Rapid and efficient cancer cell killing mediated by high-affinity death receptor homotrimerizing TRAIL variants

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The tumour necrosis factor family member TNF-related apoptosis-inducing ligand (TRAIL) selectively induces apoptosis in a variety of cancer cells through the activation of death receptors 4 (DR4) and 5 (DR5) and is considered a promising anticancer therapeutic agent. As apoptosis seems to occur primarily via only one of the two death receptors in many cancer cells, the introduction of DR selectivity is thought to create more potent TRAIL agonists with superior therapeutic properties. By use of a computer-aided structure-based design followed by rational combination of mutations, we obtained variants that signal exclusively via DR4. Besides an enhanced selectivity, these TRAIL-DR4 agonists show superior affinity to DR4, and a high apoptosis-inducing activity against several TRAIL-sensitive and -resistant cancer cell lines in vitro. Intriguingly, combined treatment of the DR4-selective variant and a DR5-selective TRAIL variant in cancer cell lines signalling by both death receptors leads to a significant increase in activity when compared with wild-type rhTRAIL or each single rhTRAIL variant. Our results suggest that TRAIL induced apoptosis via high-affinity and rapid-selective homotrimerization of each DR represent an important step towards an efficient cancer treatment.

Subject Category: Experimental Medicine

Tumour necrosis factor (TNF)-related apoptosis-inducing ligand (TRAIL) is a promiscuous member of the TNF superfamily capable of binding to five different receptors. TRAIL induces apoptosis via the death receptors (DRs) 4 and 5,1–2 followed by recruitment of the Fas-associate death domain.3–5 Formation of the death-inducing signalling complex (DISC) is completed by binding of procaspase-8 and -10, leading to their cleavage and activation, that in turn activate downstream caspases (e.g. caspase-3), resulting in apoptosis.6–8 TRAIL binds also to three decoy receptors, DcR1, DcR2 and OPG.9 Decoy receptors lack a functional death domain, and consequently they do not induce apoptosis and can prevent TRAIL binding to death receptors.2,3–11

TRAIL is generating considerable interest as a possible anticancer therapeutic agent because of its selective activation of apoptosis in cancer cells.12,13 The formation of heterotrimeric non-apoptotic TRAIL–receptor complexes7,14,15 and the receptor binding promiscuity of TRAIL endorsed the concept that DR selectivity may allow the creation of more potent TRAIL inducers.16–19 DR5 was described as contributing more than DR4 to the overall apoptotic activity of TRAIL in apoptosis signalling of cancer cells.16,20,21 Recently, DR4 has gained increased importance, with the realization that certain cancer cells depend mainly on this receptor for apoptosis via TRAIL17,18,22 Whereas potent DR5-selective variants were created previously,16,19 no enhancement of apoptosis by selective activation of DR4 has been accomplished by using TRAIL variants. A previous generation of DR4-selective variants designed by us18 or by others17 were either equally active when compared with rhTRAILWT or largely inactive.16–17

Here we report the design of highly active, DR4-specific TRAIL variants by using computational protein design and a rational combination strategy. This resulted in the first DR4-selective variants that not only have an enhanced selectivity to DR4, but also a significant increase in apoptosis-inducing activity in many cancer cells in vitro. Both receptor selectivity and increased affinity for DR4 appear to contribute to the more efficient apoptosis induction by these TRAIL variants.

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Abbreviations: TRAIL, tumour necrosis factor-related apoptosis-inducing ligand; DRs, death receptors; DR4, death receptor 4; DR5, death receptor 5; SPR, surface plasmon resonance

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These DR4-selective variants and the previously described DR5-selective variants allowed us to establish the contribution of DRs in apoptosis signalling of different cancer cells. Cell lines signalling via both DRs showed a large increase in efficacy upon combined treatment with selective TRAIL variants when compared with rhTRAILWT. Our results show these novel DR4-specific variants alone or in combination with a DR5-specific variant as a powerful strategy for targeting DR4/DR5-responsive cancer cells.

Results

Computational design of variants using FoldX. A refined model of the DR4 receptor was created using FoldX. All residues in the TRAIL–receptor interface were considered. A set of predicted energetic values for TRAIL–receptor complex formation was obtained and compared with the TRAILWT values. FoldX proposed the following mutations to favour DR4 selectivity: R149I, R149M, R149N, R149K, S159R, Q193H, Q193K, N199R, K201R, K204D, K204E, K204L, K204Y, K212R, S215D, S215E, S215H, S215K, K251D, K251E and K251Q. Mutations at position 149, 159, 199, 201 and 215 showed increased selectivity to DR4 as predicted by FoldX. Mutations at positions 148, 193, 204, 212 and 251 did not show the expected changes. Interestingly, positions 199, 201 and 215 have been found before in DR4 variants selected by phage display. Variants showing an increase in binding ratio of DR4 over DR5 (DR4/DR5) of more than fourfold as measured by pre-steady state SPR (Supplementary Table 1), and/or presenting a more than threefold increased activity on DR4-mediated cell lines, and showing inactivity in DR5-mediated cell lines were chosen for further studies. R149I, S159R and S215D were selected.

Rational combination of single variants. Several amino-acid substitutions resulted in high DR4/DR5 binding ratios relative to rhTRAIL WT, thus favouring DR4 binding. The characteristics of these single mutants were used for rational design of nine combination variants, containing between two and six mutations each. A single mutant (G131R) constructed previously showed increased affinity to both DRs and a double mutant (N199R; K201H), showing a decrease in affinity to DR5 have been combined before (to be published elsewhere), and were therefore incorporated in the new combination variants. Mutation S159R was incorporated in all the combination variants, showing a high affinity ratio change (Supplementary Table 1). Mutants displaying similar or higher binding affinities to DR4, together with a lowered affinity for DR5, were characterized. Mutations that resulted in more than 50-fold increased DR4/DR5 binding ratio compared with rhTRAILWT as measured by SPR were selected. Two combination variants showing the highest affinity increase to DR4 and affinity decrease for DR5 were selected for further studies. These variants were named 4C7 (G131R/R149I/S159R/N199R/K201H/S215D) and 4C9 (G131R/R149I/S159R/S215D).

Death receptor selectivity of 4C7 and 4C9. Binding of rhTRAILWT and variants 4C7 and 4C9 to DRs and using SPR showed an increased affinity to DR4-Ig with a 9–10-fold lower dissociation constant (Kd) when compared with rhTRAILWT (Table 1). In contrast, 4C7 and 4C9 showed a much lower affinity to DR5-Ig with regard to rhTRAILWT with a 40- and 230-fold increase in dissociation constant to DR5-Ig for 4C9 and 4C7, respectively (Table 1). DR4-Ig competed with rhTRAILWT with a 5–7-fold increased effect for the variants 4C7 and 4C9 when compared with rhTRAILWT (Figure 1a). Interestingly, DR5-Ig was less capable to compete for binding to 4C7 and 4C9 binding to immobilized DR4-Ig (Figure 1b). A 29-fold increased binding of 4C9 to immobilized DR4-Ig was observed compared with rhTRAILWT when competing with DR5-Ig, indicating a preference to bind to DR4. 4C7 was even less sensitive to competition by DR5-Ig, with 80% binding efficiency for DR4 even when competing with 5000 ng/ml of DR5-Ig. These results indicate that the variants 4C7 and 4C9 bind highly preferentially to DR4.

Decoy receptors DcR1-Ig and DcR2-Ig were found to equally compete for binding to rhTRAILWT 4C7 and 4C9 (Supplementary Figure S2). Interestingly, another combination variant (4C6: R149I/S159R/S215D) showed not only lowered binding to DR5, but also decreased binding affinity to both decoy receptors in solution (Supplementary Figure S2).

<table>
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<th>Protein</th>
<th>kₐ (M⁻¹s⁻¹)</th>
<th>kₐ (s⁻¹)</th>
<th>K₀ (nM)</th>
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<td>(1.66 ± 0.3)×10⁻⁴</td>
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<tr>
<td>WT</td>
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<td>0.033 ± 0.03</td>
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<tr>
<td>4C9</td>
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<td>(23.1 ± 1.2)×10⁻⁴</td>
<td>1.22 ± 0.2²</td>
</tr>
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</table>

*Association rate constant. †Dissociation rate constant. ‡Equilibrium dissociation constant (kₐ/kₐ). Each value was calculated using the BIAcore 3000 and the BIA evaluation 4.1 software. ‡Curves showed heterogeneity, a phenomenon that may be correlated to partial glycosylation of receptors.
different cell lines mediating apoptosis via different DRs. Analysis of cell death induction in Burkitt’s lymphoma BJAB cells inducible by both DR4 and DR5 (BJABWT) and BJAB cells deficient in DR5 (BJABDR5 DEF) showed that 4C7 and 4C9 do not require DR5 to deliver similar levels of apoptosis in both cell lines. In addition, 4C7 and 4C9 showed a significant increase in cell death induction in BJAB cells when compared with rhTRAILWT in all concentrations measured (Figure 2a and b).

Cell assays in DR5-mediating A2780 and Jurkat cells showed no apoptosis-inducing activity of the variants (<10%) at almost all concentrations measured (Figure 2c and d). These results support that 4C7 and 4C9 induce apoptosis essentially via DR4. Furthermore, enhanced activity of TRAIL variants 4C7 and 4C9 was shown on a number of colon adenocarcinoma cells sensitive to TRAIL-induced apoptosis when compared with rhTRAILWT, including Colo205, SW948, HCT-15 and DLD-1 (Figure 3a–d). A reduction of 3–5-fold in cell viability of Colo205 by variants 4C7 and 4C9 when compared with rhTRAILWT was observed (Figure 3a). The SW948 cell line showed a strong decrease in cell viability upon treatment with the variants (Figure 3b). Moreover, an impressive decrease in cell viability has been obtained in HCT-15 and DLD-1 cell lines by 4C7 and 4C9 when compared with rhTRAILWT (up to 88% cell death) (Figure 3c and d). Our results indicate that these cell lines can

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**Figure 1** Competitive inhibition by TRAIL death receptors for rhTRAILWT and variants 4C7 and 4C9, binding to immobilized DR4-Ig, using soluble DR4-Ig as a competitor (a) and soluble DR5-Ig as a competitor (b), in the presence of 100 ng/ml of rhTRAILWT or variants and 0–5000 ng/ml per well of DR4- or DR5-Ig. Binding of rhTRAILWT and variants was measured relative to the wells containing no soluble receptor.

**Figure 2** Death receptor 4 selectivity of variants in several cell lines. Cell death induction by wild-type TRAIL and variants on (a) BJAB cell line mediating apoptosis via both DR4 and DR5 receptors (BJABWT), (b) BJAB DR5-deficient cell line (BJABDR5 DEF) mediating apoptosis via DR4, (c) DR5-mediated Jurkat leukaemia and (d) DR5-mediated A2780 ovarian carcinoma cells. BJAB cells were treated with rhTRAILWT and variants for 24 h. The percentage of cell death was calculated relative to control wells containing no ligand. Jurkat leukaemia cells and A2780 ovarian carcinoma cancer cells were treated with the indicated concentrations of rhTRAILWT and variants for 24 h, after which induction of cell death was measured by Annexin V staining. The results are mean values ± S.E.M. (n = 3).
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be triggered very efficiently to undergo apoptosis by these novel DR4-specific variants.

We then tested whether selective activation of DR4 would have an impact on cancer cell lines less sensitive to TRAIL-induced apoptosis. Remarkably, our variants 4C7 and 4C9 showed a significant increase in cell killing in breast cancer MCF-7 and pancreatic cancer PANC-1 cell lines (Figure 3e and f). Both cell lines express DR4 and DR5 on their surface (Supplementary Figure S4). Furthermore, the variants were tested for activity on normal cells, namely fibroblasts. No significant activity for rhTRAILWT or variants 4C7 and 4C9 could be observed, suggesting that the increased apoptotic activity on cancer cells had no effect on normal cells (Supplementary Figure S5).

Real-time analysis of caspase activation. The variant 4C7 was subject of study by analysing caspase activation using FRET. Activation of the initiator caspase-8/10 was followed by using a probe based on CFP and YFP (Venus), interconnected by a linker containing the preferred cleavage motif for caspase-8 (IETD). Owing to its high signal/noise ratio, this probe allows the efficient detection of low caspase activities. The 4C7 variant-induced profile of intracellular IETDase activity shows increased cleavage of the CFP-IETD-Venus probe in ovarian carcinoma OVCAR-3 cells at much earlier time points when compared with rhTRAILWT (Figure 4). Cleavage by activated initiator caspases could be observed within 15–20 min after 4C7 addition. More significant differences between rhTRAILWT and 4C7 occur after 50 min of incubation with 50 and 100 ng/ml (Figure 4a and b). Effector caspases have also been shown to cleave IETD recognition sites, and consequently a more rapid increase observed after cleavage by activated caspase-8 is most likely owing to the activation of

Figure 3 Several cancer cell lines undergo apoptosis efficiently via TRAIL DR4-selective variants. (a) Colo205, (b) SW948, (c) DLD-1 and (d) HCT-15 colon carcinoma cell lines were incubated with different concentrations of rhTRAILWT and variants 4C7 and 4C9 for 24 h, and assayed for cell viability. The percentage of viable cells was calculated relative to control wells containing no ligand. The results are mean values ± S.E.M. (n = 3). Death receptor 4-selective variants induce higher levels of apoptosis in MCF-7 human breast adenocarcinoma (e) and PANC-1 human pancreatic carcinoma cells (f). Cells were treated with rhTRAILWT and variants 4C7 and 4C9 for 24 h and cell death was measured by Annexin V staining. The results are mean values ± S.E.M. (n = 3)
effector caspases during apoptosis or elevated caspase-8 activity, resulting from feedback via caspase-6. Analysis of downstream activation of effector caspase-3/7 based on DEVDase activity using a probe containing CFP and YFP indicates that the rate of cleavage remains low at initial time points, and raises rapidly after 75 min over a 30–40 min period for 4C7 (Figure 4c and d). The rate of DEVDase cleavage for rhTRAILWT was substantially lower. These results indicate a much faster and robust initiator and effector caspase activation by variant 4C7 compared with rhTRAILWT in living cells.

Combination of death receptor-selective variants. In order to establish receptor activity in different cell lines, we tested the new DR4-selective variants or our DR5-selective mutant (D269H/E195R).19 Most cell lines tested undergo apoptosis primarily via one of the DRs; however, some cells did show sensitivity via both DRs. We reasoned that selective activation of both DRs by TRAIL variants may lead to an additive effect in cell death induction and that targeting both DRs separately may be of therapeutic relevance in cancer cell lines. OVCAR-3 cells express both DRs, with almost no detectable levels of decoy receptors (Supplementary Figure S4). Both 4C7 ($ED_{50} = 6.61 \pm 1.3 \text{ ng/ml}$) and 4C9 ($ED_{50} = 7.86 \pm 1.53 \text{ ng/ml}$) caused a substantial decrease in viability when compared with rhTRAILWT ($ED_{50} = 46.8 \pm 1.1 \text{ ng/ml}$), with the DR4-selective variants reaching cell death levels of 88 and 83%, whereas rhTRAILWT was only capable of 50% cell death induction at the highest concentration measured (250 ng/ml) (Figure 5a). Similarly, activation of DR5 by D269H/E195R in this cell line resulted in increased apoptosis induction, with a nearly sevenfold increase in cell death compared with rhTRAILWT. Interestingly, the combination of equimolar concentrations of DR4 and DR5 TRAIL-selective variants resulted in an even lower cell survival at all concentrations measured, when compared with the single treatment with either only rhTRAILWT or selective variants. At the highest concentration used, the combination of selective variants was capable of killing almost 95% of cells upon 24 h incubation. In the Colo205 cell line, the cooperation between DR4- and DR5-selective induction was less pronounced, even though at high concentrations higher levels of cell death could be shown (up to 94%). At these concentrations, also a difference with D269H/E195R could be observed (Figure 5b). In the colon carcinoma CL-34 cell line, both 4C7- and DR5-specific variant showed higher activities than rhTRAILWT at almost all concentrations and when combined induced higher killing levels compared

Figure 4  TRAIL-induced IETDase and DEVDase profiles of rhTRAILWT and variant 4C7 in OVCAR-3 ovarian carcinoma cell line. Cells were treated with two different concentrations of rhTRAILWT and 4C7, (a) 50 ng/ml and (b) 100 ng/ml. IETDase activity profile displayed FRET cleavage and consequent increase in CFP/YFP emission ratio is plotted. (c) DEVDase activities displaying FRET cleavage upon addition of 100 ng/ml of rhTRAILWT and variant 4C7. The curves in (a)–(c) represent the analysis of 10 different OVCAR-3 cells, following the TRAIL treatment. Error bars indicate the standard deviations for three single curve profiles using a similar number of cells. (d) CFP/YFP ratio images of a group of OVCAR-3 cells expressing the DEVD FRET probe treated with 100 ng/ml of rhTRAILWT and 4C7 variant.
with single treatments, especially at high concentrations (Figure 5c).

**Clonogenic survival assays.** To determine the effect of these variants on long-term survival of cancer cells, we performed clonogenic assays (Figure 6). Both DR4- and DR5-selective variants significantly reduced clonogenic growth of OVCAR-3 cells when compared with rhTRAIL<sub>WT</sub> (Figure 6a and b). The combination of the receptor-specific variants results in an increased ability to kill OVCAR-3 cells.

**TRAIL-DISC immunoprecipitation.** Analysis of the components of the death receptor (DR) DISC formed upon treatment with flag-tag rhTRAIL<sub>WT</sub> and flag-tag rhTRAIL 4C9 indicate the ligation of DR4 and DR5 by flag-tag rhTRAIL<sub>WT</sub>. Only DR4 but not DR5 could be pulled down in the cells treated with flag-tag rhTRAIL 4C9 (Figure 6c), further showing the DR selectivity of these variants. In addition, the activation of procaspase-8 within the 4C9-mediated DISC was more pronounced than in the rhTRAIL<sub>WT</sub>-induced DISC, indicating a more efficient activation of pro-caspase-8 by the DR4-selective variant.

**Discussion**

TRAIL binds to five different receptors, of which only two are capable of inducing apoptosis, DR4 and DR5. It is generally accepted that decoy receptors can inhibit TRAIL-induced apoptosis because they can bind TRAIL, but lack a functional death domain to form the DISC.<sup>2,9,11</sup> However, the clarity of this decoy concept has been blurred with the observation that presence of decoy receptors not necessarily translates in protection against apoptosis.<sup>21,28</sup> In a recent study, DcR2 has been reported to heteromerize with DR5 to form inactive complexes.<sup>14–15</sup> Heteromerization of death receptors DR4 and DR5 may also lead to inactive complexes.<sup>7,15</sup> In a large number of cancer cell lines, differential contributions of the DRs have been reported,<sup>29</sup> but it is not yet clear whether they differ in their function in the apoptotic signalling. In light of these studies, the introduction of receptor selectivity can contribute to the elucidation of the function of each death receptor. Such variants are expected to have increased <i>in vivo</i> activity and are candidates for new anticancer therapies.

Computational design based on the crystal structure of the TRAIL–DR5 complex allowed the successful introduction of DR5 selectivity by two-point mutations.<sup>19</sup> The highly selective variant D269H/E195R was shown to be very efficient to induce apoptosis in an ovarian carcinoma xenograft mouse model.<sup>30</sup> For DR4, a highly refined homology model was made, using data from mutational analysis.<sup>18,19,23</sup> A set of 21 single mutation TRAIL variants predicted to show DR4-selective behaviour were designed, and experimentally characterized. Two of the positions within this set were also reported from a phage display selection study.<sup>16</sup> All the 21 mutants were characterized for their ability to bind receptors and to induce apoptosis. Several mutations showed lower affinity for DR5 and higher affinity for DR4. In most cases, these effects could be well explained upon visual inspection of the mutant models.

A representative example is given by the mutation S159R that has a large beneficial effect on the DR4/DR5 binding ratio (Supplementary Table 1). In the WT–TRAIL–DR5 complex, Arg-115 of DR5 forms a salt-bridge with Glu-155 and a hydrogen bond with His-161 of TRAIL (Figure 7b). The hydroxyl group of Ser-159 donates a hydrogen bond to the carboxyl side chain group of Glu-155. Upon mutation of
Ser-159 to Arg, the conformation of the side chain of Arg-115 of DR5 is slightly changed and the hydrogen bond interactions with His-161 are destroyed (Figure 7a).

The hydrogen bond interaction of Arg-115 with His-161 is lost and the salt-bridge with Glu-155 is somewhat weakened. This results in an overall loss of interaction energy for the S159R mutation ($\Delta \Delta G_{\text{interaction}} = +1 \text{kcal/mol}$). On the other hand, in the TRAIL–DR4 complex the equivalent residue of Arg-115 is Pro-115 and, as a consequence, no direct interactions are observed between Glu-155, Ser-159 and His-161 of TRAIL and DR4 receptor residues (Figure 7d). Mutation S159R allows Arg-159 to create a hydrogen bond with the backbone oxygen atom of Cys-113 of DR4 (Figure 7c). As no interactions are destroyed with respect to the WT situation and one new interaction is established, there is a net gain in interaction energy ($\Delta \Delta G_{\text{interaction}} = -1.4 \text{kcal/mol}$). The loss in interaction energy between TRAIL and DR5 on the one hand, and a gain in interaction energy between TRAIL and DR4 upon Ser to Arg mutation on the other hand, explain the observed increase in DR4 binding specificity.

As TRAIL was reported to have a lower affinity for DR4 than to DR5, superscript a single mutation seems not sufficient to produce a variant with high affinity and selectivity for DR4. Single mutations were therefore combined based on observed biochemical and biological properties. Two criteria were used for the selection: (1) at least fourfold increase in DR4/DR5 binding ratio based on pre-steady state SPR measurements and (2) at least threefold increased biological activity on DR4-sensitive cancer cells. Nine combinations of the selected mutations were made and most new variants showed affinity improvement to DR4 and largely reduced affinity to DR5. From these, two variants were chosen for further characterization based on observed biochemical and apoptosis-inducing characteristics: 4C7 (carrying mutations G131R/N199R/K201H/R149I/S159R/S215D) and 4C9 (carrying G131R/R149I/S159R/S215D). Characterization of the affinities of these two proteins for DR4 and DR5 showed that they have not only a decreased affinity for DR5, but also an increased affinity for DR4 (Table 1). The selective behaviour of these variants was also shown by immunoprecipitation experiments using Colo205 cells, where only DR4 but not DR5 could be detected upon treatment with a flag-tagged version of 4C9 (Figure 6c).

Variants 4C7 and 4C9 showed a remarkable capacity to induce apoptosis in several cancer cells in vitro, including a panel of human colon adenocarcinoma (Colo205, SW948, DLD-1, HCT-15 and CL-34), Burkitt’s lymphoma...
(BJAB), pancreatic carcinoma (PANC-1), the human ovarian carcinoma OVCAR-3 and human breast adenocarcinoma MCF-7 cancer cell line. The colon carcinoma cell lines showed values ranging from three- to 10-fold increased cell death induction by 4C7 and 4C9 when compared with rhTRAILWT (Figure 3). The lack of activity on DR5-mediated A2780 and Jurkat cells and the comparison of activity between BJAB and BJAB DR5-deficient cell line with rhTRAILWT clearly confirmed the DR4 selectivity of the variants 4C7 and 4C9 (Figure 2). The increased activity of the variants was also evident in TRAIL-resistant MCF-7 and PANC-1 cells (Figure 3). The higher affinity and selectivity of our variants for DR4 (Table 1 and Figure 1) seems to correlate to their higher cellular activity in cell lines sensitive via the DR4 receptor.

To further analyse the activity of the variant 4C7, we determined caspase activation in OVCAR-3 cells using FRET and fluorescent probes (Figure 4). A much faster and stronger onset of caspase activation could be observed for 4C7 in comparison to rhTRAILWT caspase-8/10 activation as seen by the IETDase probe cleavage that started already at very early time points with a nearly eightfold faster activation when compared with rhTRAILWT (Figure 4a and b). Downstream effector caspase activation based on DEVDase activity using a probe containing CFP and YFP indicated a significant cleavage rate starting at 75 min over a 30–40 min period for 4C7 (Figure 4c and d). DEVDase cleavage for rhTRAILWT was substantially slower in the time points taken. These results allow us to conclude that variant 4C7 induces a much faster and robust initiator and effector caspase activation than rhTRAILWT in the OVCAR-3 cell line, and underlines the efficacy of the DR4-specific variant.

The high efficacy of our new DR4-selective variants in addition to the earlier published DR5 variant D269H/E195R19 inspired us to test whether their combination would lead to even higher apoptotic activity. Combination treatment using the DR4-selective variants with the DR5-selective variant enhanced apoptosis-inducing activity when compared with the responses obtained for the single treatments (Figure 5). Analysis of response curves using different concentrations of DR-specific variants in combination indicates a significant additive effect on cell death induction on the ovarian carcinoma OVCAR-3 cell line. A similar effect was observed in colon carcinoma cell lines Colo205 and CL-34 at high concentrations of the two TRAIL variants. Even though part of the effect can be explained by the higher affinities and higher association rate constants of these variants for their cognate receptor, these results also point to the inability of these variants to form inactive heteromeric receptor complexes between their target DR and the other receptors, as was shown for rhTRAILWT. Furthermore, our DR4- and DR5-selective variants strongly inhibited clonogenicity of OVCAR-3 ovarian carcinoma cells (Figure 6a and b), showing the enhanced efficacy of these variants compared with rhTRAILWT in reducing long-term survival of cancer cells.

Figure 7  Structural impressions of the area around 159 for S159R and rhTRAILWT as determined by FoldX: (a and b) TRAIL-DR5 and (c and d) TRAIL-DR4. The subunits of TRAIL are depicted in lime and receptors in green. The template selected was 1D4V, the structure at 2.2 Å resolution and of monomeric human TRAIL in complex with the ectodomain of DR5 receptor. The homotrimer was generated using the protein quaternary structure server from the EBI (http://pqs.ebi.ac.uk), having the symmetry coordinates in the PDB file.

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In summary, the newly constructed DR4-specific variants lead to the formation of high-affinity homotrimeric TRAIL–DR4 complexes. These TRAIL variants alone or in combination with our DR5 agonist may prove to be a promising treatment for a range of different cancers.

Materials and Methods

Modelling and computational design of variants using FoldX. Currently, the crystal structure of TRAIL alone or in complex with DR5 is known. A homology model of DR4 was built based on the coordinates of TRAIL and DR5, from the template TRAIL in complex with the ectodomain of DR5, pdb: ID 1D4V.29 Using the high sequence identity between DR5 and DR4 (>50%), with no insertions or deletions between both death receptors. Binding effects of mutations situated in the TRAIL-receptor interface allowed the construction of a refined homology model for the TRAIL–DR4 complex using the protein design capabilities of FoldX.30,31 These coordinates were used to construct the homotrimeric TRAIL using the protein quaternary structure server from EBI (http://icps.ebi.ac.uk). Single mutants of TRAIL were predicted by FoldX. Interaction energies were calculated as the sum of the energies of the individual receptor and ligand subunits and subtracting them from the global energy of the complex. A detailed description of the protein design algorithm FoldX (version 3.0) is available elsewhere in http://foldx.org.es.

Construction, expression and purification of rhTRAIL variants. Mutants were constructed by PCR reaction using the megaprimer mutagenesis method. The polymerase used was Phusion polymerase supplied by NEB (Woburn, MA, USA) using the BamHI and NotI sites and transformed into Escherichia coli BL21 (DE3). Homotrimeric TRAIL proteins were expressed and purified as described before.16,23 Flag-tag rhTRAIL WT and flag-tag rhTRAIL C49 were introduced by the sequence encoding a flag-tag N-terminally to the rhTRAIL (aa 114–281) sequence in pET15b. Analytical gel filtration, dynamic light scattering and non-reducing gel electrophoresis confirmed that rhTRAIL WT and variants were stable trimeric molecules and did not form higher-order molecular weight aggregates.

Receptor binding by surface plasmon resonance and competitive ELISA. Binding experiments were performed using a surface plasmon resonance-based Biacore 3000 (GE Healthcare, Eindhoven, The Netherlands). Research grade CM4 sensor chips, plasmon resonance-based Biacore 3000 (GE Healthcare, Eindhoven, The Netherlands) on the sensor surface of a CM4 sensor chip was performed following a protocol. Plasmids pSCAT8 and pDEVD were a kind gift from Dr. Markus Rehm. With Fugene 6 (Roche, Almere, The Netherlands) according to the manufacturer’s protocol. Plasmids pSCAT8 and pDEVD were a kind gift from Dr. Markus Rehm. Ratio images were obtained using the Image J software and available custom plugins by calculating the ratio from the obtained CFP and YFP images corrected for background. Signals were normalized by subtracting the minimum value across all time points from each single time-course experiment.

Cell viability and apoptosis assays. To determine sensitivity to rhTRAIL and variants, cells were plated in 96-well plates, allowed to adhere for 24 h and then treated with rhTRAIL in various concentrations, ranging from 1 to 250 ng/ml. After 24 h incubation, the cells were subjected to an MTS viability assay (Promega, Leiden, The Netherlands) following the manufacturer’s protocol. Cell viability was determined after 1–2 h of incubation by measuring the absorption at 490 nm on a microplate reader (Thermo Labsystems). Apoptosis induction was measured by Annexin V staining and quantified by flow cytometric analysis as described before.

Live cell microscopy: caspase activation in living cells using FRET. Time lapse images were collected using a Leica SP2 AOBS CSLM microscope equipped with an environmental chamber at 37 °C and 5% CO2 and 10 or 20 objective magnification. Images were collected every minute. OVCAR-3 ovarian carcinoma cell line expressing pSCAT6 producing CFP-DEVD-Venus and pDEVD containing CFP-DEVD-YFP were derived by transfection with Turbofect 6 (Roche, Almere, The Netherlands) according to the manufacturer’s protocol. Plasmids pSCAT8 and pDEVD were a kind gift from Dr. Markus Rehm. Images were obtained using the Image J software and available custom plugins by calculating the ratio from the obtained CFP and YFP images corrected for background. Signals were normalized by subtracting the minimum value across all time points from each single time-course experiment.

Clonogenic survival assay. The clonogenic ability of ovarian carcinoma cells (OVCAR-3) was determined by plating the cells at the appropriate dilutions in DMEM medium, treated for 24 h with rhTRAIL WT and variants and the colonies were counted after 2 weeks. Briefly, 100 or 200 cells were plated out in 94 mm Petri dishes (Greiner) and, after washing with PBS, the cell pellet was weighted and the cells lysed with nine volumes of Extraction buffer A (Dynabeads

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Co-immunoprecipitation kit, Invitrogen) supplemented with 50 mM NaCl and incubated on ice for 15 min. The lysates were cleared by centrifugation and mixed with 1.5 mg of antibody-coupled Dynabeads. The lysates were rotated with the beads for 3 h at 4°C, after which the supernatant was separated from the beads, the beads washed and the DISC complex eluted by following the manufacturer’s instructions. The immunoprecipitate was denatured in Laemmli buffer before loading on gels and analysis of the DISC components.

Conflict of interest

Wim J Quax, Luis Serrano and Afshin Samali are founding members of TRISKELE Therapeutics.

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