in both alleles (Figure 1A), while RT-PCR showed that mRNAs encoding Lhcb4 isoforms were absent in the mutant (Figure 1B).
During the course of this work, two additional lines (N124926 and SK32480) with a T-DNA insertion mapped to the \textit{Lhcb4.1} and \textit{Lhcb4.2} genes, respectively, became available. Both lacked the corresponding protein. The lines showed very similar phenotypic characteristics to N376476 (\textit{koLhcb4.1}) and N877954 (\textit{koLhcb4.2}), respectively. Below, we present data from the N376476 and N877954 lines, unless otherwise stated.

By screening the F2 generation, it was possible to isolate double mutants expressing single Lhcb4 isoforms, namely, \textit{koLhcb4.2 4.3} (retaining Lhcb4.1), \textit{koLhcb4.1 4.3} (retaining Lhcb4.2), and \textit{koLhcb4.1 4.2} (retaining Lhcb4.3). All of the isolated genotypes did not show a significant reduction in growth with respect to the wild type under control light conditions (100 \(\mu\text{mol photons m}^{-2}\text{s}^{-1}\), 24°C, 8/16 day/night). Electron microscopy analysis of plastids from mesophyll cells of the wild type and mutants was performed to test if the thylakoid structure was changed as an effect of the missing Lhcb (see Supplemental Figure 1 online). All the mutants showed a membrane organization similar to that of wild-type chloroplasts with well-defined grana, containing approximately six discs per granum, as well as...
similar amounts of grana, stroma lamellae, and end membranes per plastid (see Supplemental Table 1 online).

Thylakoid membranes isolated from the koLhcb4 mutant lacked the corresponding gene product (Figure 1C). A better resolution in the 22 to 35 kD molecular mass range was obtained using a gel incorporating 7 M urea, which split Lhcb4 into a doublet in wild-type thylakoids, the upper band corresponding to Lhcb4.2 as shown by comparison with the pattern from double KO genotypes (Figure 1D). The gel without urea (Figure 1C) revealed that an additional band with lower molecular mass was also missing, which corresponded to Lhcb6, as revealed by immunoblotting (Figures 2A and 2B). The pigment content of the koLhcb4 mutant did not differ from the wild type in either chlorophyll content per leaf area or Chl/Car ratio but showed a significant decrease in the Chl a/Chl b ratio (Table 1).

To determine whether the capacity of the antenna system and its ability to transfer absorbed energy to RCs was affected by the mutation, the functional antenna size of PSII was measured on thylakoids by estimating the rise time of chlorophyll fluorescence in the presence of DCMU and nigericin. No significant differences were observed in $t_{2/3}$ $F_{\text{max}}$ (see Methods for details) between koLhcb4 and the wild type (Table 1; see Supplemental Figure 2 online), suggesting that Lhcb4 depletion did not impair the overall light-harvesting capacity. Analysis of the fluorescence induction in dark-adapted leaves (Butler and Strasser, 1978) revealed a higher F0 value and a significant decrease of maximum quantum efficiency of PSII ($F_v/F_m$) in koLhcb4 with respect to the wild type. Thus, a larger fraction of absorbed energy is lost as fluorescence in the mutant, implying that the connection between the major LHCII complex and PSII RC is less efficient in the absence of Lhcb4 (Table 1). Nevertheless, parameter $S_{\text{ET}}$ which is used for quantifying PSII electron transport (ET) activity, was essentially the same in koLhcb4 and the wild type, suggesting that ET was not limited downstream from QA in mutant leaves.

**Figure 2.** Polypeptide Composition of Thylakoid Membranes from Wild-Type and koLhcb4 Mutant.

(A) Immunoblotting used for the quantification of photosynthetic subunits in the wild type (WT) and koLhcb4 thylakoids. Immunoblot analysis was performed with antibodies directed against individual gene products: minor antenna proteins, the LHCII subunit, the PSII core subunit PsbB (CP47), the PSI core subunit PsbA, cytochrome f, and plastocyanin. Thylakoids corresponding to 0.25, 0.5, 0.75, and 1 mg of chlorophyll were loaded for each sample. All samples were loaded on the same SDS-PAGE slab gel.

(B) Results of the immunotitration of thylakoid proteins. Data of PSII antenna subunits were normalized to the core amount, PsbB content (Ballottari et al., 2007), and expressed as a percentage of the corresponding wild-type content. Significantly different values from wild-type membranes are marked with an asterisk.

(C) Thylakoid pigment-protein complexes were separated by nondenaturing Deriphat-PAGE upon solubilization with α-DM. Thylakoids corresponding to 25 μg of chlorophylls were loaded in each lane.

[See online article for color version of this figure.]
Pigment-Protein Complexes
Organization and Stoichiometry of Pigment-Protein Complexes

The organization of pigment binding complexes was analyzed by non-denaturing Deriphat-PAGE upon solubilization of wild-type and koLhcb4 thylakoid membranes with 0.7% dodecyl-α-D-maltoside (α-DM). Seven major green bands were resolved (Figure 2C). In the wild type, the PSI-LHCI complex was found as a major band (B7) in the upper part of the gel, while the components of the PSII-LHCII migrated as multiple green bands with different apparent masses, namely, the antenna core dimers and monomer (B6 and B5, respectively) and the antenna moieties, including the Lhcb4-Lhcb6-LHCII-M supercomplex (B4), LHCII trimer (B3), and monomeric Lhcb6 (B2). Four faint green bands with high apparent molecular mass, which contained undissociated PSII supercomplexes of different LHCII composition, were detected in the upper part of the gel (B8). The major differences detected in koLhcb4 with respect to the wild type were the lack of B4 and a reduced level of PSI supercomplexes. Densitometric analysis of the Deriphat-PAGE showed a higher PSI/PSII ratio in koLhcb4 mutant (1.38 ± 0.11) with respect to the wild type (1.04 ± 0.03).

To detect possible alterations in the relative amount of protein components of the photosynthetic apparatus, we determined the stoichiometry of several subunits by immunoblotting titration using CP43 (PsbB) as an internal standard (Ballottari et al., 2007). KoLhcb4 plants lacked both Lhcb4 and Lhcb6 proteins (Figure 2A), while there was a 30% increase in Lhcb1 (Figure 2B) with respect to the wild-type level. The other pigment-protein complexes, as well as cytochrome (cyt) f and plastocyanin, were present in wild-type amounts. A slight but significant increase in PsaA content in koLhcb4 compared with the wild type (Figure 2A) was detected.

We also analyzed the Lhc protein composition in double mutants expressing individual Lhcb4 isoforms (Figure 1D; see Supplemental Table 2 and Supplemental Figure 3 online). The koLhcb4.2 4.3 mutant retained 87% of wild-type Lhcb4 levels, while koLhcb4.1 4.3 had a significantly lower amount of Lhcb4 (59%) with respect to the wild type (see Supplemental Table 2 online). The koLhcb4.1 4.2 mutant did not show any band with mobility similar to Lhcb4.1 and Lhcb4.2 (Figure 1D). Nevertheless, the Lhcb4.3 gene product is expected to have a molecular mass significantly lower than the 0.1 and 0.2 isoforms. Therefore, we cannot exclude that it migrates together with the bulky LHCII band. Immunoblotting analysis also revealed that the level of Lhcb6 polypeptide underwent changes in agreement with the amounts of Lhcb4.1 + Lhcb4.2 protein. In the koLhcb4.1 4.2 mutant, Lhcb6 was not detectable (see Supplemental Table 2 online).

Photosynthesis-Related Functions: ET Rate, State Transition, and P700 Redox State

Since pigment-protein complexes participate in modulating ET between photosystems, PSI and PSII function during photosynthesis was further analyzed by chlorophyll floometry. KoLhcb4 showed no significant differences with respect to wild-type plants either in the linear electron flow (LEF) or in the Qo redox state (qL), as measured at different light intensities in the presence of saturating CO2 on leaves (Figures 3A and 3B), thus indicating no limitations in photosynthesis downstream of PSII. This was further confirmed by measuring rates of O2 evolution in CO2-saturating conditions: no significant reduction in the quantum yield of photosynthesis was observed in koLhcb4 versus wild-type plants at any of the light intensities used (Figure 3C).

The capacity for state transitions (Allen, 1992) was measured from the changes in chlorophyll fluorescence on leaves (Jensen et al., 2000). The final amplitude of the state transitions (qT) after a 15-min illumination was the same in both genotypes (0.091 ± 0.007), indicating no limitations in photosynthesis downstream of PSI. Furthermore, it was observed that the switching on of far-red light produced a sudden decrease of fluorescence intensity in the wild type but not in koLhcb4. In fact, fluorescence level in the latter was already very low, consistent with a faster reoxidation of the PQ pool in koLhcb4 with respect to the wild type under low-intensity blue light.

Fluorescence induction analysis on intact leaves showed a F0 value up to 40% higher in koLhcb4 versus the wild type, implying the absence of Lhcb4 caused a lower efficiency in energy transfer between LHCII and PSII RC (Table 1). Photosynthetic electron flow through PSI during steady state photosynthesis in vivo was measured from the dependence of the P700/total P700 ratio on intensity of PSI+PSII light (λ = 630 nm; Figure 3F). In the wild type, this value approached saturation level at ~1000 μmol photons m⁻² s⁻¹. In koLhcb4, the saturation was observed already at 600 μmol photons m⁻² s⁻¹, which implies a higher

### Table 1. Chlorophyll Content and Fluorescence Induction Parameters Determined for Leaves of Arabidopsis Wild-Type and koLhcb4 Plants and Mutants Retaining a Single Lhcb4 Isoform

<table>
<thead>
<tr>
<th>Genotype</th>
<th>Chl a/b</th>
<th>Chl/Car</th>
<th>μg Chl/cm²</th>
<th>F0</th>
<th>Fv/Fm</th>
<th>t½s (ms)</th>
<th>S½n/Fmax (ms⁻¹)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Wild type</td>
<td>3.06 ± 0.07</td>
<td>3.63 ± 0.08</td>
<td>20.7 ± 3.0</td>
<td>0.195 ± 0.002</td>
<td>0.790 ± 0.007</td>
<td>182 ± 6</td>
<td>1.2 ± 0.1</td>
</tr>
<tr>
<td>koLhcb4.2 4.3</td>
<td>2.83 ± 0.06*</td>
<td>3.58 ± 0.03</td>
<td>19.0 ± 1.1</td>
<td>0.253 ± 0.021*</td>
<td>0.747 ± 0.021*</td>
<td>176 ± 8</td>
<td>1.5 ± 0.3</td>
</tr>
<tr>
<td>koLhcb4.1 4.3</td>
<td>2.97 ± 0.03</td>
<td>3.70 ± 0.13</td>
<td>22.5 ± 1.2</td>
<td>0.213 ± 0.012</td>
<td>0.796 ± 0.004</td>
<td>182 ± 5</td>
<td>1.1 ± 0.4</td>
</tr>
<tr>
<td>koLhcb4.1 4.2</td>
<td>2.83 ± 0.09*</td>
<td>3.63 ± 0.08</td>
<td>22.9 ± 2.5</td>
<td>0.266 ± 0.004*</td>
<td>0.745 ± 0.007*</td>
<td>184 ± 6</td>
<td>1.4 ± 0.2</td>
</tr>
</tbody>
</table>

Data are expressed as mean ± SD (n > 5); significantly different values (Student’s t test, P = 0.05) with respect to the wild type are marked with an asterisk.

Functional Role of Lhcb4 2663
Figure 3. Characterization of Photosynthetic Electron Flow in Wild-Type and koLhcb4 Plants.

(A) Dependence of the LEF rate on light intensity in wild-type (WT) and koLhcb4 leaves. LEF rate is calculated as $\phi_{\text{PSII}} \cdot \text{PAR} \cdot A_{\text{leaf}} \cdot \text{fraction}_{\text{PSII}}$ (see Methods for details).

(B) Photosynthetic oxygen evolution in saturating CO$_2$. Rate of oxygen evolution was measured on whole leaves during illumination with various levels of actinic red light.

(C) Amplitude of qL measured at different light intensities on wild-type and koLhcb4 leaves. qL reflects the redox state of the primary electron acceptor $Q_{A}$, thus, the fraction of open PSII centers.

(D) and (E) Measurement of state I–state II transition in the wild type (D) and koLhcb4 (E). Upon 1-h dark adaptation, plants were illuminated with blue light (40 $\mu$mol photons m$^{-2}$ s$^{-1}$, wavelength <500 nm) for 15 min to reach state II. A far-red light source was then superimposed on the blue light to induce a transition to state I. Values of $F_{m}$, $F_{m}^{-}$, and $F_{m}^{+}$ were determined using light saturation pulses (4500 $\mu$mol photons m$^{-2}$ s$^{-1}$, 0.6 s). a.u., arbitrary units.

(F) and (G) Dependence of the P700 oxidation ratio ($\Delta A/\Delta A_{\text{max}}$) on light intensity. $\Delta A/\Delta A_{\text{max}}$ was measured on wild-type and koLhcb4 leaves at varying actinic intensities, exciting either PSII + PSI ($\lambda = 630$ nm; [F]) or PSI only ($\lambda = 730$ nm; [G]).
NPQ of Chlorophyll Fluorescence

Thermal energy dissipation is a major photoprotection mechanism in plants. Although a quenching component has been localized in PSII core (Finazzi et al., 2004), most quenching activity was associated to the antenna system (Horton and Ruban, 2005; Havaux et al., 2007). In a previous work (Betterle et al., 2009), we observed that the koLhcb4 mutant, among other KO mutants for Lhcb genes, cannot assemble the CP29-CP24-LHClII complex (B4C), whose dissociation is essential for triggering of NPQ. Here, we focus on the elucidation of the mechanistic role of Lhcb4 in NPQ and photoprotection. The NPQ activity of koLhcb4 plants is shown in Figure 4A. Upon exposure of the wild type to saturating light intensity (1200 μmol photons m⁻² s⁻¹, 24°C), NPQ showed a rapid rise to 1.4 in the first minute followed by a slower raise, reaching a value of 2.3 after 8 min. Induction of NPQ in koLhcb4 was slower (0.7 at t = 1 min) and reached a lower amplitude (1.6 at t = 8 min). Recovery in the dark was faster and wider in koLhcb4 compared with the wild type (Figure 4A), as previously shown (Betterle et al., 2009). NPQ activity of koLhcb4 reached the wild-type level upon longer light treatment (25 min). When a second illumination period was applied (Figure 4B), the delay in NPQ rise was reduced, while dark recovery was still faster in koLhcb4. It should be noted that Zea synthesis had the same kinetic in the wild type and koLhcb4 (Figure 4C), implying that the delayed NPQ onset was not due to a delayed Zea synthesis.

We also investigated whether the three Lhcb4 isoforms had the same activity in NPQ by analyzing the kinetic of NPQ rise in double mutants retaining one single Lhcb4 gene. The results reported in Figure 4D show that mutants conserving the Lhcb4.1 gene had slightly faster onset of NPQ than the wild type, while those retaining Lhcb4.2 gene were unable to reach the final NPQ level of wild-type plants. Mutant plants retaining only Lhcb4.3 behaved like koLhcb4. The faster fluorescence recovery observed in koLhcb4 compared with the wild type is present in the mutants retaining only Lhcb4.2 or Lhcb4.3 genes, whereas mutant retaining Lhcb4.1 recovered with the slower wild-type kinetic.

Photosensitivity under Short- and Long-Term Stress Conditions

Treatment of plants with strong light produces photooxidative stress, whose severity is enhanced by low temperature. Under these conditions, enhanced release of O₂ radicals bleaching of pigments, lipid oxidation, and PSII photoinhibition, which is accompanied by a decrease in Fv/Fm (Zhang and Scheller, 2004). The sensitivity to photooxidative stress of wild-type and Lhcb4-depleted plants was assessed upon transfer from control conditions to HL + low temperature (500 μmol photons m⁻² s⁻¹, 4°C). The level of Fv/Fm was monitored for 2 d (Figure 5A). In wild-type plants, the Fv/Fm parameter gradually decreased from 0.8 to 0.35 during the treatment, while in koLhcb4 plants, the decrease was stronger, reaching a value of 0.15 at the end of the treatment. As a reference, we compared the photosensitivity of different antenna mutants, namely, koLhcb5/Lhc6, koLhcb3, and LHClII antisense plants (asLHClII). Interestingly, mutants lacking antenna components other than Lhcb4 showed the same level of photoinhibition as wild-type plants, implying that Lhcb4 is the only antenna component indispensable for full level of photoprotection under photooxidative stress (Figure 5A).

Measurements of Fv/Fm recovery after photoinhibitory treatment (see Supplemental Figure 6 online) clearly showed that wild-type and koLhcb4 leaves had the same capacity of PSII quantum efficiency recovery, implying that a higher photosensitivity of mutant plants is due to a less effective photoprotection rather than to impaired PSII repair mechanism (Aro et al., 1994). Loss of Lhcb4 caused a decrease in PSII quantum yield (Figure 5A) that could be caused either by damage of the PSII core complex or by an incomplete excitation transfer to PSII.
To provide a more complete characterization of the photodamage, leaf discs from the wild type and koLhcb4 were submitted to HL + cold stress (1500 \( \mu \)mol photons m\(^{-2}\) s\(^{-1}\), 4\(^{\circ}\)C) and the time course of pigment photobleaching and lipid peroxidation was measured (Figures 5B and 5C). Analysis indicates that chlorophyll bleaching was faster and malondialdehyde (MDA) production was higher in koLhcb4 with respect to wild-type leaves, implying a higher level of lipid peroxidation (Havaux et al., 2005). Photodamage can be caused by the production of ROS, including singlet oxygen (\(^1\)O\(_2\)) (Triantaphylides et al., 2008). To identify if this species was involved in the preferential photodamage of koLhcb4, we quantified production of \(^1\)O\(_2\) directly in wild-type and koLhcb4 leaves using vacuum-infiltrated \(^1\)O\(_2\)-specific probe (see Methods for details). After illumination with strong light at 4\(^{\circ}\)C, koLhcb4 leaves clearly showed a significantly higher release of singlet oxygen compared with the wild type (Figure 5D).

An interesting question is whether the different Lhcb4 isoforms have a specific importance in photoprotection. We measured the level of PSII photoinhibition on KO mutants retaining individual Lhcb4 genes. Plants retaining either Lhcb4.1 or Lhcb4.2 showed a wild-type level of resistance to HL + cold stress, while those

![Graph A](image1)

![Graph B](image2)

![Graph C](image3)

![Graph D](image4)

**Figure 4.** Kinetics of the Formation and Relaxation of Photoprotective Energy Dissipation.

(A) Measurements of NPQ kinetics on wild-type (WT) and koLhcb4 leaves illuminated with 1200 \( \mu \)mol photons m\(^{-2}\) s\(^{-1}\), 24\(^{\circ}\)C.

(B) NPQ kinetics of wild-type and koLhcb4 plants during two consecutive periods of illumination with white light (1200 \( \mu \)mol photons m\(^{-2}\) s\(^{-1}\), 25 min, 24\(^{\circ}\)C) with an 18-min period of darkness in between, as indicated by the white and black bars.

(C) Time course of viola deepoxidation in wild-type and koLhcb4 plants. Leaf discs from dark-adapted leaves were illuminated at 1200 \( \mu \)mol photons m\(^{-2}\) s\(^{-1}\), 24\(^{\circ}\)C (white actinic light). At different times, discs were frozen in liquid nitrogen and total pigments extracted before HPLC analysis.

(D) NPQ kinetics of the wild type, koLhcb4, and mutants retaining single Lhcb4 isoforms. The expression of the isoform Lhcb4.1 (koLhcb4.2 4.3) leads to complete compensation of the NPQ phenotype of koLhcb4; instead, the expression of the Lhcb4.2 isoform partially recovered the quenching ability in the first minutes of induction; nevertheless the mutant failed to fully match the wild type quenching capacity within 8 min of illumination. The presence of the Lhcb4.3 gene did not contribute to NPQ activity. Symbols and error bars show means ± SD (n > 3).
retaining the Lhcb4.3 gene (koLhcb4.1 4.2 mutant) were similar to koLhcb4 (Figure 6). More detailed analysis was performed after 3 and 8 d of treatment by determining the leaf chlorophyll content, a target of photoxidative stress. Mutants koLhcb4 and koLhcb4.1 4.2 underwent a significant reduction of leaf chlorophyll content (Table 2), while mutants retaining either Lhcb4.1 or 4.2 did not show this effect. This indicates that a compensatory accumulation of Lhcb4.1 or 4.2 isoforms can restore photoprotection to the wild-type level. Interestingly, the presence of wild-type alleles of Lhcb4.3 as the only Lhcb4 isoform did not restore photoprotection, thus prompting us to get further insight into the Lhcb4.3 expression. We investigated accumulation of the Lhcb4.3 isoform upon several stress conditions, namely, (1) 500 μmol photons m⁻² s⁻¹, 4°C for 2 d; (2) 900 μmol photons m⁻² s⁻¹, 4°C for 10 d; and (3) 1600 μmol photons m⁻² s⁻¹, 24°C for 10 d. At the end of these treatments, thylakoids were isolated from the wild type, koLhcb4, and koLhcb4.1/4.2 and analyzed by SDS-PAGE/immunoblotting using a polyclonal antibody. Under all experimental conditions, no Lhcb4 immune-reactive bands were detected in the koLhcb4 sample from all the tested conditions. In order to verify that our antibody was indeed able to detect Lhcb4.3, the three recombinant apoproteins were assayed by the anti-Lhcb4 antibody and were recognized with the same efficiency (see Supplemental Figure 7 online). Based on these experiments, we conclude that plants retaining the Lhcb4.3 gene only were unable to accumulate the encoded protein to a significant level in the conditions explored in this work.
Data are expressed as mean ± SD (n = 5); for each genotype, significantly different values (Student’s t test, P = 0.05) with respect to t₀ are marked with an asterisk.

### Table 2. Chlorophyll Content of the Wild Type and Lhcb4 Mutants upon 3 and 8 d of Stress (900 μmol Photons m⁻² s⁻¹, 4°C)

<table>
<thead>
<tr>
<th>Days of Stress</th>
<th>Wild Type</th>
<th>koLhcb4</th>
<th>koLhcb4.2 4.3</th>
<th>koLhcb4.1 4.3</th>
<th>koLhcb4.1 4.2</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>21.7 ± 2.86</td>
<td>19.5 ± 0.16</td>
<td>22.5 ± 1.2</td>
<td>21.5 ± 2.1</td>
<td>22.2 ± 2.6</td>
</tr>
<tr>
<td>3</td>
<td>19.1 ± 0.7</td>
<td>16.5 ± 1.6*</td>
<td>18.9 ± 2.6*</td>
<td>17.7 ± 0.9*</td>
<td>17.2 ± 1.4*</td>
</tr>
<tr>
<td>8</td>
<td>20.3 ± 2.2</td>
<td>13.7 ± 1.7*</td>
<td>19.7 ± 2.0</td>
<td>18.1 ± 2.3</td>
<td>15.8 ± 1.8*</td>
</tr>
</tbody>
</table>

Figure 6. Photoinhibition of the Wild Type and Mutants Retaining a Single Lhcb4 Isoform Exposed to HL and Low Temperature.

Kinetics of Fv/Fm decay were measured on the wild type (WT), koLhcb4, koLhcb4.1 4.2, koLhcb4.1 4.3, and koLhcb4.2 4.3. Whole plants were treated as described for Figure 5A.

### Role of the Zea–Lhcb4 Interaction in Long-Term Membrane Lipid Photoprotection

Earlier work has shown that Lhcb4 exchanges Viola with Zea both in vitro (Bassi et al., 1997; Morosinotto et al., 2002) and in vivo upon HL treatment (Bassi et al., 1993), which leads to an increased activity of several photoprotection mechanisms, including ROS scavenging, improved Chl triplet quenching (Mozzo et al., 2008), and formation of carotenoid radical cations (Holt et al., 2005; Avenson et al., 2008). In order to investigate the role of Zea on Lhcb4-dependent photoprotection, koLhcb4 was crossed with npq1 and selected a genotype lacking both the capacity of producing Zea and Lhcb4. The level of stress caused by HL + cold treatment in the wild type, koLhcb4, and koLhcb4 npq1 was measured from the extent of lipid peroxidation detected by thermoluminescence (TL) (Ducruet and Vavilin, 1999). Figure 7 shows plots of TL amplitudes at different time points during exposure of leaf discs to HL stress (800 μmol photons m⁻² s⁻¹, 4°C). In koLhcb4 genotypes, HL treatment produced higher levels of lipid peroxidation with respect to the wild type, while npq1 behavior was intermediate. Experimental points were fitted using a first-order exponential function (Y = A e⁻∝t), and the resulting equations were used to obtain the differential effect of Zea in the presence or absence of Lhcb4 (Figure 7, inset). Clearly, a much stronger differential effect of the npq1 mutation was observed in koLhcb4 versus the wild type background, implying that photoprotection mediated by Lhcb4 is enhanced in the presence of Zea.

### Structural Analysis of Isolated Grana Membranes

The above results suggest that the koLhcb4 mutant was more strongly affected than genotypes lacking other components of the PSII antenna system. Previous work with koLhcb6 (koCP24) (Kovács et al., 2006; de Bianchi et al., 2008) and koLhcb3 (Damkjaer et al., 2009) has shown that mutations in members of the Lhcb subfamily can affect the macro-organization of PSII supercomplex and PSII-associated regulatory functions. To verify whether a structural effect was induced by lack of Lhcb4, we analyzed both the organization of PSII supercomplexes in grana membranes and isolated PSII supercomplexes by transmission electron microscopy and single particle analysis. Grana membranes from wild-type and koLhcb4 plants exhibit randomly distributed tetrameric stain-excluding particles, corresponding to the PSII-OEC complexes exposed on the luminal surface of dimeric PSII core complexes (Figures 8A and 8B) (Boekema et al., 2000; Betterle et al., 2009). In case of the wild type, a density of PSII complexes in the grana membrane was −1.5 PSII per 1000 nm² (Figure 8B). Samples from koLhcb4 were clearly different (Figure 8A), being characterized by a wider spacing of the tetrameric particles (0.9 PSII particles per 1000 nm²). To further investigate how the lack of Lhcb4 affects the structure of PSII supercomplex, purified PSII supercomplexes were analyzed by single-particle analysis. To this aim, freshly isolated thylakoids were mildly solubilized with 0.3% α-DM followed by rapid fractionation by gel filtration (Damkjaer et al., 2009). The first eluted fractions, containing membrane fragments and PSII supercomplexes, were analyzed by electron microscopy. Single projections of PSII supercomplexes images, identified on the basis of their shape as determined by earlier work (Caffarri et al., 2009), were collected and subjected to image analysis, including translational and rotational alignments, multivariate statistical analysis, and averaging of homogeneous classes. Figure 8C shows the average projection map from a set of 1024 projections of C₅S₂ supercomplexes from plants lacking Lhcb4. Clearly, the projection map is rather similar to the projection map of the wild-type C₅S₂ supercomplex (Figure 8E). In both genotypes, typical features of the core complex were resolved, together with the S-trimer within the peripheral antenna. However, a detailed comparison between the complexes from koLhcb4 (Figures 8C and 8D) and from the wild type (Figure 8E) (adopted from Boekema et al., 2000) showed a different mode of binding of...
the S-trimer to the core complex and, thus, a different overall shape of the C$_2$S$_2$ particles. In addition, electron density, at the position where Lhcb4 binds in wild-type C$_2$S$_2$ particles, is less evident but still present in koLhcb4; however, the space between the core complex and LHCII-S seems to be too small to accommodate an Lhcb subunit, suggesting it could represent a lipid-filled area. The simplest way to interpret this result is that PSII supercomplex associates in the absence of Lhcb4 leading to a reorganization of the overall shape of the particle with partial occlusion of the Lhcb4 area and bending of the LHCII-S mass toward the CP47 complex of the core. To verify this hypothesis, we determined the polypeptide composition and relative amounts in the PSII particles from the wild type versus koLhcb4.

Grana membranes from wild-type and mutant plants were solubilized with low $\alpha$-DM concentration (0.3%) and fractionated by nondenaturing Deriphat-PAGE (see Supplemental Figure 8A online), yielding a pattern superimposable to that previously reported (Caffarri et al., 2009). Fractions containing the C$_2$S$_2$M (Band 10) and C$_2$S$_2$M$_2$ (Band 11) supercomplexes were still detectable in koLhcb4, albeit their amounts were reduced compared with the wild type (14 and 9% of C$_2$S$_2$M and C$_2$S$_2$M$_2$). We chose to analyze C$_2$S$_2$M supercomplexes (Band 10) since this fraction is homogeneous while fractions with higher mobility contain more comigrating complexes (Caffarri et al., 2009) (see Supplemental Figure 9 online). Band 10, and other bands used as a reference, were eluted from the gel and their protein composition was determined by SDS-PAGE, quantitative protein gel blotting analysis, and densitometry (see Supplemental Figures 8B to 8D online). In the C$_2$S$_2$M supercomplex (Band 10) from the wild type, polypeptides of PSII core were found along with Lhcb4, Lhcb5, and LHCl. In the same fraction from koLhcb4, the amounts of Lhcb5 and LHCl with respect to PSII core subunits were the same despite the lack of Lhcb4, implying that this C$_2$S$_2$M supercomplex, although exhibiting a similar overall organization and mobility in green gels, lacked one of its inner subunits. Lhcb4 was not substituted by any other protein component that we could confirm by Coomassie blue staining or immunoblotting (see Supplemental Figure 8 online for details).

**DISCUSSION**

**The Role of Lhcb4/Lhcb6 in the Topology of Grana Membranes and in the Assembly of the PSII-LHCII Supercomplex**

Lhcb4 is one of the six homologous Lhc proteins composing the PSII antenna system. These pigment-protein complexes are...
expected to share a common three-dimensional organization on the basis of the structural data available (Ben Shem et al., 2003; Liu et al., 2004). Previous work with antisense lines has shown a high degree of redundancy among Lhcb subunits; in fact, the PSII supercomplex organization was maintained in the absence of Lhcb1+2 components by overaccumulating Lhcb5 (Ruban et al., 2003). Similarly, plants lacking Lhcb5 and/or Lhcb6 overaccumulated Lhcb1 or Lhcb4 gene products, leading to maintenance of the C2S2 central architecture of the PSII supercomplex (de Bianchi et al., 2008). In the case of koLhcb4, structural redundancy is broken as shown by the destabilization of all supercomplex bands in green gels (Figure 2C; see Supplementary Figure 3 online) and by the differences in the structure of C2S2 particles (Figure 8). These results are consistent with Lhcb4 being the only Lhc subunit that can occupy the position between CP47 and LHCII-M building blocks within the PSII supercomplex (Figures 8C to 8E). This conclusion is supported by the lack of compensation by other gene products in the C2S2M of the mutant (see Supplemental Figures 8B to 8D online).

Isolation of the C2S2 supercomplex from koLhcb4 grana membranes shows that antenna proteins can be associated with the core complex in the absence of Lhcb4 as a docking subunit, which is consistent with the recent isolation of a stable monomeric core with CP26 and the LHCII-S trimer complex (Caffarri et al., 2009). Supercomplexes with a similar shape around the Lhcb4 position were previously found after salt treatment (Boekema et al., 2000). We conclude that in the absence of Lhcb4, PSII does assemble in grana membranes. However, because of the missing subunit, the complex is less stable and assumes a different overall structure with a low-density area located in the position where Lhcb4 is present in the wild type, which is likely occupied by lipids. As for the organization of the outer shell of the PSII antenna system, which is formed by LHCII-M, LHCII-L and Lhcb6, it appears to be strongly affected in koLhcb4. Titration of the different Lhc proteins with respect to PSII RC showed that the Lhcb6 complex was completely missing in koLhcb4 (Figure 2B). Since the Lhcb6 messenger level was unchanged with respect to the wild type, this implies that removal of Lhcb4 decreases Lhcb6 stability. This is consistent with Lhcb4 being the docking site for Lhcb6 (Andersson et al., 2001; Caffarri et al., 2009), both participating in a pentameric complex called B4C (band 4 complex), which connects inner and outer antenna moieties (Bassi and Dainese, 1992; Betterle et al., 2009). One component of B4C, interacting with Lhcb4 and/or Lhcb6, is Lhcb3 (Betterle et al., 2009), which is still present in koLhcb4 membranes; this result is consistent with the observation that, besides Lhcb4, Lhcb3 can interact with Lhcb1 and Lhcb2 in complexes containing LHCII-M (see Supplemental Figure 8B online, lanes B9 to B11), while it is absent in complexes containing only LHCII-S, such as CS complexes (lanes B6). Less stable C2S2M2 supercomplexes can still form in the koLhcb4 mutant, which lacks both Lhcb4 and Lhcb6, although their molecular interactions are rather weak and the abundance of this complex is only 5% with respect to the wild type. Previous work with koLhcb6 showed that a large part of the outer antenna formed by LHCII-M and LHCII-L is not directly bound to PSII supercomplexes; rather, trimers L and M form LHCII-only domains that segregate from arrays of C2S2 particles (de Bianchi et al., 2008). No such arrays were observed in koLhcb4 membranes (Figure 8A) nor was PQ diffusion restricted (Figure 3), suggesting that the altered shape of C2S2 particles and/or their instability prevents cooperative interactions with arrays. Besides this, the distance between neighbor PSII centers is higher than in the wild type (Figures 8A and 8B), implying a higher number of LHCII trimers is interposed between PSII centers.

**Consequences for Photosynthesis: Light Harvesting and ET**

The loss of Lhcb4 in *Arabidopsis* did not strongly affect growth rate and pigment compositions under control light conditions (Figure 3, Table 1). Indeed, linear and cyclic ET rates were similar to those in wild-type plants, as well as the functional antenna size. However, alterations in the PSII macrostructure in mutant plants did result in differences in chlorophyll fluorescence parameters (Table 1). Increased F0 (Table 1) showed that the efficiency of excitation energy transfer from the antenna to the PSII RC is decreased in mutant plants. In grana membranes from the wild type (Figure 8B), the distribution of PSII particles is homogeneous through the whole surface. This is not the case for koLhcb4, whose grana membranes contained discrete patches of LHCII trimers interspersed by fewer, randomly distributed PSII cores (Figure 8A); therefore, in some discrete areas of koLhcb4 grana membranes, the LHCII/PSII core ratio is increased. In these domains, LHCII fluorescence is probably not efficiently photochemically quenched, thus yielding higher F0 (Table 1). This is consistent with the results obtained in Ko mutants for Lhcb5 and Lhcb6 (Kovács et al., 2006; de Bianchi et al., 2008), while no such effect was observed in asLHCII (Andersson et al., 2003). Increased F0 suggests that vacancy of Lhcb4 forces the excitation energy to follow restricted pathways from the peripheral antenna to the core; indeed, the migration time of excitations from antenna to the PSII RC was significantly reduced in koLhcb4 with respect to the wild type (van Oort et al., 2010). Thus, despite the fact that fluorescence induction in the presence of DCMU showed no changes in functional antenna size, the number of LHCII per PSII RC in grana membranes was higher (Figures 8A and 8B). We conclude that in the absence of a well-organized PSII-LHCII supercomplex, the efficiency of excitation energy trapping in koLhcb4 was lower than in the wild type and that the compensatory increase in peripheral LHCII complexes (Figures 2A and 2B) contributed with lower efficiency to light harvesting.

Although functional measurements indicate that there was no major perturbation of PSII and PSI functions, a slight increase in the PSI/PSII ratio was observed in koLhcb4 (Figure 2). The consequences of a lower PSII level were not strong: the parameter \( F_{m}/F_{0}\), expressing the average fraction of open RCs per mm2, was lower in the mutant, which is likely due to the fact that the maximal rate of photochemical reduction of P700 with respect to the wild type (Figure 3F). Since both LEF and CEF are not affected in the mutant, we propose that this effect could be related to the higher PSII functional antenna size of the mutant in
HL. Indeed, when measurements were performed by changing the quality of actinic light (PSII rather than PSI+PSII light), differences in P700 oxidation rate between wild-type and mutant leaves disappeared (Figure 3G), implying the increase of PSI functional antenna size in HL is due to PSI components. This is consistent with an increased level of spillover in koLhcb4 at higher light intensity. This suggestion was verified by 77K fluorescence spectroscopy (see Supplemental Figure 5 online), showing that HL treatment induces an increased contribution of LHCII to PSI excitation in koLhcb4 but not in the wild type. Since the amplitude of the state I–state II transition is the same in the wild type and in the mutant and, moreover, the stn7 kinase is inhibited in HL (Tikkkanen et al., 2006), this spectral change cannot be attributed to state transitions. Instead, we interpret these results as evidence of spillover of excitation from LHCII to PSI. We suggest that this effect ensues from the lower stability of PSII-LHCII supercomplexes in the mutant, with the disappearance of the B4 complex (Figure 2C) and the faster kinetic of state transitions (Figures 3D and 3E). In koLhcb4, an overall weakening of interactions between the PSII core and LHCII would make weakly bound LHCII at the grana margins able to contribute to PSI antenna size particularly in HL, when grana membrane reorganization occurs with segregation of LHCII-rich domains (Kirchhoff, 2008; Betterle et al., 2009; Johnson et al., 2011).

Consequences for Regulation of Light Harvesting: State Transitions

State transition is the mechanism by which the complement of LHCII is balanced between PSII and PSI, depending on the reduction state of the intermediate electron carrier PQ, through its reversible phosphorylation, which induces its disconnection from PSII and binding to PSI (Allen and Nilsson, 1997; Jensen et al., 2000). Lhcb4 can be phosphorylated in monocots (Bergantino et al., 1995; Bergantino et al., 1998) and in Arabidopsis (Hansson and Vener, 2003). In Chlamydomonas reinhardtii, P-Lhcb4 was found to be connected to PSI-LHCI supercomplexes in state II conditions (Takahashi et al., 2006), and Lhcb4-depleted cells showed changes in state transitions (Tokutsu et al., 2009). Thus, changes in state transitions can be expected in Lhcb4-less plants. Results displayed in Figures 3D and 3E show that koLhcb4 was not affected in its state transition total activity, in agreement with the similar reduction state of the PQ pool at all light intensities (Figure 3B) (Bellafiore et al., 2008). We observed that in koLhcb4 the fluorescence changes, upon switching off the far-red light, are faster than in the wild type, implying that the transiently reduced state of the free PQ pool is more promptly relaxed by migration of the LHCII to PSI. The same state transition phenotype has been described for koLhcb6 and koLhcb5 Lhcb6 plants (de Bianchi et al., 2008), and this is a clear indication that the connection between the PSII core and the bulk trimeric LHCII is weaker in these mutants with respect to the wild type. In the case of the koLhcb6 plants, the faster state transition was attributed to the displacement of the M-trimer from the PSI macrostructure, thus enhancing its migration to PSI. We suggest that, in the koLhcb4 mutant, the weaker binding of S trimer (and, possibly, of the M trimer) and the higher amount of weakly connected LHCII trimers available increase the probability of migrating toward PSI upon phosphorylation. In C. reinhardtii, state transitions do not only fulfill the role of balancing light absorption between photosystems, they also increase PSI electron flow at the expense of PSII, acting as a switch between LEF and CEF (Vallon et al., 1991). This was not the case in Arabidopsis since, despite an effect on state transitions (Figures 3D and 3E), no changes in linear (Figure 3A) versus CEF rates (see Supplemental Figures 4A and 4B online) were observed.

Nonphotochemical Fluorescence Quenching

Nonphotochemical dissipation of excess energy is affected in koLhcb4 plants with respect to the wild type: mutant plants showed a delayed rise and a lower NPQ amplitude after 8 min of light (Figure 4A). Upon prolonged illumination and/or upon repeated light treatments (Figure 4B), the amplitude of NPQ was similar to that of the wild type. A similar effect was previously observed in the koLhcb5 Lhcb6 double mutant (de Bianchi et al., 2008). Since Lhcb6 is destabilized in koLhcb4 (Figure 2), this mutant phenocopies a double koLhcb4 Lhcb6 mutant. NPQ kinetic may be modified by changes in the trans-thylakoid ΔpH gradient (de Bianchi et al., 2008), by changes in the level of the pH sensor PsbS (Li et al., 2002, 2004), or by changes in the number/relative abundance of protein subunits hosting quenching sites (Bonente et al., 2008). Changes in luminal pH appear unlikely; in fact, the kinetics of Zea synthesis, catalyzed by the pH-dependent enzyme VDE (Yamamoto and Higashi, 1978), were the same in wild-type and mutant leaves (Figure 4C). Moreover, the level of PsbS was unchanged (Figure 2B), leaving modification in the abundance and identity of quenching sites localized in Lhcb proteins as the most likely cause for the observed phenotype. We interpret our results in the framework of the recently proposed model (Miloslavina et al., 2008; de Bianchi et al., 2010), based on the formation of quenching sites within each of two distinct domains in grana discs: (1) C2-S2 particles (containing PSII RC, Lhcb4, Lhcb5, and LHCII-S) and (2) the peripheral antenna, including Lhcb6, LHCII-M, and LHCII-L (Miloslavina et al., 2008; Betterle et al., 2009), which segregate because of the action of PsbS. Zea-dependent quenching activity has been detected within monomeric Lhcb4, b5, and b6 in detergent solution (Avenson et al., 2008), while, in LHCII, it was activated by aggregation and was independent from Zea (Ruban et al., 2007). The similar NPQ kinetics of koLhcb4 and of koLhcb5 Lhcb6 can thus be explained because they both retain a quenching site within the C2-S2 domain; the faster quenching kinetic upon repeated illumination is due to the enhancement by Zea of the quenching activity (Niyogi et al., 1998) associated with Lhcb4 (Figure 4) or Lhcb5. This is consistent with the capacity of exchanging Viola with Zea in binding site L2 observed in monomeric Lhcb4 (Morosinotto et al., 2002; Wehner et al., 2006) rather than in trimeric LHCII. In koLhcb5 Lhcb6, the recovery of NPQ upon the initial delay is faster and more complete than in koLhcb4, implying that Lhcb4 is more active as a quencher than Lhcb6 and Lhcb5 (de Bianchi et al., 2008). If there are two distinct quenching domains within PSII antenna (Miloslavina et al., 2008) and monomeric Lhcb4-6 proteins are the only sites of quenching in vivo, the peripheral antenna domain should remain unquenched in both koLhcb5 Lhcb6 (de Bianchi et al., 2008) and...
koLhcb4 because they both lack Lhcb6. Since both genotypes reach NPQ levels similar to the wild type, although with a delayed kinetic, it is likely that some kind of quenching does occur in the peripheral antenna domain disconnected from $S_{2}$ particles in these genotypes. Previous work with koLhcb6 has shown that this component has a quenching effect on the major LHCII antenna (van Oort et al., 2010); here, we observe that, even in the absence of Lhcb4, the overall quenching activity is high, consistent with quenchers being activated in both PSII antenna domains, and conclude that LHCII is likely to contribute to NPQ in vivo, in spite of the fact that it does not exhibit the spectroscopic and structural features of quenching sites (Ahn et al., 2008; Avenson et al., 2008). Thus, it likely activates quenching by a different mechanism (Ruban et al., 2007). Consistent with this view is the finding that a red-shifted fluorescence lifetime component was observed in vivo under NPQ conditions (Müller et al., 2010), which has properties similar to LHCII aggregated in vitro.

While differences in qE were minimized by prolonged/repeated illumination, we observed that the qL component, which is responsible for the slowly relaxing component of quenching, is equally decreased upon short/long or repeated light treatments (Figure 4B). Previous work suggested that qL was specifically due to Zea binding to Lhc65 (Dall’Osto et al., 2005); instead, removal of Lhc65 has a negligible effect on qL amplitude, since the double mutant koLhcb5 Lhcb6 did not further decrease its qL level (de Bianchi et al., 2008). Here, we show that removal of Lhc4 reduces the extent of qL, implying that qL is modulated by both Lhcb4 and Lhcb5.

**Consequences for Photoprotection: Resistance to Photooxidative Stress Is Decreased in koLhcb4 Plants with Respect to the Wild Type**

Lhcb4-less plants showed a reduced photoprotection capacity when exposed to high irradiance at low temperature. The highest sensitivity to photooxidative stress of koLhcb4, among all other Lhcb KO mutants, is consistent with the higher reduction in fitness of plants lacking Lhcb4 (Andersson et al., 2001; Ganeteg et al., 2004), supporting the importance of this gene product for PSII performance and chloroplast photoprotection. The structural integrity of photosynthetic supramolecular complexes is essential for the resistance to photooxidative stress (Horton and Ruban, 2005), although the reasons are not completely clear. Depletion of light-harvesting antennae has been reported to favor photoinhibition of both PSI (Alboresi et al., 2009) and PSII (Kim et al., 2009; Dall’Osto et al., 2010). KoLhcb4 mutant plants are more sensitive to photooxidative stress than the wild type (Figure 5), and this effect is associated with increased production of $O_{2}$. This effect cannot be ascribed to a pleiotropic effect on photosynthetic ET efficiency, since lack of Lhcb4 does not significantly affect either the rate or the regulation of photosynthetic ET (Figures 3A to 3C). A putative mechanism for photo-inhibition is the reduction of $Chl^{*}$ dissipation (qE and qL) (Johnson et al., 2007); however, in koLhcb4, qE activity is affected at the onset of illumination, while it is similar to the wild type on a longer timescale. Since our photoinhibitory light treatment was performed at constant light intensity, we conclude that photosensitivity of koLhcb4 plants is not due to differences in qE or qL.

The connection between the PSII core complex and outer LHCII was partially impaired. Indeed, a steady increase in $F_{v}/F_{m}$ was measured in koLhcb4 plants with respect to the wild type (Table 1); a higher $Chl^{*}$ level might lead to $Chl^{*}$ formation through intersystem crossing. Although we cannot exclude that the inefficient connection of LHCII to PSII RC could contribute to the higher photosensitivity of koLhcb4, we notice that the koLhcb5 Lhcb6 double mutant was as resistant as wild-type plants (Figure 5A) despite a $F_{v}/F_{m}$ reduction comparable to that of Lhcb4-less plants (de Bianchi et al., 2008). Since photoprotection in koLhcb5 Lhcb6 plants was similar to that in the wild type (Figure 5A), neither the increase in $F_{v}$ per se nor the absence of Lhcb6 can be the cause of higher photosensitivity of koLhcb4 plants. Instead, this effect appears to be specific for the absence of Lhcb4, implying that this gene product is of particular importance in providing PSII photoprotection (Figure 5A). Indeed, Lhcb4 appears to be the most conserved among Lhc proteins associated with PSII (Koziol et al., 2007), and it maintains its stoichiometry with the RC even under stressful growth conditions that lead to a depletion of Lhcb1, 2, and 6 (Ballottari et al., 2007). The most obvious peculiarity of Lhcb4 is that it cannot be replaced in PSII supramolecular architecture by other Lhcb5 and is thus required for allowing structural integrity of PSII (Boekema et al., 1999) (Figures 8C to 8E). Thus, the higher photosensitivity in koLhcb4 could be the effect of a less stable PSII supercomplex; indeed, the extreme photosensitivity of Arabidopsis mutants such as ch1 (Dall’Osto et al., 2010) can be ascribed to the absence of light-harvesting complexes surrounding the PSII RC, leaving the PSII core complex exposed to the lipid phase, where radical chain reactions of peroxy-lipids occur during photooxidative stress.

Measurement of lipid peroxidative damage by thermoluminescence in intact leaves of wild type versus koLhcb4 and koLhcb4 npq1 plants showed that Zea and Lhcb4 are synergistically active in protection from photodamage (Figure 7). Therefore, it appears that a specific effect for Zea in providing efficient photoprotection is preferentially amplified when bound to Lhcb4. This observation is consistent with a previous report of a protective effect of Zea (Dall’Osto et al., 2010) in addition to qE enhancement (Niyogi et al., 1998) and to ROS scavenging in the lipid phase (Havaux and Niyogi, 1999; Baroli et al., 2003, 2004; Havaux et al., 2007). Upon light-induced synthesis, Zea enters the protein-bound xanthophyll pool by binding to the L2 site of minor Lhc (Avenson et al., 2008; Jahns et al., 2009) and to the V1 site of LHCII (Caffarri et al., 2001), in addition to qE protection in the lipid phase. The molecular mechanism of enhanced resistance to photooxidation by Zea is not yet clear. The hypotheses include (1) draining excitation energy from the PS core by xanthophylls bound at the interface with Lhcb5; (2) quenching of $Chl^{*}$ energy from RC, possibly by the Dexter exchange mechanism (Nayak et al., 2002) by Zea located in sites L2 on specific Lhcs; and (3) preferential scavenging of $O_{2}$ by Zea into site V1 of LHCII (Johnson et al., 2007). Although all of these mechanisms are plausible for explaining the higher photosensitivity of koLhcb4, their relative contribution needs further investigation.
Effect of the Accumulation of the Individual Isoforms on koLhcb4 Phenotype

The attenuation of the F0 and NPQ phenotypes produced by the triple koLhcb4 mutations is different in the three double koLhcb4 mutants. Retention of Lhcb4.1 restores F0/Fm and NPQ to wild-type levels, while Lhcb4.2 is only partially effective in this function, Lhcb4.3 is completely inefficient (Figures 4D and Figure 6, Table 1). Indeed, plants retaining only Lhcb4.3 show the same reduction in Chl a/b ratio (Table 1), increased photosensitivity (Figure 6), and delayed NPQ kinetic as the triple koLhcb4. Although we cannot exclude that Lhcb4.1, 4.2, and 4.3 gene products are intrinsically different, the penetration of the phenotype appears to be essentially due to the level of the gene product. Indeed, levels of Lhcb4.1, Lhcb4.2, and Lhcb4.3 in the three double mutant are 1, 0.6, and 0, respectively, per PSII RC (see Supplemental Table 2 online). In fact, although Lhcb4.3 transcription is lightly increased under stress conditions (Alboresi et al., 2011), we were unable to detect the corresponding protein even by keeping plants under a variety of stress conditions, implying it is either not translated or rapidly turned over.

The Lhcb4.3 gene appeared recently in plant genome. Its sequence is not present in algal genomes, and no orthologs were found in the moss Physcomitrella patens; therefore, it appeared only at later stages in the evolution of the green lineage (Alboresi et al., 2008). It seems unlikely that a new gene conserved in higher plants has no function or is not translated at all. EST data from Arabidopsis show that Lhcb4.3 is expressed at low levels (Jansson, 1999), and its transcription seems to be confined to dicots (Goff et al., 2002; Yu et al., 2002); furthermore, clustering (Jansson, 1999), and its transcription seems to be confined to dicots (Goff et al., 2002; Yu et al., 2002); furthermore, clustering analysis of Lhc superfamily expression data (Klimmek et al., 2006) confirmed that Lhcb4.3 is regulated in an opposite way with respect to Lhcb4.1 and Lhcb4.2. The putative Lhcb4.3 protein is shorter than other Lhcb4 isoforms, and its sequence is considerably different from both Lhcb4.1 and Lhcb4.2 isoforms, suggesting it might have a different function in the stress conditions in which this gene is actively transcribed. However, we cannot exclude that the Lhcb4.3 isoform might accumulate in special and still uninvestigated environmental/developmental conditions; so far, the role of this isoform remains elusive.

We have shown that Lhcb4 has a specific function in protecting PSII from photoinhibition, which accounts for the increased sensitivity of koLhcb4 to HL stress. This gene product, different from any other PSII antenna component, cannot be replaced by homologous subunits in the PSII-LHCCI supramolecular complex. This leads to the formation of incomplete PSII particles with a gap in the position normally occupied by Lhcb4, a modified shape and a lower stability. This is consistent with evidence that a core group of antenna proteins developed prior to green algal diversification (Koziol et al., 2007) and included Lhcb4 protein associated with PSII RC. The weaker interaction between PSII-LHCCI components leads to a faster migration of LHCCI complexes to the stroma membrane upon phosphorylation, similar to the case of koLhcb6 and koLhcb3. Although PSII activity is not impaired in moderate light conditions, there is a lower level of PSII RC in grana membranes, probably because the PSII core complex is less strongly retained within grana discs, because one of the two monomeric Lhcb5 that bridges it to LHCCI is missing. Under stress conditions, koLhcb4 is photoinhibited, and this effect is selectively enhanced in the absence of Zea, which is active in preventing the synthesis of ROS species and promoting their scavenging. By binding in between LHCII trimers and the CP47 subunit of PSII, Lhcb4 is crucial for the protection of PSII RC from ROS produced during photosynthesis either by neighbor-damaged PSII complexes (Krieger-Liszkay et al., 2008) or by overexcited antennae (Santabarbara et al., 2002; Mozzo et al., 2008). Thus, the absence of a specific antenna subunit, although it does not restrict light harvesting and photosynthetic ET rate, impairs PSII photoprotection capacity through its effect on the assembly of supercomplexes. This is an example of optimization of the building blocks of photosynthetic complexes and the tuning of their interactions with each other toward overcoming the inhibitory effect of increasing oxygen concentration over evolution time. It is worth noting that functional alterations described in koLhcb4 plants likely result from the reduced ability of PSII macroassemblies to undergo proper organization in low versus high light, thus confirming the crucial role of Lhcb4 in modulating structural and functional flexibility of grana membranes.

METHODS

Plant Material and Growth Conditions

Arabidopsis thaliana T-DNA insertion mutants (Columbia ecotype) GK Line ID282A07 (insertion into the Lhcb4.1 gene), SAIL_910_D12 (insertion into the Lhcb4.2 gene), and SALK_023779 (insertion into the Lhcb4.3 gene) were obtained from NASC collections (Alonso et al., 2003). Homozygous plants (Beltere et al., 2009) were identified by PCR analysis using the following primers: forward 5′-TACCAGATAACGCAAGTATTATAG-3′ and reverse 5′-CACATGATAATGTTTAAAGTGGAG-3′ for the Lhcb4.1 sequence, 5′-CATCATACACATTGTACTGATCCAG-3′ for the Lhcb4.2 sequence, 5′-GCCCTTCAGAATGTTAAGATGTCCTGTC-3′ for the Lhcb4.3 sequence, and 5′-GACATACCTGGTAAAGGTTGACACAG-3′ for reverse 5′-GTGAGCTGATCCATGGAAGGTGGG-3′ for the Lhcb4.1 sequence, 5′-GACATACCTGGTAAAGGTTGACACAG-3′ for reverse 5′-GTGAGCTGATCCATGGAAGGTGGG-3′ and reverse 5′-GACATACCTGGTAAAGGTTGACACAG-3′ for the Lhcb4.3 sequence. Genotypes koLhcb4, koLhcb4.1, koLhcb4.2, koLhcb4.3, and koLhcb4.1.4.3 were obtained by crossing single mutant plants and selecting progeny by PCR analysis. For RT-PCR, total RNA was isolated from 4-week-old plants with the Trizol protocol. Reverse transcription was performed using M-MLV reverse transcriptase with oligo(dT) primer and 1.5 µg of total RNA. For normalization purposes, actin2 was chosen as an endogenous control.

The primers used were as follows: 5′-ATGGTGTCCCGGTATTCATCATTCC-3′ and 5′-CTGAAAGGCGAAAACCA-AGAATC-3′ for Lhcb4.1 cDNA; 5′-GTGCCATTAGCTCCATCCATTC-3′ and 5′-GACCTTTGCGAATTGGAGAG-3′ for koLhcb4 cDNA; 5′-AGGCTTCAATGAGTATGTTTCGCTGG-3′ and 5′-GACATACCTGGTAAAGGTTGACACAG-3′ for the Lhcb4.3 sequence. Genotypes koLhcb4, koLhcb4.1, koLhcb4.2, koLhcb4.3, and koLhcb4.1.4.3 were obtained by crossing single mutant plants and selecting progeny by PCR analysis. For RT-PCR, total RNA was isolated from 4-week-old plants with the Trizol protocol. Reverse transcription was performed using M-MLV reverse transcriptase with oligo(dT) primer and 1.5 µg of total RNA. For normalization purposes, actin2 was chosen as an endogenous control.

The Arabidopsis T-DNA insertion mutant koLhcb3 (SALK_020342) from NASC was obtained by selecting progeny by immunoblotting using a specific antibody against the Lhcb3 subunit; asLHCCI (Anderson et al., 2003) was obtained from the NASC. The double mutant koLhcb4 npq1 was obtained by crossing single mutants and selecting progeny by immunoblotting (with the Lhcb4 antibody) and HPLC upon HL treatment of leaves. Insertion mutants koLhcb6 and koLhcb5 Lhcb6 were isolated as previously described (de Bianchi et al., 2008). Mutants were
grown for 5 weeks at 100 μmol photons m\(^{-2}\) s\(^{-1}\), 23°C, 70% humidity, and 8 h of daylight.

**Stress Conditions**

Short-term HL treatment was performed at 1200 μmol photons m\(^{-2}\) s\(^{-1}\), 45 min, and room temperature (24°C), to measure the kinetics of Zea accumulation, on detached leaves floating on water. Samples were rapidly frozen in liquid nitrogen prior to pigment extraction. Longer photooxidative stress was induced by exposing whole plants to either 550 μmol photons m\(^{-2}\) s\(^{-1}\) at 4°C for 2 d, 900 μmol photons m\(^{-2}\) s\(^{-1}\) at 4°C for 10 d, or 1600 μmol photons m\(^{-2}\) s\(^{-1}\) at 24°C for 10 d. Light was provided by halogen lamps (Focus S; Prisma) and filtered through a 2-cm recirculation water layer to remove infrared radiation.

**Pigment Analysis**

Pigments were extracted from leaf discs, either dark adapted or HL treated, with 85% acetone buffered with Na\(_2\)CO\(_3\), and then separated and quantified by HPLC (Gilmore and Yamamoto, 1991).

**Membrane Isolation**

Functional thylakoids were isolated from dark-adapted or HL-treated leaves as previously described (Casazza et al., 2001).

**Gel Electrophoresis and Immunoblotting**

SDS-PAGE analysis was performed with the Tris-Tricine buffer system (Schägger and von Jagow, 1987), with the addition of 7 M urea to the running gel to separate Lhcb4 isoforms. Non-denaturing Deriphat-PAGE was performed following the method developed by Peter et al. (1991) with modification described by Havaux et al. (2004). Thylakoids concentrated at 1 mg/mL chlorophylls were solubilized in a final concentration of 1% α/β-DM, and 25 mg of chlorophyll were loaded in each lane. For immunotratification, thylakoid samples corresponding to 0.25, 0.5, 0.75, and 1 μg of chlorophyll were loaded for each sample and electroblotted on nitrocellulose membranes, and proteins were detected with alkaline phosphatase-conjugated antibody according to Towbin et al. (1979). Signal amplitude was quantified (n = 4) using GelPro 3.2 software (Bio-Rad). To avoid any deviation between different immunoblots, samples were compared only when loaded in the same slab gel.

**In Vivo Fluorescence and NQ Measurements**

PSII function during photosynthesis was measured through chlorophyll fluorescence on whole leaves at room temperature with a PAM 101 fluorometer (Heinz-Walz) (Andersson et al., 2001), a saturating light pulse of 4500 μmol photons m\(^{-2}\) s\(^{-1}\), 0.6 s, and white actinic light of 100 to 1100 μmol photons m\(^{-2}\) s\(^{-1}\), supplied by a KL1500 halogen lamp (Schott). NQ, \(\phi_{Qs}\), \(\phi_{psii}\), \(\phi_{psii}^{max}\), \(\phi_{Qs}^{max}\), and LET were calculated according to the following equations (Van Kooten and Snel, 1990; Baker, 2008; NQ = \(\frac{\text{F}(\text{m}) - \text{F}(\text{n})}{\text{F}(\text{m})}\), \(\phi_{psii} = \text{F}(\text{m}) - \text{F}(\text{n})\), \(\phi_{psii}^{max} = \text{F}(\text{m}) - \text{F}(\text{n})\), \(\phi_{Qs} = \text{Q}_{s}\), \(\phi_{Qs}^{max} = \phi_{Qs}\), \(\phi_{psii}^{PAR} = \text{PAR}_{\text{abs}} - \text{fraction}_{\text{psii}}\), where \(\phi_{psii}^{PAR}\) is the minimal fluorescence from dark/light-adapted leaf, \(\phi_{psii}^{PAR}\) is the maximal fluorescence from dark/light-adapted leaves measured after the application of a saturating flash, \(\phi_{Qs}\) the stationary fluorescence during illumination, and \(\phi_{psii}^{PAR}\) the photosynthetic active radiation; \(\text{fraction}_{\text{psii}}\) was measured by quantitative immunoblot: wild type, 0.50 ± 0.01; koLhcb4, 0.47 ± 0.06.

State transition experiments were performed using whole plants according to established protocols (Jensen et al., 2000). Preferential PSI excitation was provided by illumination with blue light (40 μmol photons m\(^{-2}\) s\(^{-1}\)), excitation of PSI was achieved using far-red light from an LED light source (Heinz-Walz; 102-FR) applied for 15 min simultaneously with blue light, \(\text{F}_{\text{n}}\) level in state I (\(\text{F}_{\text{n}}\)) and state II (\(\text{F}_{\text{n}}\)) was determined at the end of each cycle by the application of a saturating light pulse. The parameter \(\text{q}^T\) (PSII cross-section changes) was calculated as \(\frac{\text{F}(\text{m}1) - \text{F}(\text{n})}{\text{F}(\text{m})}\) (where \(\text{F}_{\text{m}1/2}\) is the maximal fluorescence yield in state I/II).

Fluorescence induction kinetics was recorded with a home-built apparatus to measure functional antenna size on leaves. \(\text{S}_{\text{th}}/\text{F}_{\text{max}}\) was calculated from variable fluorescence curves induced with green light (1100 μmol photons m\(^{-2}\) s\(^{-1}\)) (Strasser et al., 1995). For measurements of PSII functional antenna size, variable fluorescence was induced with a green light of 15 μmol photons m\(^{-2}\) s\(^{-1}\), on thylakoids (10 μg Chls/mL) in a measuring buffer containing 10 mM HEPES, pH 7.8, 5 mM MgCl\(_2\), 30 μM DCMU, and 50 μM nigericin. The reciprocal of time corresponding to two-thirds of the fluorescence rise (\(\text{T}_{2/3}\)) was taken as a measure of the PSII functional antenna size (Malin et al., 1981). For measurements of the PSII repair process, whole plants were illuminated at 900 μmol photons m\(^{-2}\) s\(^{-1}\), 4°C for 4 h to induce photooinhibition of PSII, and restoration of the \(\text{F}_{\text{n}}/\text{F}_{\text{m}}\) ratios was subsequently followed at irradiances of 20 μmol photons m\(^{-2}\) s\(^{-1}\), 4°C (Aro et al., 1994).

**Analysis of P700 Redox State**

All in vivo spectroscopic measurements were performed using a LED spectrophotometer (JTS10; Biologic Science Instruments) in which absorption changes are sampled by weak monochromatic flashes (10-nm bandwidth) provided by LEDs. P700\(^+\) reduction following a flash was assayed as described by Golding et al. (2004). Leaves were infiltrated with 50 μM DCMU in 150 mM sorbitol. A 400-ms saturating flash of red light (\(\lambda_{\text{max}} = 630 \text{ nm}; 3000 \text{ μmol photons m}^{-2} \text{ s}^{-1}\)) was delivered to the leaf, and changes in absorbance at 705 nm were used to measure the kinetics of P700\(^+\) reduction. Oxidized P700 (\(\text{F}_{\text{S}2}\)) was recorded during far-red light illumination (2500 μmol photons m\(^{-2}\) s\(^{-1}\), \(\lambda_{\text{max}} = 720 \text{ nm}\)). The level of oxidized P700 in the leaf (\(\Delta\lambda\)) was determined during illumination with either orange light (630 nm, from 50 to 980 μmol photons m\(^{-2}\) s\(^{-1}\), \(\lambda_{\text{max}} = 720 \text{ nm}\)) or far-red light (720 nm, from 200 to 2500 μmol photons m\(^{-2}\) s\(^{-1}\)) (Zygadlo et al., 2005). Antenna size of PSI was estimated according to Kim et al. (2009).

**Analysis of Cytochrome b6f Redox Kinetics**

Absorption transients were measured on thylakoids (30 μg Chls/mL) in a measuring buffer containing 10 mM HEPES, pH 7.8, 5 mM MgCl\(_2\), 1 mM sodium ascorbate, 20 μM nigericin, and 50 μM methylviologen (Kirchhoff et al., 2000). Cytochrome b6f redox changes were measured in continuous light (630 nm, 300-ms pulse, 560 μmol photons m\(^{-2}\) s\(^{-1}\)) at four wavelengths: 548, 554, 563, and 573 nm; redox changes of cytochromes b and f were calculated according to Joliot and Joliot (1984).

**Analysis of Steady State Proton Flux across Thylakoid Membranes**

Light-induced pmf was estimated on intact leaves, from DIRK changes in absorbance associated with the ECS at 520 nm, as described previously (Livingston et al., 2010). ECS was induced with orange (630 nm) actinic intensities ranging from 15 to 960 μmol photons m\(^{-2}\) s\(^{-1}\). Relative values of steady state proton flux across the thylakoid membrane (\(\nu_{\text{r}}\)) was estimated from the initial slope of the ECS decay upon rapid light–dark transitions.

**Gas Exchange Measurement**

Measurements of \(O_2\) evolution in saturating CO\(_2\) were performed using an S101 \(O_2\) electrode (Qubit System). Light response curves were determined using broadband red light.
Determination of the Sensitivity to Photooxidative Stress

Photooxidative stress was induced in detached leaves by HL treatment at low temperature. Detached leaves floating on water were exposed to 1500 μmol photons m⁻² s⁻¹ for 8 h, in a growth chamber at 4°C, and then immediately frozen in liquid nitrogen. Photooxidative stress was assessed by measuring MDA formation as an indirect quantification of lipid peroxidation (Havaux et al., 2005). Lipid peroxidation was measured on whole plants by TL with a custom-made apparatus (Ducruet, 2003). The amplitude of the TL peak at 135°C was used as an index of lipid peroxidation (Havaux, 2003). Measurements of singlet oxygen production from leaves were performed with SOSG (Molecular Probes), a O₂-specific fluorogenic probe, as described by Dall'Osto et al. (2010).

Electron Microscopy

Samples were negatively stained with 2% uranyl acetate on glow discharged carbon-coated copper grids. Electron microscopy was performed on a Philips CM120 electron microscope equipped with a LaB₆ tip operating at 120 kV. Images were recorded with a Gatan 4000 SP 4K slow-scan CCD camera at 80,000 magnification at a pixel size (after binning the images) of 0.375 nm at the specimen level with GRACE software for semiautomated specimen selection and data acquisition (Oostergetel et al., 1998). Single-particle analysis of a data set of 1350 PSII particles was performed using Groningen Image Processing software, including multireference and no-reference alignments, multivariate statistical analysis, classification, and averaging of homogeneous classes (van Heel et al., 2000).

Accession Numbers

Sequence data from this article can be found in the Arabidopsis Genome Initiative or GenBank/EMBL databases under accession numbers At5g10130 (Lhcb4.1), At3g08940 (Lhcb4.2), At2g40100 (Lhcb4.3), At5g54070 (Lhcb3), At4g10340 (Lhcb5), At1g15820 (Lhcb6), At1g08550 (npq1), and At3g18780 (actin2). The KO lines mentioned in the article can be obtained from the NASC under the stock numbers N376476 (npq1), and At3g18780 (actin2). The KO lines mentioned in the article can be obtained from the NASC under the stock numbers N376476 (npq1), and At3g18780 (actin2).

Supplemental Data

The following materials are available in the online version of this article.

Supplemental Figure 1. Transmission Electron Micrographs of Plastid from Mesophyll Cells of the Wild type and Mutants.

Supplemental Figure 2. Induction Curves of Chl Fluorescence Measured on Thylakoids Treated with DCMU.

Supplemental Figure 3. Biochemical Characterization of koLhcb4 Mutants.

Supplemental Figure 4. P700 Measurements: Cyclic Electron Transport, PSI Functional Antenna Size, and Cytochrome b6f Redox Kinetics.

Supplemental Figure 5. Low-Temperature Fluorescence Spectra of Chloroplasts.

Supplemental Figure 6. PSII Repair Efficiency under Photoxidative Stress.

Supplemental Figure 7. Insights on the Lhcb4.3 Expression and Function.

Supplemental Figure 8. Insights on the PSII Supercomplex Composition.

Supplemental Figure 9. Schematic Panel of Wild-Type and koLhcb4 PSII Supercomplex Composition.

Supplemental Table 1. Quantitative Analysis of Thylakoid Morphological Traits.

Supplemental Table 2. Polypeptide Composition of Thylakoid Membranes from Wild-Type and KO Mutant Expressing Single Lhcb4 Isoform.

ACKNOWLEDGMENTS

We thank Giovanni Finazzi (Commissariat à l’Energie Atomique, Grenoble, France) for helpful discussion. Financial support for this work was provided by the Programmi di Ricerca di Interesse Nazionale (2008XB774B) and by a grant of the Italian Ministry of Research and Marie Curie Actions Initial Training Networks HARVEST (Grant 238017).

AUTHOR CONTRIBUTIONS

S.d.B. isolated single KO mutants, carried out the crossings to obtain all the genotypes used, and performed a biochemical and physiological characterization of their photosynthetic apparatus. S.C., N.B., and L.D. were involved in the photoxidative treatments and stress measurements, in the spectroscopy investigation on thylakoids or isolated complexes, and in data analysis. R.K., E.B., and N.B. carried out electron microscopy structural analysis of isolated grana and PSII particles. R.B. and L.D. conceived the study, participated in its design and coordination, and helped to draft the manuscript.

Received May 11, 2011; revised June 21, 2011; accepted July 17, 2011; published July 29, 2011.

REFERENCES


