Feasibility of measuring renal blood flow by phase-contrast magnetic resonance imaging in patients with autosomal dominant polycystic kidney disease

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Abstract

Objectives Renal blood flow (RBF) has been shown to predict disease progression in autosomal dominant polycystic kidney disease (ADPKD). We investigated the feasibility and accuracy of phase-contrast RBF by MRI (RBFMRI) in ADPKD patients with a wide range of estimated glomerular filtration rate (eGFR) values.

Methods First, we validated RBF MRI measurement using phantoms simulating renal artery hemodynamics. Thereafter, we investigated in a test-set of 21 patients intra- and interobserver coefficient of variation of RBF MRI. After validation, we measured RBFMRI in a cohort of 91 patients and compared the variability explained by characteristics indicative for disease severity for RBFMRI and RBF measured by continuous hippuran infusion.

Results The correlation in flow measurement using phantoms by phase-contrast MRI was high and fluid collection was high (CCC=0.969). Technical problems that precluded RBFMRI measurement occurred predominantly in patients with a lower eGFR (34% vs. 16%). In subjects with higher eGFRs, variability in RBF explained by disease characteristics was similar for RBFMRI compared to RBFHip, whereas in subjects with lower eGFRs, this was significantly less for RBFMRI.

Conclusions Our study shows that RBF can be measured accurately in ADPKD patients by phase-contrast, but this technique may be less feasible in subjects with a lower eGFR.

Key points

- Renal blood flow (RBF) can be accurately measured by phase-contrast MRI in ADPKD patients.
- RBF measured by phase-contrast is associated with ADPKD disease severity.
- RBF measurement by phase-contrast MRI may be less feasible in patients with an impaired eGFR.

Keywords Renal blood flow measurement · Phase-contrast MRI · Hippuran · ADPKD · Phantom

Introduction

Most autosomal dominant polycystic kidney disease (ADPKD) subjects show progressive kidney function decline and develop end-stage renal disease (ESRD) between their 4th to 7th decade of life [1]. With renoprotective treatments emerging, such as vasopressin V2-receptor antagonists and somatostatin analogues [2, 3], it will be important to assess disease severity. Especially those patients with a high likelihood of disease progression towards ESRD will need treatment. Assessing disease severity and prognosis in ADPKD is not without difficulty. Glomerular filtration rate (GFR) may not be the best measure in this respect, because GFR remains within normal limits for an extended period of time [4]. Therefore, total kidney volume (TKV) assessed by MRI has been
investigated as an alternative [5]. TKV measurement is, however, laborious and the association between TKV and disease severity is limited at an individual patient level. Renal blood flow (RBF) has been proposed as a third marker to assess disease severity [6]. A decrease in RBF flow paralleled TKV growth in the observational CRISP cohort study [6] and is thought to precede decline in GFR [7].

However, the value of RBF has been investigated in a single study, using gadolinium-contrast to obtain a reference image of the renal artery to position the phase-contrast perpendicular to the vessel contour and included only ADPKD subjects with a creatinine clearance > 70 ml/min [6, 7]. Confirmation, especially in subjects with lower kidney function, is therefore needed. Moreover, a method without gadolinium administration would be preferable to avoid the risks of developing gadolinium-associated contrast-induced nephropathy [8] and nephrogenic systemic fibrosis [9, 10]. Besides MRI, RBF can also be measured by infusion of para-aminohippuric acid (RBF Hip) [11, 12].

Given these considerations, the aim of the present study was to investigate whether measurement of RBF by MRI without using gadolinium contrast (RBF MRI) could be performed accurately and precisely in a group of ADPKD patients with a wide range in kidney function. First, we validated RBF MRI measurement using phantoms simulating renal artery hemodynamics. Thereafter, we investigated in a test-set of patients the intra- and inter-observer variability. After validation, we compared RBF MRI and RBF Hip, and investigated associations of RBF MRI and RBF Hip with characteristics indicative for disease severity in ADPKD subgroups.

Methods and Materials

Patient population

For this study, all consecutive patients with ADPKD visiting our out-patient clinic from January 2007 until September 2012 were asked to participate. The diagnosis of ADPKD was made based upon the revised Ravine criteria [13]. Subjects were considered ineligible if they received renal replacement therapy, had undergone renal surgery, were unable to undergo MRI, and had other systemic diseases or treatments potentially affecting kidney function. One hundred forty six subjects met these criteria, and were invited for an extensive, one-day screening. This screening included, among others, an MRI and assessment of effective renal plasma flow (ERPF), which were used to measure RBF MRI and RBF Hip. Thirteen patients refused to participate and in thirteen patients, RBF MRI was not measured due to logistical reasons, leaving 120 patients for the present analyses. This study was conducted in accordance with the guidelines of the Helsinki Declaration. All participants gave written informed consent.

Measurement and calculations

All participants collected a 24-hour urine sample the day preceding the screening at our facility, in which albuminuria was measured. During the visit, blood pressure was measured (Dinamap) and fasting blood samples were drawn for determination of osmolality, copeptin and creatinine. GFR was estimated using the CKD-EPI (Chronic Kidney Disease Epidemiology Collaboration) equation [14, 15]. MRI and measurement of renal hemodynamic parameters were performed per protocol on the same day.

Renal blood flow measured by MRI

Imaging was performed on a 1.5-Tesla Magnetom Avento (Siemens, Erlangen, Germany). Two coils were placed on the anterior and posterior abdominal walls directly over the kidneys. Two fast, respiratory-triggered, coronal and transversal T2-weighted image sets were acquired to determine the kidney volumes [16]. To localize the renal arteries, coronal and transversal data sets were scanned during the breath hold using a T2-weighted TrueFISP sequence. RBF was assessed with phase-contrast MRI (PC-MRI), using a two-dimensional fast gradient echo phase-contrast sequence with retrospective cardiac gating (30 cardiac phases) and flow compensation [17]. Acquisition parameters TR/TE/FA were 43.6 ms/4 ms/30°. A single slice was positioned perpendicular to the renal artery. Slice thickness was 5 mm, FOV was 32 cm, and the number of phase-encoding steps was 260-384, depending on patient size. Acquisition time was 150-160 seconds. The encoding velocity (VENC) was 100 cm/s and flow acquisition was obtained in the slice direction. Supplementary Figure 1 shows a representative sample of data recorded during the cardiac cycle and shows a realistic renal arterial waveform. RBF MRI measurement lasted between 150 and 160 seconds. Flow analysis was performed using ARGUS flow software (Siemens, Erlangen, Germany) for phase-contrast imaging. Images were magnified to 5.0 and renal artery contours were drawn manually on anatomical images to define the region of interest for the corresponding phase-contrast images at all phases of the cardiac cycle. The drawing of these contours required 5-10 minutes per kidney. Flow rates of all renal arteries (left, right and any additional arteries) were added to calculate total RBF MRI. Flow measurements were judged technically unsatisfactory when they met one or more of the following criteria: indistinct vessel contour (blurred as a result of motion), artefacts in the vessel area, ghosting (movement artefacts), or non-physiologic, heterogeneous measurement or weak signal [18]. TKV was measured on T2-weighted coronal images using Analyze Direct 8.0 (Analyze Direct, Inc., Overland Park, KS, USA) and divided by height. Intra- and inter-observer coefficients of variation for TKV measurement were 1.8 and 2.3 %, respectively.
Validation of flow measured by MRI in a phantom

Two flexible silicon phantoms, each representing the aorta and two renal arteries perpendicular to the aorta (Elastrat, Geneva, Switzerland) (Fig. 1) were used to assess accuracy of the PC-MRI measurements. The four renal arteries in these two phantoms had diameters of 5, 6, 7 and 8 mm, representing the normal range of renal artery diameters. The phantoms were suspended in a container filled with water. An MRI-compatible pump (Cardioflow 1000MR, Shelly Medical Imaging Technologies, London, Canada) was positioned proximally; one renal artery was connected to a container, the other renal artery and distal aorta were connected to a reservoir (Fig. 1). A mix of 40-% glycerol and 60-% purified water was used as flow medium to mimic blood viscosity (~3.5 mPa/s) [19]. A synchronized trigger pulse signal was generated by the MRI to simulate electrocardiographic (ECG) gating at a heart rate of 60 beats per minute. The flow in the phantom was determined simultaneously by PC-MRI according to the protocol described above and by fluid collection (volume pumped through the renal artery was collected in a container). These flow collections were considered the gold-standard measure to evaluate the accuracy of the MR-calculated flow. Importantly, the system was positioned in the horizontal plane and the container was placed floating in the reservoir to prevent any steady flow bias due to the principle of communicating vessels (Fig. 1). During our phantom experiment, the pump was set at different flow rates (800, 1000, 1200 and 1400 ml/min) and acted as "cardiac output". In this phantom, "cardiac output" is divided over two renal arteries and the aorta. Thus, the blood flow rates in the phantom renal arteries were dependent on the pump rates ("cardiac output") and the diameters of the phantom renal arteries and aorta. Flow was measured in one renal artery at a time, but several times to assess reproducibility.

Variability of RBF_{MRI} measurement

RBF_{MRI} measurements were performed by two research physicians (ES and EM) whom were trained to perform RBF_{MRI} measurements. During their training period, they measured 60 RBF_{MRI} values under supervision of an experienced MRI technician (PK). Subsequently, they measured RBF_{MRI} on 2 occasions, blinded to previous results, in 21 patients (i.e., 42 kidneys) to determine intra- and inter-observer variability.

RBF measured by 131I-hippuran

Measurement of renal hemodynamic parameters was performed using a constant infusion method with 131I-hippuran to measure ERPF [11, 20]. During these assessments, patients were not water-loaded, but allowed to drink ad libitum. At 08.00 am, a priming solution of 0.04 mL/kg body weight was administered, followed by a constant infusion of the tracer at a rate depending on eGFR for 5.5 hours. After a stabilisation period of 1.5 hours, two 2-hour clearance periods followed, in which ERPF was measured as (I*V)/P, where I*V represents the infusion of the tracer and P represents the tracer concentration in plasma. Intra-subject day-to-day variability using this method is 5 % [11]. RBF_{Hipp} was calculated using the formula ERPF/(1-hematocrit).

Statistical analysis

Lin’s concordance correlation coefficient (CCC) and Bland-Altman analyses were used to assess agreement between the gold standard fluid collection and the flow measured by MRI in the phantom. Reproducibility was assessed by CCC, Bland-Altman analyses and intra- and inter-observer coefficient of variation (CV).

Subsequently, RBF_{MRI} and RBF_{Hipp} were measured in an ADPKD patient cohort of which baseline characteristics were calculated for the overall population and for participants with an eGFR >70 and ≤70 ml/min*1.73m², separately. Normality was assessed by a Q-Q-plot. Normal distributed variables were expressed as the mean ± standard deviation (SD), whereas non-normal distributed variables were expressed as the median (interquartile range). Difference in baseline characteristics between two eGFR subgroups was tested using a Chi-square test for categorical data, and a Student’s t-test when normally distributed or a Mann-Whitney test when not normally distributed.

Disease characteristics were correlated to RBF_{Hipp} and to RBF_{MRI} in the overall cohort and in subcohorts stratified for eGFR >70 and ≤70 ml/min*1.73m². To investigate the best fit for RBF_{Hipp} and RBF_{MRI}, we performed a stepwise backwards

Fig. 1 Schematic representation of the phantom used to study accuracy and reliability of flow measured by MRI when compared to fluid collection: A silicon phantom of the aorta and two renal arteries; B pulsatile pump; C reservoir; and D container in which fluid output was collected for flow measurement.
regression analysis. Clinically important disease characteristics were entered and deleted stepwise from the model in order of weakest association until a model with the best fit (i.e., adjusted R$^2$) was obtained. Model discrimination was determined by bootstrapping (2000 times) to calculate the difference in adjusted R$^2$ of two multivariate linear regression models [21].

All analyses were performed with SPSS version 20.0 (SPSS Inc., Chicago, IL, USA). A two-sided $p$-value less than 0.05 was considered to indicate statistical significance.

Results

Validation of flow measured by MRI

Figure 2 shows a scatter-plot comparing pulsatile flow measured by fluid collection and flow measured by PC-MRI, combined for all flow rates and vessel diameters. The 5-mm diameter renal artery images were judged technically unsatisfactory, because of artefacts in the vessel area and measurement heterogeneity. An excellent agreement between flow measured by the gold standard of fluid collection and by PC-MRI was observed. The CCC for 6 to 8 mm was 0.969 ($p < 0.001$), indicating high agreement. Bland-Altman analysis shows that flow measured by MRI on average underestimated flow measured by fluid collection by 2.9% (Fig. 2). A significant correlation was noted between the mean (flow measured by fluid collection and by MRI) and the absolute difference between these two measures ($r = -0.47$, $p = 0.004$).

Reproducibility of RBF$_{MRI}$ measurement in a test-set of ADPKD patients

Figure 3 shows the agreement between blood flow measured by observers ES and EM in 21 ADPKD patients. The average intra-observer CV was 2.3% (ES 2.0% and EM 2.6%). The estimated reliability, i.e., intra-class correlation coefficient, for observers ES and EM were 0.997 and 0.995, respectively (both $p < 0.001$). The inter-observer CV was 3.5% with a reliability coefficient of 0.991 ($p < 0.001$). In patients with an eGFR $>70$ ml/min*1.73m$^2$, the intra- and inter-observer CVs were 1.5 and 3.5%, respectively, whereas in patients with an eGFR $\leq 70$ ml/min*1.73m$^2$, these values were 3.2 and 3.5%, respectively.

RBF$_{MRI}$ measurement in an ADPKD patient cohort

Out of the 120 patients, 29 (24%) had RBF measurements that were judged technically unsatisfactory, and these patients were excluded for further analysis, leaving 91 ADPKD patients for analyses (in patients with an eGFR $\leq 70$ and $>70$ ml/min*1.73m$^2$, this held for 18 (34%) and 11 (16%) patients, respectively). The excluded patients had a significantly lower eGFR and ERPF compared to the patients that could be studied [eGFR 79.2 vs. 58.1 ml/min*1.73m$^2$ ($p = 0.001$)], and ERPF 263.5 vs. 201.7 ml/min*1.73m$^2$ ($p = 0.004$). No significant difference in TKV was observed between the groups (969 vs. 1526 mL/m, $p = 0.28$). The included patients were relatively young with a mean age of 39.8 ± 11.4 years and their eGFR ranged from 15 to 136 ml/min*1.73m$^2$. Nine patients (9.9%) had multiple renal arteries (one bilateral, seven

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unilateral and one unilateral with two additional renal arteries). RBF MRI ranged from 244 to 1547 ml/min·1.73m². Right artery RBF MRI was significantly higher than left artery RBF MRI (456 vs. 358 ml/min·1.73m², p < 0.001). The smallest renal artery diameter was 3.9 and the largest was 11.4 mm. The additional renal arteries had smaller diameters, with a mean of 6 mm compared to 7 mm of "single" renal arteries. As expected, RBF was significantly lower in patients with lower eGFR whereas age, TKV and use of antihypertensives were significantly higher in this subgroup (Table 1). Figure 4 shows examples of PC-MRI flows that were judged technically satisfactory and unsatisfactory.

**Associations of RBF MRI and RBF Hip with disease characteristics.**

RBF MRI correlated significantly with RBF Hip (r = 0.81, p < 0.001), although RBF MRI was higher than RBF Hip over the entire range of RBF MRI. In univariate regression analyses, both RBF MRI as well as RBF Hip were positively correlated with eGFR and urinary sodium excretion, and negatively with age and TKV (Table 2). A stepwise backward multiple regression model showed that RBF Hip and RBF MRI were significantly and especially associated with eGFR (both <0.001). In the whole group, the variability explained by the model for RBF MRI was significantly lower than for RBF Hip (adjusted R² = 0.61 and 0.77, respectively; p < 0.001). In patients with an eGFR >70 ml/min·1.73m², the adjusted R² was not significantly different for RBF MRI than RBF Hip (0.36 and 0.51, respectively; p = 0.19), whereas in patients with an eGFR ≤70 ml/min·1.73m², disease characteristics explained a smaller fraction of variability in RBF MRI compared to RBF Hip (0.17 and 0.72, respectively; p < 0.001; Table 3).

In addition, we performed a sensitivity analysis in which we excluded patients with additional renal arteries in order to investigate whether multiple renal arteries caused methodological problems. Multiple renal arteries were found only in patients with eGFR >70 ml/min·1.73m². We found similar variability in measurement of RBF MRI and MRI Hip as in the whole group (adjusted R² = 0.63 and 0.77, respectively), and the variability explained by the model increased slightly for RBF MRI (adjusted R² = 0.42) and remained similar for RBF Hip (adjusted R² = 0.53).

**Discussion**

In this study, we investigated whether measurement of RBF by MRI without using gadolinium contrast (RBF MRI) could be performed accurately and precisely. First, we validated RBF MRI measurement using phantoms simulating renal artery hemodynamics. Thereafter, we investigated in a test-set of patients the reproducibility of RBF MRI. After validation, we measured RBF MRI in a cohort of ADPKD patients and compared RBF MRI and RBF measured by hippuran infusion with characteristics indicative for disease severity.

In our phantom experiments, mimicking the clinical renal hemodynamic profile, RBF MRI showed perfect agreement to the gold-standard for flow measurement, i.e., fluid collection. We observed no differences in accuracy in RBF MRI between the "renal arteries" varying in diameter or between flow...
velocities, although it should be noted that measurement of RBF\textsubscript{MRI} was not feasible in the artery with the smallest diameter (5 mm). Our findings are in line with the values obtained by Dambreville et al [19], who showed that steady and pulsatile flow can be measured reliably in polyvinyl alcohol tubes with different diameters. These authors also experienced difficulties in measuring MRI flow in tubes with small diameters. In our test set of 21 patients, reproducibility was high and comparable with values obtained in the literature with contrast-measured RBF [6, 19]. These data indicate that RBF\textsubscript{MRI} can be measured reliably in vitro and in vivo. In patients without any additional renal arteries, we found similar variability in measurement of RBF\textsubscript{MRI} and MRI\textsubscript{Hip} compared to all patients. These data indicate that the presence of additional renal arteries also does not influence the reliability of RBF measurement.

We used MRI to measure RBF\textsubscript{MRI} in a cohort of ADPKD patients. Our technical success rate was 76 %, which is comparable with previous literature on this method [18]. We compared RBF\textsubscript{MRI} with RBF\textsubscript{Hip} in these patients. It should be noted that RBF\textsubscript{Hip} is another indirect measurement of RBF. RBF\textsubscript{Hip} was lower compared to RBF\textsubscript{MRI} in patients with relatively preserved as well as impaired kidney function. It is known that RBF\textsubscript{Hip} underestimates RBF, because the extraction of hippuran by tubular cells is not 100 % and dependent on kidney function [12]. In two previous studies, it was shown that RBF\textsubscript{MRI} correlated well with RBF determined by clearance of para-aminohippurate (r = 0.94 and r = 0.93, respectively) [22, 23]. These studies used healthy subjects. In the present study we applied hippuran instead of para-aminohippurate [12]. We showed that RBF\textsubscript{MRI} correlated well with RBF\textsubscript{Hip} (r = 0.90) and that RBF\textsubscript{MRI} was associated with anatomical, hemodynamic and laboratory parameters indicating disease severity. This corroborates the findings by the CRISP consortium that found the same associations and additionally described that RBF\textsubscript{MRI} predicted kidney function decline [6]. Measurement of RBF can, therefore, be important to assess disease severity, to predict prognosis and potentially also act as a surrogate endpoint to assess the efficacy of renoprotective interventions early in the disease. RBF could also be used in combination with GFR to assess the filtration fraction [24].

Although on average we showed that RBF could be measured reliably by MRI without using contrast, there are a number of issues rendering such measurement less feasible in ADPKD patients with impaired kidney function. First, RBF\textsubscript{MRI} measurement in the smallest phantom artery (5 mm), which is a surrogate for more severe ADPKD, was not possible. Second, in the phantom experiment, we found a significant correlation between mean flow and difference in flow measured by MRI and fluid collection, with a larger difference in the lower flow range. This indicates that systematic bias increases at the lower flow range. Third, variability was higher in patients with a low eGFR than in patients with a
Fourth, RBF\textsubscript{MRI} measurement in patients with lower eGFRs resulted less often in interpretable images. MRI images were judged technically unsatisfactory in 34% of patients with an eGFR ≤70 vs. in 16% of patients with an eGFR >70 ml/min*1.73m\textsuperscript{2}. Fifth, in subjects with a lower eGFR, only a small fraction of variability in RBF\textsubscript{MRI} was explained by disease characteristics. We speculate that due to the lower RBF in smaller renal arteries the phase-contrast measurement may give a (too) weak signal, causing the lower success rate. Two methods could be used to overcome this problem. The spatial resolution could be increased or the VENC could be adjusted. However, for this study, we used the protocol of the CRISP consortium [6, 7, 19] with predetermined variables (FOV 32 cm, slide thickness 5 mm, VENC of 100cm/s) as to maintain the protocol and to compare results without bias. In addition, renal arteries with larger...
Table 2  Associations of renal blood flow (ml/min*1.73m²) measured by MRI or hippuran with characteristics indicative for disease severity in ADPKD (n = 91)

<table>
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<th>Univariate analyses</th>
<th>Multivariate analyses</th>
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<tr>
<td></td>
<td>MRI</td>
<td>Hippuran</td>
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<td></td>
<td>r</td>
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<tr>
<td>Adjusted R²</td>
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<tr>
<td>Age (years)</td>
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<td>Female vs male sex (1 or 0)</td>
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<tr>
<td>Systolic blood pressure (mmHg)</td>
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</tr>
<tr>
<td>Antihypertensive drugs (Yes=1)</td>
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<td>Plasma osmolality (mOsm/kg)</td>
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<tr>
<td>Log Copeptin (pmol/l)</td>
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<tr>
<td>eGFR (ml/min*1.73m²)</td>
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<tr>
<td>Ln TKV/h (ml/m)</td>
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<td>24h urine volume (mL)</td>
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<tr>
<td>Urinary sodium excretion (mmol/24h)</td>
<td>0.23</td>
<td>0.03</td>
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<tr>
<td>Ln Urinary albumin excretion (mg/24h)</td>
<td>0.04</td>
<td>0.73</td>
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TKV/h and UAE were natural logarithm transformed and copeptin was log transformed. Abbreviations: eGFR, estimated glomerular filtration rate; TKV/h, total kidney volume/height. * Model discrimination assessed by bootstrapping (2000 times), p-value < 0.001

Table 3  Multivariate associations of renal blood flow (ml/min*1.73m²) measured by MRI or hippuran with characteristics indicative for disease severity in patients stratified for kidney function

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<th>MRI</th>
<th>Hippuran</th>
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<tr>
<td></td>
<td>Std β</td>
<td>p</td>
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<td>eGFR &gt;70 ml/min*1.73m²</td>
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<tr>
<td>N = 57</td>
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<tr>
<td>Adjusted R²</td>
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</tr>
<tr>
<td>Age (years)</td>
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<tr>
<td>Weight (kg)</td>
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<td>Antihypertensive drugs (Yes = 1)</td>
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<td>Plasma osmolality (mOsm/kg)</td>
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<td>eGFR (ml/min*1.73m²)</td>
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<tr>
<td>24h Urine volume (mL)</td>
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<td>0.03</td>
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<tr>
<td>Ln (Urinary albumin excretion mg/24h)</td>
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<tr>
<td>eGFR ≤ 70 ml/min*1.73m²</td>
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<tr>
<td>N = 34</td>
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<tr>
<td>Adjusted R²</td>
<td>0.17*</td>
<td>0.72*</td>
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<tr>
<td>Female vs male sex (1 or 0)</td>
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<tr>
<td>Weight (kg)</td>
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<td>Plasma osmolality (mOsm/kg)</td>
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<tr>
<td>Log (Copeptin pmol/l)</td>
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<td>eGFR (ml/min*1.73m²)</td>
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<td>24h Urine volume (mL)</td>
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<tr>
<td>Ln (Urinary albumin excretion mg/24h)</td>
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* Model discrimination assessed by bootstrapping (2000 times), p-value < 0.001.
diameters allow better phase-contrast positioning and easier measurement of the flow. Notwithstanding these observations, we think that this does not necessarily disqualify the use of RBF MRI measurement for ADPKD staging and risk classification. The clinical need for risk classification in this patient group is especially present early in the disease, when GFR remains within the normal limits for an extended period of time [1]. At a later stage of the disease, when kidney function starts to decline, eGFR in association with age can be used as a parameter to assess disease severity and prognosis. It should be stated that in patients with an eGFR >70 mL/min*1.73 m², gadolinium-enhanced techniques can also be used to measure RBF, because in this patient subgroup there is a low risk of toxicity. Lastly, there are other unenhanced techniques to measure flow, such as arterial spin labelling [25], but this technique is especially useful to measure regional variations in renal perfusion, and investigating this technique was beyond the scope of the present study. Of note, a pilot study showed no difference for RBF values between arterial spin labelling and RBF MRI measurement [26].

Limitations of our study are, first, that we studied renal hemodynamical parameters in a cross-sectional setting. Therefore, we were not able to investigate associations with disease progression. Second, this study assessed reliability and accuracy of RBF MRI using a phantom simulating the clinical situation. However, it lacks a gold standard method for RBF measurement in vivo. Such a method would necessitate a flow probe directly introduced in the renal arteries, which, because of its invasive character, is not feasible. We, therefore, assessed reliability of RBF MRI in an indirect way, by studying the degree of variability of RBF MRI that was explained by accepted characteristics of ADPKD severity, and by comparing this with the degree of variability of RBF HIP explained by the same disease characteristics. Strengths of our study were that we measured RBF MRI and ERPF by gold-standard techniques on the same day. These measurements were performed in a relatively large, well-phenotyped cohort of ADPKD patients with relatively preserved as well as impaired kidney function. Furthermore, our method avoids use of gadolinium, which is important because of the potential side effects of this contrast agent in patients with kidney dysfunction [27].

In conclusion, our study shows that in ADPKD patients, RBF can be measured by MRI accurately and reliably without using gadolinium contrast, but that assessment of RBF by PC-MRI may be less feasible in patients with impaired kidney function.

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Some study subjects or cohorts have been previously reported in:

Methodology: retrospective, cross sectional study, performed at one institution.

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References


