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**RESEARCH COMMUNICATION**

**Ink4a and Arf differentially affect cell proliferation and neural stem cell self-renewal in Bmi1-deficient mice**

Sophia W.M. Bruggeman,1,4 Merel E. Valk-Lingbeek,1,4 Petra P.M. van der Stoop,1,5 Jacqueline J.L. Jacobs,1,5 Karin Kieboom,1 Ellen Tanger,1 Danielle Hulsman,1 Carly Leung,2 Yvan Arsenijevic,3 Silvia Marino,2 and Maarten van Lohuizen1,6

1Division of Molecular Genetics, The Netherlands Cancer Institute, 1066 CX Amsterdam, The Netherlands; 2Department of Pathology, Institute of Clinical Pathology, University of Zürich, 8091 Zürich, Switzerland; 3Laboratory of Oculogenetics, Ophthalmic Hospital Jules Gonin, Lausanne University, 1004 Lausanne, Switzerland.

The Polycomb group (PcG) gene *Bmi1* promotes cell proliferation and stem cell self-renewal by repressing the *Ink4a/Arf* locus. We used a genetic approach to investigate whether *Ink4a* or *Arf* is more critical for relaying *Bmi1* function in lymphoid cells, neural progenitors, and neural stem cells. We show that *Arf* is a general target of *Bmi1*, however particularly in neural stem cells, derepression of *Ink4a* contributes to *Bmi1*−/− phenotypes. Additionally, we demonstrate haploinsufficient effects for the *Ink4a/Arf* locus downstream of *Bmi1* in vivo. This suggests differential, cell type-specific roles for *Ink4a* versus *Arf* in PcG-mediated (stem) cell cycle control.

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The *Bmi1* gene, originally identified as a collaborating oncogene in c-Myc-induced lymphomagenesis, is a member of the Polycomb group (PcG) gene family of chromatin modifiers and transcriptional repressors [Haupt et al. 1991; Van Lohuizen et al. 1991]. Loss of a PcG gene alters *Homeobox* (Hox) gene expression resulting in skeletal malformations [Van der Lugt et al. 1994; Akasaka et al. 1996; Core et al. 1997; Del Mar Lorente et al. 2000]. However, *Hox* genes are not the only targets of PcG as the tumor suppressor locus *Cdkn2a* (hereafter *Ink4a/Arf* locus) is also negatively regulated by *Bmi1* and other PcG members [Jacobs et al. 1999; Voncken et al. 2003; Core et al. 2004; Gil et al. 2004].

**Results and Discussion**

**Differential effects of Ink4a and Arf derepression in Bmi1-deficient lymphoid organs**

*Bmi1−/−* mice suffer from progressive hypoplasia of the spleen and thymus as illustrated by a large reduction in lymphocyte counts [Van der Lugt et al. 1994; Fig. 1]. We have previously shown that *Ink4a/Arf* is an in vivo target of *Bmi1* in the lymphoid system [Jacobs et al. 1999]. Here we investigated the impact of *Ink4a* versus *Arf* derepression on lymphocyte counts in *Bmi1−/−* mice. In the thymus but not in spleen, *Arf* loss gives a significantly smaller rescue than *Ink4a/Arf* loss, revealing a role for *Ink4a* restricting thymocyte growth downstream of *Bmi1* (*p < 0.01*) [Fig. 1A,C]. However, loss of *Ink4a* alone does not alleviate the phenotype showing that reduction of *Arf* levels is a prerequisite [Fig. 1B,D]. Lack of rescue from *Bmi1* deficiency upon *Ink4a* deletion is not unique for splenocytes. Also *Bmi1−/−* mouse embryonic fibroblasts (MEFs), which up-regulate p16*ink4a* and p19*arf* in a dose-dependent manner [Supplementary Fig. 1A], are...
only rescued from premature senescence upon loss of Arf (Supplementary Fig. 1B,C). Intriguingly, in the Bmi1-deficient spleen but not in thymus, heterozygosity for either Arf or Ink4a/Arf alleviates the reduced cell counts to a minor yet significant extent (p < 0.001), indicating that specific threshold levels of Ink4a and Arf are required (Fig. 1A,C).

Similar effects occur in the composition of the splenic B-cell population. Bmi1−/− mice harbor increased thymic populations of CD4−CD8− immature T-cells and a decreased number of splenic mature B220−Slg− B-cells (Van der Lugt et al. 1994; Jacobs et al. 1999). Applying flow cytometry using standard T- and B-cell differentiation markers, we found that in spleen, heterozygosity for either Arf or Ink4a/Arf completely restores the relative population frequency of mature B-cells to wild-type levels [Supplementary Fig. 2A,B]. In contrast in thymus, only complete loss of Arf or Ink4a/Arf results in a complete rescue (p < 0.05). Note that heterozygosity does not lead to a rescue. p53 loss induces a minor rescue in lymphocyte counts.

### Figure 1. Differential effects of Ink4a and Arf dosage and derepression in Bmi1−/− lymphoid organs.

(A) Splenocyte counts are dramatically reduced in Bmi1−/− mice. Heterozygosity for Arf or Ink4a/Arf partially rescues this phenotype (p < 0.01) but complete loss of Arf or Ink4a/Arf results in a better partial rescue. p53 loss also induces a partial rescue. (B, D) Neither Bmi1−/− splenocyte nor thymocyte counts are significantly restored by loss of Ink4a. (C) Thymocyte counts are dramatically reduced in Bmi1−/− mice. Loss of the complete Ink4a/Arf locus gives a substantially better rescue than loss of Arf alone (p < 0.05). Note that heterozygosity does not lead to a rescue. p53 loss induces a minor rescue in lymphocyte counts.

### Figure 2. Ink4a/Arf derepression upon Bmi1 loss prevents efficient Shh-induced CGNP proliferation.

When investigating the neurological defects of Bmi1−/− mice, we recently pinpointed a reduced number of poorly proliferating cells of the cerebellar external granular layer [EGL] to be partially responsible (Leung et al. 2004). During normal early post-natal development, a wave of proliferation of cerebellar granule neuron progenitors (CGNPs) in the EGL is induced by Purkinje neuron secreted Sonic Hedgehog (Shh) [Dahmane and Ruiz-I-Altaba 1999, Wechsler-Reya and Scott 1999]. In time, CGNPs differentiate and migrate inward past the molecular layer (ML) until they reach the internal granular layer (IGL) where they reside as mature granule neurons (for review, see Wang and Zoghbi 2001). We were able to link this process to PCG by showing that Bmi1 is a target of Shh, thus explaining the impaired proliferative response of Bmi1−/− CGNPs upon Shh stimulation (Leung et al. 2004). As Bmi1−/− CGNPs are still partially responsive to Shh, we propose a model where Shh controls proliferation via at least two routes, i.e., through induction of N-Myc/Cyclin D2 (Kenney et al. 2003) and through Bmi1-mediated repression of Ink4a/Arf.

Analysis by quantitative real-time PCR (qRT-PCR) shows that Ink4a and to a lesser extent Arf expression is increased in Bmi1−/− CGNPs and cerebella (Fig. 2A). Moreover, we demonstrate that stimulation of wild-type CGNPs with Shh leads to an induction of Bmi1 protein expression and a concomitant significant down-regulation of both Ink4a and Arf transcript levels, consistent with the idea of Shh regulating Ink4a/Arf through Bmi1 (Fig. 2C,D). To assess to what extent Ink4a and Arf derepression underlies the proliferative defects of Bmi1−/−...
CGNPs, we investigated the Shh response of Bmi1;Arf and Bmi1;Ink4a/Arf doubly deficient CGNPs and their respective controls. Notably, neither Arf nor Ink4a/Arf loss causes enhanced proliferation in absence of Shh in line with the multiple levels of regulation downstream of Shh [Fig. 2B,E]. However, in the context of Bmi1 deficiency, both Arf loss alone and complete loss of the Ink4a/Arf locus rescues Shh-induced proliferation of Bmi1<sup>−/−</sup> CGNPs to control levels (p < 0.05) [Fig. 2B,E]. Notably, we observe a clear effect of heterozygosity for Ink4a/Arf in Bmi1<sup>−/−</sup> CGNPs similarly to thymus [Fig. 2E].

We demonstrated earlier that loss of Ink4a/Arf gives a qualitative rescue of the Bmi1<sup>−/−</sup> cerebellar defects in vivo [Jacobs et al. 1999]. Since we found that Arf loss fully rescued Bmi1<sup>−/−</sup> CGNPs in vitro, we questioned what extent this affects in vivo proliferation of cerebellar progenitor cells. Histological analysis revealed that the reduced thickness of the Bmi1<sup>−/−</sup> granular layer is rescued to a similar extent in either an Arf<sup>−/−</sup> or Ink4a/Arf<sup>−/−</sup> background [Fig. 3A,B]. Surprisingly, the cell density of the Bmi1<sup>−/−</sup> IGL is fully rescued in an Ink4a/Arf<sup>−/−</sup> background, but only partially in an Arf<sup>−/−</sup> background (p < 0.01) pointing at a subtle role for Ink4a-repression in the IGL in vivo [Fig. 3A,C]. Disturbed EGL proliferation can explain multiple abnormalities of the cerebellum. Particularly later during development, granule neurons signal to Purkinje and Basket cells to create clusters of stem cells and progeny (Reynolds and Weiss 1992). One major neurogenic region in the adult cerebrum harboring stem cells is the subventricular zone (SVZ) of the lateral ventricle wall [for review, see Doetsch 2003]. Neural stem cells isolated from the SVZ can be grown as either adherent colonies or as “neurospheres”, floating clusters of stem cells and progeny (Reynolds and Weiss 1992; Morshede et al. 1994).

Recently, Molofsky et al. demonstrated that Bmi1 is essential for the self-renewal of SVZ-derived neurospheres and that Ink4a repression is partially mediating this effect [Molofsky et al. 2003]. Here we set out to determine whether Arf is required as well. Western blot analysis on wild-type and Bmi1 knockout cerebral tissue (isolated from post-natal day 50 [P50] mice) revealed

**Figure 3.** Ink4a and Arf differentially contribute to histological abnormalities of the Bmi1<sup>−/−</sup> cerebellum. (A) Morphological analysis of wild-type (left panels), Bmi1<sup>−/−</sup> (middle left panels), Bmi1<sup>−/−</sup>;Arf<sup>−/−</sup> (middle right panels), and Bmi1<sup>−/−</sup>;Ink4a/Arf<sup>−/−</sup> (right panels) adult cerebellum. Haematoxylin and eosin (H&E, top and middle panels) staining shows rescue of overall cerebellar size and granular layer thickness in Bmi1<sup>−/−</sup>;Ink4a/Arf<sup>−/−</sup> mice. (Bottom panels) Aberrant arborization of basket neurons (NF200 staining) is observed in all cerebella lacking Bmi1. Final magnification, 5x and 60x. (IGL) Internal granular layer; (ML) molecular layer. (B) IGL area measurements reveal significant rescues in Bmi1<sup>−/−</sup>;Arf<sup>−/−</sup> and Bmi1<sup>−/−</sup>;Ink4a/Arf<sup>−/−</sup> mice. (C) Ink4a/Arf loss induces a complete rescue of the number of Bmi1<sup>−/−</sup> granule neurons, whereas Arf loss leads to a partial rescue (p < 0.01). (D) The partial rescue in number of Bmi1<sup>−/−</sup> molecular layer neurons is significantly better in an Ink4a/Arf deficient background (p < 0.05) [HPF, high-power field; n = 4 mice].
up-regulated p16ink4a expression in the Bmi1−/− brain (Fig. 4B). In addition, qRT-PCR demonstrated that alike in MEFs and cerebellum, Ink4a and Arf transcripts are up-regulated, establishing Ink4a/Arf as a bona fide in vivo Bmi1 target (Fig. 4C). Next, we prepared adult SVZ neural stem cell cultures at clonal density and assured that the vast majority of primary neurospheres was derived from multipotent stem cells by staining for neuronal and glial markers (Fig. 4A, right panel). Strikingly, Bmi1−/− primary neurospheres not only form less frequently, but also differentiate less efficiently (Fig. 4A right panel) and are much smaller in size than control neurospheres (Fig. 4A [left panel], D). The latter observation suggests that proliferation within a Bmi1−/− neurosphere, reflecting the sum of cell divisions from self-renewing stem cells and their progeny, is impaired. Indeed, Bmi1−/− colonies incorporate far less BrdU than control colonies (Fig. 4F). Notably, this defect in proliferation can be rescued by both loss of Arf and by complete deletion of the Ink4a/Arf locus implicating Arf as an important downstream effector of Bmi1 for stem cell and progenitor proliferation (Fig. 4A,F).

Next, we performed a neurosphere assay to specifically study stem cell self-renewal. In this assay, the capacity of a primary neurosphere to form new multipotent neurospheres after dissociation is measured. We report a dramatic decrease in the self-renewing capacity of both adult (P30) and P7-derived Bmi1−/− neurospheres in agreement with previous findings (Fig. 4E). Deletion of the Ink4a/Arf locus in Bmi1−/− neurospheres completely rescues this defect. Importantly, loss of Arf alone gives a partial rescue (p < 0.05), reinforcing the earlier observation that proper repression of Ink4a is also required for neurosphere self-renewal. This does not simply reflect a tissue culture phenomenon as is highlighted by Molosky et al. (2005) who found that Ink4a loss partially restores Bmi1−/− stem cell frequency and neurogenesis in vivo. Upon induction of differentiation, all secondary neurospheres stained positive for neuron and astrocyte specific markers (data not shown). Surprisingly, there is a clear increase in both proliferation and self-renewing capacity of Arf and Ink4a/Arf deficient neurospheres compared with wild types (Fig. 4E,F). This strongly suggests that the Ink4a/Arf locus actively restricts self-renewing cell divisions thus playing a role in controlling the stem cell compartment. Lastly, we tested these neurospheres for long-term self-renewal as, for instance, exhaustion of hematopoietic stem cells sometimes occurs after a prolonged period of time (Park et al. 2003). However, we were able to keep Bmi1−/−:Arf−/− and Bmi1−/−:Ink4a/Arf−/− neurospheres in culture for at least five weekly passages, suggesting that the Bmi1 knockout phenotype is fully reversed.

A specific subset of PcG proteins appears to regulate stem cell self-renewal and progenitor proliferation

How could PcG distinctly regulate its target genes in different cell types? Emerging studies indicate that different “flavors” of PcG complexes may exist (De Napoles et al. 2004; Kuzmichev et al. 2004). An increase in the relative amount of a PcG member such as Bmi1, may alter the affinity of a PcG complex towards the chromatin in such a way that for instance self-renewing divisions are favored (for review, see Valk-Lingbeek et al. 2004). Underscoring a special role for Bmi1, we found

![Figure 4.](image-url)
that loss of Ring1a or M33, PcG proteins belonging to the same complex as Bmi1, does not affect neurosphere self-renewal or CGNP proliferation (Supplementary Fig. 3A,B). Only loss of Mel18, the closest homolog of Bmi1, has a modest though not significant negative effect on these processes in accordance to our observations in hematopoietic stem cells [Iwama et al. 2004]. It remains striking that lymphoid cell counts, cerebellar development, skeletal transformations, and overall body growth in Bmi1;Ink4a/Arf doubly deficient mice are at best partially rescued [Figs. 1, 3; Supplementary Table 1; Supplementary Fig. 4]. Therefore, other Bmi1 regulated genes must exist. In line with Drosophila not possessing genes resembling the Ink4a/Arf locus, the acquisition of PcG-mediated proliferation control likely evolved later, perhaps reflecting a demand for protection of long lasting stem cells. Candidates for additional Bmi1 targets are the Hox genes. Interestingly, a subset of Hox genes has been implicated in mammalian brain development and control of hematopoiesis. Moreover, several Hox genes are differentially expressed in Bmi1−/− SVZ neurospheres [Molofsky et al. 2003]. However, it cannot be excluded that a substantial number of additional targets exists, as PcG proteins as chromatin modifiers are known to act on large gene sets [Kirmizis et al. 2004]. Altogether, we have demonstrated profound tissue and cell type specific differences in the effects of Ink4a versus Arf derepression in Bmi1-deficient mice. We propose a general role for Bmi1-mediated Arf/p53 repression in curtailing proliferation. Ink4a repression on the other hand is required only in a subset of cells. Furthermore, we show haploinsufficient effects of the Ink4a/Arf locus in vivo selectively for certain tissues, and suggest that appropriate threshold levels of these two proteins are required to ensure proper proliferation. These observations implicate PcG proteins not only in embryonic developmental fate decisions, but also in discriminative processes between cell cycle control of stem- and more differentiated cells.

Materials and methods

Mice breeding, lymphocyte counts and flow cytometry

Bmi1−/+ FVB or C57BL/6 mice [Van der Lugt et al. 1994] were crossed with Ink4a/Arf−/+ FVB mice [Serrano et al. 1996], with Arf−/+ FVB mice [Kamijo et al. 1997], with p53−/+ FVB mice [Donehower et al. 1992], or Ink4a−/− C57BL/6 mice [Krimpenfort et al. 2001]. In addition, Ring1a FVB mice [Del Mar Lorente et al. 2000], M33 FVB mice [Core et al. 1997] and Mel18 [mixed background] knockout mice [Akasaka et al. 1996] were used. All mice were genotyped routinely by PCR (list of primers available upon request). Multiple independent animals of the respective genotypes were assayed and all results were subjected to Student’s t-tests and Bonferroni correction when appropriate. Preparation of cell suspensions from lymphoid organs, cell counts, and flow cytometric analyses were done as described previously [Jacobs et al. 1999].

Western blot analysis and quantitative real-time PCR

Equal amounts of protein were separated on 13% SDS-PAGE or precast gels [Invitrogen] and blotted onto Immobilon-P membranes [Amersham Biosciences]. Bands were visualized using enhanced chemiluminescence [Amersham]. Primary antibodies were F6 for Bmi1, Mel18 for p16Ink4a [Santa Cruz], or R562 for p19Arf [Abcam]. Secondary antibodies were goat-anti-mouse [ZyMed] or goat-anti-rabbit [BioSource], both HRP conjugated. Total RNA was extracted using TRIZOL reagent [Invitrogen] and cDNA was prepared using Superscript II RT and oligo(T)18 primers [Invitrogen]. qRT-PCR was performed with 50 ng cDNA on an ABI PRISM 7000 using SYBR Green PCR mastermix (Applied Biosystems). For primer sequences and formulas, see Supplemental Material.

CGNP isolation and histology of cerebellar tissue

CGNP cultures were isolated from 7 d-old mice and cultured according to established protocols [Weehler-Reya and Scott 1999; Leung et al. 2004]. Quantification of Shh-induced proliferation was performed as described previously [Leung et al. 2004]. Immunocytochemistry was performed using mouse monoclonal anti-BrdU (DAKO) and Alexa Fluor m488 goat-anti-mouse [Molecular Probes]. DAPI [Molecular Probes] was used to visualize nuclei. Immunohistochemistry and histological analyses were performed as described before [Leung et al. 2004].

Neural stem cell isolation, neurosphere proliferation, and self-renewal assays

For neural stem cell isolation and culture conditions, see Supplemental Material. To assess proliferation, primary adherent stem cell colonies were pulsed for 1 h with 1 µM BrdU [7 d after isolation]. Immunocytochemistry was performed as for CGNPs. The self-renewal capacity was determined by dissociating 10-d-old primary neurospheres and replating them in six-well ultra-low binding plates at clonal density. After 10 d, the newly generated (secondary) neurospheres were counted. To reveal multipotency, primary and secondary neurospheres were plated onto polyornithine and fibronectin- or laminin-coated [Sigma] plates and differentiated for 4 d in neurosphere medium supplemented with 2% FBS [ICN Biomedicals]. Cells were labeled with antibodies against GFAP [DAKO] and β-tubulin-III [Sigma]. Secondary antibodies were FITC-conjugated goat-anti-rabbit and Cy3-conjugated goat-anti-mouse [Jackson Immunoresearch]. Nuclei were stained with DAPI.

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