Advanced glycation end products in patients with peripheral artery disease

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Chapter 8

Diverging effects of diabetes mellitus in patients with peripheral artery disease and abdominal aortic aneurysm and the role of advanced glycation end products: ARTERY study. Protocol for a multicenter cross-sectional study


Submitted
Chapter 8

Abstract

Introduction
Diabetes mellitus is a well-defined risk factor for peripheral artery disease (PAD), but protects against the development and growth of abdominal aortic aneurysm (AAA). Diabetes mellitus is associated with arterial stiffening and peripheral arterial media sclerosis. Advanced glycation end products (AGEs) are increased in diabetes mellitus and cardiovascular disease. AGEs are known to form cross-links between proteins and are associated with arterial stiffness. Whether AGEs contribute to the protective effects of diabetes mellitus in AAA is unknown. Therefore, the ARTERY study is designed to evaluate the role of AGEs in the diverging effects of diabetes mellitus on AAA and PAD.

Methods and analysis
This cross-sectional multicenter study will compare the amount, type and location of AGEs in the arterial wall in a total of 120 patients with AAA or PAD with and without diabetes mellitus (n=30 per subgroup). Also, local and systemic vascular parameters, including pulse wave velocity, will be measured to evaluate the association between arterial stiffness and AGEs. Finally, AGEs will be measured in serum, urine and assessed in skin with skin autofluorescence using the AGE Reader™.

Ethics and dissemination
This study is approved by the Medical Ethics committees of University Medical Center Groningen, Martini Hospital and Medisch Spectrum Twente, The Netherlands. Study results will be disseminated through peer-reviewed journals and scientific events.

Trial registration
Trialregister.nl NTR 5363.
Abstract

Introduction

Diabetes mellitus is a risk factor for various diseases, including retinopathy, renal insufficiency and atherosclerosis. Remarkably, diabetes mellitus is a risk factor for peripheral artery disease (PAD), but protects against development of abdominal aortic aneurysm (AAA). Evidence for an inverse association between diabetes mellitus and AAA was shown in a large case-control study with veterans (n=73,451) in which an AAA was detected in 1,031 subjects. This study demonstrated an odds ratio of 0.54 for the presence of diabetes mellitus and AAA. This inverse association was confirmed by a meta-analysis consisting of 17 large studies. In addition, AAA growth was shown to be decreased substantially in patients with diabetes mellitus (n=49) as compared to patients without diabetes mellitus (n=311) after 36 months of follow-up. The explanation for the diverging effects of diabetes mellitus on occlusive versus dilating arterial disease remains unclear.

Differences in effects of diabetes mellitus on aorta versus large peripheral arteries may contribute to these diverging effects. AAA is characterized by inflammation of the tunica media with matrix metalloproteinase and macrophage activation, causing proteolysis and smooth muscle cell apoptosis. In contrast, PAD results from inflammation of the tunica intima and increased endothelial permeability, macrophage infiltration, retention of cholesterol and recruitment of smooth muscle cells which form a fibrous cap. Nevertheless, AAA and PAD appears often simultaneously in patients, possibly due to their common risk factors. Diabetes-induced changes in arterial pulse wave propagation and reflection may be another factor contributing to the diverging effects of diabetes on increased occlusive arterial disease in the lower extremities, and relative protection against AAAs. Type 2 diabetes is associated with increased aortic stiffness, without abnormalities in aortic diameter or carotid stiffness. Subjects with type 2 diabetes demonstrate a decreased reflection magnitude, which may indicate an increased penetration of pulsatile energy to distal vascular beds.

Advanced glycation end products (AGEs) might also play a role in these different effects of diabetes mellitus on vascular tissues. AGEs are formed by non-enzymatic glycemic and oxidative stress reactions. Diseases with increased glycemic or oxidative stress are associated with increased AGEs levels. Indeed, increased serum and skin AGEs are found in patients with diabetes mellitus compared to controls. Oxidative stress plays a role in the pathogenesis of both AAA and PAD. In line with this
reasoning, increased serum AGEs and skin AGEs were found in patients with PAD as compared to controls.\textsuperscript{16,17} Furthermore, our data show increased skin AGEs, noninvasively assessed, in 252 AAA patients compared to controls (Boersema J et al., unpublished data, 2016). Two effects of AGEs have been described, namely formation of cross-links on long-lived proteins, and induction of cellular stress responses by engagement of the receptor for AGEs (RAGE).\textsuperscript{18} Cross-linking causes stiffness, but may also protect against mechanical structure loss of the affected tissues. The association between AGEs and arterial stiffness is shown in several studies. In patients with end-stage renal disease as well as in type 1 diabetes mellitus, skin AGEs level was associated with pulse wave velocity, a measure of arterial stiffness.\textsuperscript{19,20} Increased arterial stiffness is associated with atherosclerotic disease.\textsuperscript{21} However, in a mouse model homogenous stiffening reduced aneurysm growth, while segmental aortic stiffening caused aneurysm growth.\textsuperscript{22}

Therefore, the aim of the ARTERY study (Advanced glycation end products in patients with peripheral artery disease and abdominal aortic aneurysm study) is to identify the association between the presence of diabetes mellitus and increased accumulation of AGEs and arterial stiffening in the vascular wall in AAA and in PAD patients. If an association would be found, it would support our hypothesis that increased AGE accumulation may cause homogenous stiffness of the vascular wall and protects against AAA development and growth, but increases atherosclerosis in PAD.

Methods

Objectives

Primary objectives
To compare the amount of AGEs in the vascular wall between patients with and without diabetes mellitus suffering from:
- Abdominal aortic aneurysm.
- Peripheral artery disease.

Secondary objectives
Our secondary objectives are:
- To compare types and location of the AGEs between patients with and without diabetes mellitus.
- To evaluate the relation between the amount of AGEs and vascular stiffness within all subgroups.
To correlate AGEs in the vascular wall to serum, urine and skin AGEs in the whole study group.

**Study design**

The ARTERY study is designed as a cross-sectional multi-center study, performed in three centers in The Netherlands.

**Participants**

Patients from at least 18 years on, willing to participate are eligible for the study. Patients will be included in case of the diagnosis AAA or PAD in whom an indication for open surgery exists. Patients with concomitant AAA and PAD will be excluded as well as patients with inflammatory disorders will be excluded. Detailed inclusion and exclusion criteria are shown in Table 1. Patients will be divided into four groups, based on the diagnosis AAA or PAD and presence or absence of diabetes mellitus. American Diabetes Association guidelines will be used to define diabetes mellitus. The diagnosis diabetes mellitus will be supposed by a random glucose ≥11.1 mmol/L, HbA1c ≥6.5% or 48 mmol/mol, or a history of diabetes mellitus.¹

### Table 1. Inclusion and exclusion criteria

<table>
<thead>
<tr>
<th>Inclusion criteria</th>
<th>Exclusion criteria</th>
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</thead>
<tbody>
<tr>
<td><strong>AAA</strong></td>
<td></td>
</tr>
<tr>
<td>An aneurysm of the abdominal aorta ≥50 mm for women, ≥55 mm for men, or a growth rate of ≥10 mm per year as demonstrated on duplex ultrasound, computed tomographic angiography, or magnetic resonance angiography. And scheduled for an open repair of the aneurysm.</td>
<td>Patients with signs of an inflammatory or mycotic aneurysm on computed tomographic angiography.</td>
</tr>
<tr>
<td>PAD</td>
<td></td>
</tr>
<tr>
<td>Proven PAD with an ankle-brachial index &lt;0.80, or a toe pressure &lt;50 mmHg, or confirmation of PAD on duplex ultrasound, computed tomographic angiography, magnetic resonance angiography, or angiography. And scheduled for open bypass surgery or open endarterectomy of the lower extremity arteries.</td>
<td>Patients with PAD and an abdominal aorta ≥30 mm. PAD caused by local radiation therapy.</td>
</tr>
</tbody>
</table>

AAA indicates abdominal aortic aneurysm; PAD, peripheral artery disease.
In total, 120 patients will be included in the ARTERY study. This number is based on two sample size calculations. For our primary objective, calculation was based on a study which compared the content of the specific AGE pentosidine in plaques of diabetic and non-diabetic patients with carotid artery stenosis. In this study, a significant difference was found in 7 patients with diabetes mellitus (197.0 ± 34.0 pmol/mg collagen) and 20 patients without diabetes mellitus (111.9 ± 16.2 pmol/mg collagen). With a power of 80% and an alpha of 5%, each group has to consist of 3 patients. With four groups in our study this would result in at least 12 patients. However, for our secondary objectives, an additional sample size calculation was performed. This calculation was based on skin AGEs in patients with and without diabetes mellitus. This study reported a mean skin AGE level of 2.79 ± 0.8 arbitrary units (AU) in 973 patients with diabetes mellitus and 2.14 ± 0.6 AU in 231 patients without diabetes mellitus. To show a similar difference between patients with and without diabetes mellitus of skin AGEs, 25 patients per group are required. Taking a failure plus dropout rate of 20% into account together with the possibility of subgroup analysis, we should include 30 patients per subgroup, and a total of 120 patients.

Recruitment
The patients will be recruited from the outpatient clinic of vascular surgery located in one of the three following Dutch hospitals: University Medical Center Groningen (UMCG), Martini Hospital Groningen, and Medisch Spectrum Twente (MST).

Patients will be informed about the study by their treating vascular surgeon. Detailed written and oral information will be given by the investigator or research coordinator.

Study procedures
After signing the informed consent form, patients are asked to fill in a questionnaire and have an appointment in the vascular laboratory. Blood will be drawn and 24-hour urine will be collected before surgery. Several biopsies will be obtained during surgery, depending on the type of surgery, and of which more details are shown below. Collected tissue biopsies will be stored until analysis.

Blood and urine sampling
Blood will be sampled for routine analysis and stored to determine AGEs. For direct analysis blood will be drawn in an EDTA tube, lithium-heparin tube and a sodium fluoride tube. Citrate plasma, EDTA plasma and serum will be stored at -80° Celsius. For the analysis of AGEs, EDTA tubes will be used for blood collection, which will be
transported on ice, centrifuged on 4° Celsius and stored at -80° Celsius until batch analysis.

Patients will be asked to collect urine for 24 hours and to keep the container cool until the urine is returned to the laboratory. The urine will be centrifuged, routine analysis will be performed and samples will be stored at -80° Celsius.

**Biopsy**
During surgery, full-thickness biopsies from the arterial wall will be taken at predefined locations. These biopsies will be marked with sutures to assure cranio-caudal directions.

- In case of an open aneurysm repair, a biopsy will be obtained 5 centimeter below the lowest renal artery on the left ventral side with a size of 3 (length) x 1 (width) centimeter.
- In case of bypass surgery for PAD, a biopsy will be obtained from the proximal anastomosis in the groin of 1 (length) x 0.5 (width) centimeter.
- In case of an endarterectomy for PAD, usually the common femoral artery, a biopsy will be obtained from the proximal part of the artery with a size of 1 (length) x 0.5 (width) centimeter.

In addition, skin tissue, venous tissue and fat tissue will be obtained and stored as described below. Skin tissue will be obtained from the incision with a width of 1 (length) x 0.5 (width) x 1 (depth) centimeter. Venous tissue is only available in case of an autologous venous bypass. In that case, a circular segment from the venous graft will be obtained at the level of the distal anastomosis with a size of 0.25-0.50 centimeter. Peri-aortic adipose tissue will be obtained during open aneurysm repair.

**Storage**
After surgery, biopsy material of the arterial tissue will be divided into three equal parts. The first two parts, together with skin, venous and adipose tissue, will be snap frozen and stored at -80° Celsius. The third part, marked with a suture on the most distal part of the biopsy, will be stored in formaldehyde for 24-72 hours. Afterwards, tissue will be dehydrated with ethanol and embedded in paraffin. Materials of MST will be fixed with formaldehyde, stored in 96% ethanol, and transported to the UMCG per batch of 10 samples where these tissues will be embedded in paraffin.
Chapter 8

Outcome measures

Table 2 shows an overview of the outcome measures and measurement methods, which will be described in detail in the following paragraphs.

Demographics
The patients’ characteristics such as date of birth, sex, cardiovascular risk factors, cardiovascular disease, cardiovascular family history and drug use will be assessed by evaluating medical records and will be confirmed or complemented by a questionnaire consisting of 25 questions.

Weight and length will be recorded during visit at the vascular laboratory. Routine blood and urine analysis will be performed to evaluate the presence of diabetes mellitus and to determine several cardiovascular risk factors, including renal function.

AGEs in arterial wall
Analysis of AGEs in the arterial wall, serum and urine will be performed by the laboratory for Metabolism and Vascular Medicine, Maastricht University Medical Center, The Netherlands. Protein-bound Nε-carboxymethyl-lysine (CML), Nε-carboxyethyl-lysine (CEL) and methylglyoxal-derived hydroimidazolone (MG-H1) will be measured in the first part of the arterial biopsy material with ultra-performance liquid chromatography tandem mass spectrometry (UPLC-MS/MS), as described previously. To localize different types of AGEs, 4 millimeter slices will be made from the paraffin embedded arterial tissue. Consecutive slices will be stained with several antibodies, including polyclonal anti-AGE antibody (Abcam, ab23722), monoclonal anti-AGEs antibody for lysine derivates (Biorbyt, orb27490) and monoclonal anti-Pentosidine (Biorbyt, orb27502).

AGES in fluids
Specific protein-bound plasma and urine AGEs, CML, CEL and MG-H1, will be measured with UPLC-MS/MS, as described previously. Protein-bound plasma and urine pentosidine will be measured using high performance liquid chromatography (HPLC), as described previously.
Skin AGES
AGEs in the skin will be assessed noninvasively with the AGE Reader™ (DiagnOptics Technologies BV, Groningen, The Netherlands). This method uses ultraviolet light to excite specific AGEs. Reflection and emission light are detected and converted into a skin autofluorescence level expressed in arbitrary units. Detailed information about this method is described elsewhere. Both inner forearms will be scanned three times. Patients will be asked not to use sun blockers, skin tanners or skin creams two weeks before the measurement, since these creams influence the skin autofluorescence measurement.

Vascular parameters
Several vascular parameters will be obtained for the ARTERY study. These parameters include blood pressures, toe pressures, and duplex ultrasound of the aorta. In addition, carotid artery intima-media thickness and carotid-femoral pulse wave velocity will be measured in the vascular laboratory of the UMCG only. Measurements will be performed by trained vascular technicians.

Brachial artery blood pressures will be measured three times on both arms in sitting position after 10 minutes of rest.

For the ankle-brachial index measurement, patients should be in supine position. For the ankle-brachial index, the systolic pressure will be measured of the brachial artery, and both the posterior tibial artery and the dorsalis pedis artery bilaterally. This will be measured during cuff release using a Doppler probe. Toe-brachial index will be measured using toe photo-electric plethysmography during gradual pressure release of a cuff placed at the base of the first toe, the toe pressure being defined by reappearance of the plethysmographic signal.

Screening to exclude concomitant AAA and PAD will be performed with duplex ultrasound in supine position. An AAA will only be reassessed in case the last imaging procedure was more than two months ago.

Carotid intima-media thickness will be measured as a marker of generalized atherosclerotic burden. The far wall of the left and right common carotid artery will be measured with the MyLabOne (Esaote Europe B.V., Maastricht, The Netherlands). Three measurements will be performed with an average of 6 heart beats of the wall segment of 10 millimeters before the transition to the carotid bulb.
For the measurement of the carotid-femoral pulse wave velocity, patients are asked not to drink coffee or alcohol and not to smoke three hours before the measurement. Patients are allowed to have a light meal. Arterial waveforms will be obtained from the carotid and femoral artery and the time delay will be measured between the feet of the two waveforms. The distance covered by the waves will be established as 80% of the distance between the two recording sites. Pulse wave velocity will be calculated as distance/time delay in meters/seconds with the use of the Sphygmocor EM3 (AtCor Medical Pty Ltd, West Ryde, Australia).

Table 2. Measures and methods

<table>
<thead>
<tr>
<th>Domain</th>
<th>Outcome measure</th>
<th>Methods/equipment</th>
</tr>
</thead>
<tbody>
<tr>
<td>Demographics</td>
<td>Cardiovascular risk factors</td>
<td>Evaluation medical records</td>
</tr>
<tr>
<td></td>
<td>Cardiovascular disease</td>
<td>Questionnaire</td>
</tr>
<tr>
<td></td>
<td>Cardiovascular family history</td>
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<td></td>
<td>Drug use</td>
<td></td>
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<tr>
<td></td>
<td>Diabetes mellitus</td>
<td>Routine blood analysis including random glucose, HbA1c,</td>
</tr>
<tr>
<td></td>
<td>Renal function</td>
<td>creatinine, total cholesterol, LDL, HDL, triglycerides,</td>
</tr>
<tr>
<td></td>
<td>Lipid profile</td>
<td>ALAT, ASAT, gamma-gt, LDH, TSH, T4, hsCRP.</td>
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<tr>
<td></td>
<td>Liver function</td>
<td>EDTA, li-heparine tube, NaF tube.</td>
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<tr>
<td></td>
<td>Thyroid function</td>
<td></td>
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<tr>
<td></td>
<td>Weight</td>
<td>Seca 877 scale</td>
</tr>
<tr>
<td>AGEs in arterial wall</td>
<td>Quantitative measurement</td>
<td>High performance liquid chromatography</td>
</tr>
<tr>
<td></td>
<td>Types and localization</td>
<td>Immunohistochemistry</td>
</tr>
<tr>
<td>AGEs in fluids</td>
<td>Serum AGEs</td>
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<tr>
<td></td>
<td>Urine AGEs</td>
<td>High performance liquid chromatography</td>
</tr>
<tr>
<td>Skin AGEs</td>
<td>Skin autofluorescence</td>
<td>AGE Reader™</td>
</tr>
<tr>
<td>Vascular parameters</td>
<td>Blood pressure</td>
<td>Microlife WatchBP Office</td>
</tr>
<tr>
<td></td>
<td>Ankle-brachial index</td>
<td>Vicorder</td>
</tr>
<tr>
<td></td>
<td>Diameter aorta</td>
<td>Siemens Acuson S2000</td>
</tr>
<tr>
<td></td>
<td>Intima-media thickness</td>
<td>Esaote MyLabOne</td>
</tr>
<tr>
<td></td>
<td>Pulse wave velocity</td>
<td>Sphygmocor EM3</td>
</tr>
</tbody>
</table>

AGEs indicates advanced glycation end products.

Data handling and statistical analysis

Informed consent forms will be collected and stored at the UMCG. Biomaterials from patients will be coded. All generated data will be stored coded in an electronic Case Report File with the software OpenClinica. Data will be exported to SPSS software (IBM Corp. Armonk, NY, USA) for statistical analysis.

Described data will be shown as mean with standard deviation for normal distributed variables, mean with interquartile range for skewed data and number with
percentages for categorical data. Difference between groups will be tested with the independent Student $t$-test or non-parametric tests for skewed data. For categorical variables, Chi-square test will be performed. A $P<0.05$ will be considered significant.

**Ethical considerations**

Ethical approval was obtained from the UMCG Medical Ethics committee (METc 2014/269). Local approval for the ARTERY study was given by the Medical Ethics committee and the board of directors of the Martini Hospital (METc 2014/269) and MST (METc H15-076). All investigators performed and passed the exam of the Dutch Basic course on Regulations and Organisation for clinical investigators.

**Discussion**

The ARTERY study is designed to define the possible role of AGEs in the protection against AAA formation in diabetes mellitus. The protocol for this cross-sectional multicenter study addresses this issue by combining local and systemic AGE assessments with vascular functional and imaging methods.

Several research groups have evaluated the localization and content of AGEs in human tissue before. Most studies have been performed assessing skin AGE levels.\(^{29}\) Furthermore, Hoffman and colleagues described the measurement of AGEs in right atrial appendages and vein graft material from patients with coronary artery disease.\(^{30,31}\) However, to our knowledge, this is the first study that will compare AGEs accumulation in human vascular tissue and will investigate the effects of diabetes mellitus on AGE accumulation and distribution in different vascular diseases. Furthermore, the current study has been designed to provide answers on several additional research questions, including the association between tissue AGEs and vascular stiffness. In addition, this study will show the association between skin AGEs measured noninvasively and AGEs measured in vascular tissue, urine and blood.

The AGE Reader\(^{\text{TM}}\) uses fluorescence for assessing skin AGEs. Skin autofluorescence (SAF) is strongly associated with AGEs from skin biopsies.\(^ {27}\) Several studies have shown that skin autofluorescence, measured with the use of the AGE Reader\(^{\text{TM}}\), is related to arterial parameters such as pulse wave velocity and intima-media thickness, and predicts cardiovascular events.\(^ {19,32-35}\) However, the correlation between AGEs measured in the arterial wall compared to skin autofluorescence is unknown. It is likely that there is a correlation between both, since Hofmann et al. showed that skin AGE
levels were strongly correlated to AGEs in cardiac tissue from patients with coronary artery disease. A strong association between AGEs of the arterial wall and AGEs measured with skin autofluorescence would underscore the role of the AGE Reader™ as an easy and noninvasive technique as a marker of vascular tissue accumulation.

Although we designed our study carefully, there are some limitations. We do not exclude patients suffering from renal insufficiency. These patients also have increased AGEs accumulation. We have chosen not to exclude these patients, since patients with vascular disease frequently have renal insufficiency. Exclusion of concurrent renal failure and vascular disease would result in a selection bias. Therefore, we have decided to increase our sample size to allow subgroup analysis. In addition, renal function will be a variable which will be included into multivariate analyses. In addition, biopsy material obtained at the location of disease may not be representative for vascular tissue in general. We standardize the biopsies procedure to minimize location specific differences. However, it is technically and ethically not possible to obtain additional biopsies from different vascular sites to determine whether AGEs are distributed equally.

Conclusion

In conclusion, the ARTERY study will investigate the diverging effects of diabetes mellitus on AAA and PAD and the role of AGEs as possible explanatory factor. The study will be conducted in a multicenter setting. Primary objective is to analyze AGEs in the arterial wall. As secondary objectives, AGEs of the arterial wall will be compared to AGE measurements in blood, urine and skin, and with vascular wall parameters.

Acknowledgements

The authors thank Hannie Westra, PhD and Berber Doornbos-van der Meer, MSc, Department of Rheumatology and Clinical Immunology, Groningen, The Netherlands for their expert advice of immunohistochemical analysis and antibodies. We thank also the technicians of the vascular laboratory UMCG for helping with the protocol of the vascular measurements and performing preliminary tests: Saskia van de Zande, MSc, Anne van Gessel, MSc, Arie van Roon, PhD. Finally, the authors wish to thank the research coordinator Anja Stam for her help setting up the study in MST.
References


