Kinetic Characterization and X-ray Structure of a Mutant of Haloalkane Dehalogenase with Higher Catalytic Activity and Modified Substrate Range†‡

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ABSTRACT: Conversion of halogenated aliphatics by haloalkane dehalogenase proceeds via the formation of a covalent alkyl−enzyme intermediate which is subsequently hydrolyzed by water. In the wild type enzyme, the slowest step for both 1,2-dichloroethane and 1,2-dibromoethane conversion is a unimolecular enzyme isomerization preceding rapid halide dissociation. Phenylalanine 172 is located in a helix−loop−helix structure that covers the active site cavity of the enzyme, interacts with the Cl of 1,2-dichloroethane during catalysis, and could be involved in stabilization of this helix−loop−helix region of the cap domain of the enzyme. To obtain more information about the role of this residue in dehalogenase function, we performed a mutational analysis of position 172 and studied the kinetics and X-ray structure of the Phe172Trp enzyme. The Phe172Trp mutant had a 10-fold higher kcat/Km for 1-chlorohexane and a 2-fold higher kcat for 1,2-dibromoethane than the wild-type enzyme. The X-ray structure of the Phe172Trp enzyme showed a local conformational change in the helix−loop−helix region that covers the active site. This could explain the elevated activity for 1-chlorohexane of the Phe172Trp enzyme, since it allows this large substrate to bind more easily in the active site cavity. Pre-steady-state kinetic analysis showed that the increase in kcat found for 1,2-dibromoethane conversion could be attributed to an increase in the rate of an enzyme isomerization step that preceeds halide release. The observed conformational difference between the helix−loop−helix structures of the wild-type enzyme and the faster mutant suggests that the isomerization required for halide release could be a conformational change that takes place in this region of the cap domain of the dehalogenase. It is proposed that Phe172 is involved in stabilization of the helix−loop−helix structure that covers the active site of the enzyme and creates a rigid hydrophobic cavity for small apolar halogenated alkanes.

The first step in the degradation of 1,2-dichloroethane by the bacterium Xanthobacter autotrophicus GJ10 is catalyzed by the enzyme haloalkane dehalogenase (Janssen et al., 1985). The enzyme converts a broad range of haloalkanes to the corresponding alcohols with water as the sole cosubstrate (Keuning et al., 1985). The protein consists of two domains (Verschueren et al., 1993a): a main domain that is formed by an eight-stranded β-sheet surrounded by α-helices, and a cap domain formed by five helices and loops. The hydrophobic active site is buried and located between these two domains. The main domain has structural similarity with several other hydrolytic enzymes, which were classified as α/β-hydrolase fold enzymes (Ollis et al., 1992).

The reaction mechanism and kinetic mechanism of haloalkane dehalogenase were deduced from X-ray crystallography, site-directed mutagenesis and pre-steady-state kinetic studies (Figure 1). The reaction is initiated by binding of 1,2-dichloroethane in the Michaelis complex, where the Cl of the substrate is stabilized between NH’s of the Trp125 and Trp175 side chains. These two tryptophans stabilize the chloride ion when the carbon−chlorine bond is cleaved upon attack of the nucleophilic O of Asp124 (Verschueren et al., 1993b; Pries et al., 1995a). Asp260 is hydrogen bonded to His289, which may activate the histidine by stabilizing the positive charge that develops on the imidazole ring when His289 accepts a proton from water. The slowest step at high substrate concentration is not cleavage of the carbon−halogen bond, but release of the charged halide ion out of the buried active site cavity (Schanstra & Janssen, 1996a; Schanstra et al., 1996b). In the wild type enzyme, halide release and halide binding can occur via two parallel routes. Halide release was found to follow mainly the route in which actual release of the halide ion from the enzyme was preceded by a slow unimolecular enzyme isomerization step (Schanstra & Janssen, 1996a).

The largely hydrophobic active site cavity of the dehalogenase is lined with the catalytic-triad residues Asp124, His289, and Asp260 and with four phenylalanines, two
MATERIALS AND METHODS

Materials. Restriction enzymes and isopropyl β-D-thiogalactopyranoside (IPTG) were obtained from Boehringer Mannheim. Bacteriophage R408, T7 polymerase, and DNA sequencing reagents were obtained from Pharmacia. Monodeoxyribonucleoside-5′-triphosphates (dNTPs) were obtained from Promega. The oligonucleotides for site-directed mutagenesis and sequencing were supplied by Eurosequence, Groningen (The Netherlands). The oligonucleotides used were pF172X (5′-AGCCCTGAGATGCNNACCCCTGGGAAA-3′, PstI restriction site underlined and mutated codon in bold) and pF172C (5′-AGCCCTGAGATGGCTTACCCCTGGGAAA-3′, PstI restriction site underlined, and mutated codon in bold). Escherichia coli strain BW313 (Kunkel et al., 1985) was used for the production of single-stranded uracil containing DNA used in mutagenesis. E. coli JM101 (Promega) was the host for production of single-stranded DNA used for sequencing and E. coli strain BL21(DE3) (Studier et al., 1990) was used for overexpression of both wild type and mutant dehalogenases.

DNA Methods. All DNA and E. coli manipulations were performed using standard molecular biology techniques described by Sambrook et al. (1989). Mutants were constructed using the method described by Kunkel et al. (1985), using vector pGELAF+ (Schanstra et al., 1993). Sequences were confirmed by dyeoxy sequencing (Sanger et al., 1977).

Protein Expression and Purification. Wild type and mutant enzymes were expressed and purified as described earlier (Schanstra et al., 1993). The cultures expressing the mutant enzymes were, after induction with IPTG, cultivated overnight at 17 °C to prevent inclusion body formation. The buffers used during purification were TEMAG [10 mM Tris-sulfate, pH 7.5, 1 mM EDTA, 1 mM 2-mercaptoethanol, 3 mM sodium azide, and 10% (v/v) glycerol] and PEMAG [10 mM sodium phosphate, pH 6.0, 1 mM EDTA, 1 mM 2-mercaptoethanol, 3 mM sodium azide, and 10% (v/v) glycerol]. The enzyme was concentrated with an Amicon ultrafiltration cell using a PM10 filter and stored in TEMAG-buffer at 4 °C.

Protein Analysis and Dehalogenase Assays. Protein concentrations of crude extracts were determined with Coomassie Brilliant Blue using bovine serum albumin as a standard. The concentration of purified wild type enzyme was determined using ε280 = 4.87 × 10^4 M⁻¹ cm⁻¹ (Schanstra & Janssen, 1996a). ε280 = 5.44 × 10^4 M⁻¹ cm⁻¹ was used for the Phe172Trp enzyme, and ε280 = 5.00 × 10^4 M⁻¹ cm⁻¹ was used for the Phe172Tyr enzyme, based on the modified protein sequences calculated with the program DNASTAR (DNASTAR Inc., Madison, WI). The purified proteins were analyzed with SDS–PAGE which showed that the purity of the preparations was greater than 99% (not shown). Dehalogenase assays were performed using colorimetric detection of halide release as described by Keuning et al. (1985). Solvent kinetic isotope effects were determined similarly with substrate dissolved in increasing concentrations of 2H₂O. The Kₘ and kcat were determined as described (Schanstra et al., 1996b). The experimental error in the data was less than 15% for the kcat values and less than 25% for the Kₘ values.

Pre-Steady-State Kinetic Experiments. Rapid-quench-flow experiments were carried out at 30 °C on a rapid-quench-flow instrument (RQF-63) from KinTek Instruments (Johnson, 1992) as described by Schanstra et al. (1996b). All reported concentrations are in the reaction line of the rapid-quench-flow instrument.

Stopped-flow fluorescence was used to study the kinetics of halide binding and substrate conversion. All experiments...
were performed on an Applied Photophysics model SX17MV stopped-flow fluorospectrophotometer at 30 °C. Excitation was at 290 nm and fluorescence emission from Trp residues was followed through a 320 nm cutoff filter supplied with the instrument. The reported concentrations are those in the reaction chamber.

The buffer used during stopped-flow fluorescence and rapid-quench-flow experiments was T50EMA-buffer (50 mM Tris-sulfate, pH 8.2, 1 mM EDTA, 1 mM 2-mercaptopoethanol, and 3 mM sodium azide) or T50EDA-buffer (50 mM Trissulfate, pH 8.2, 1 mM EDTA, 1 mM dithiotheritol, and 3 mM sodium azide). T50EDA-buffer was used in rapid-quench experiments with 1,2-dibromoethane, since 2-mercaptopoethanol interfered with 2-bromothiolactone during gas chromatographic analysis. All substrate solutions were prepared in T50EMA or T50EDA. Halide binding was performed in T50EMAG [T50EMA-buffer with 10% (v/v) glycerol]. Glycerol was present during halide binding experiments since it improved the reproducibility of the stopped-flow fluorescence binding curves, but did not affect the kinetics.

**Kinetic Data Analysis.** Analysis of rapid-quench and stopped-flow fluorescence data was done by numerical simulation using the computer program Gepasi (Mendes, 1993; Gepasi for MS-Windows, version 2.0, release 2.08) coupled to the spreadsheet program Quattro-Pro 5.0 for MS-Windows (Borland International, Inc.).

The combined schemes for halide release (Schanstra & Janssen, 1996a) and substrate conversion (Schanstra et al., 1996b) by the dehalogenase are given in Scheme 1 (1A, bromide; 1B, chloride). In this scheme, E₁ is the normal native enzyme, Eₙ₁ is the kinetically observed conformational isomer to which halide ions can rapidly bind, and E₁X is the kinetically observed collision complex in which halide is weakly bound to native enzyme (Schanstra & Janssen, 1996a). Rate and equilibrium constants for halide binding under pseudo-first-order conditions were obtained by numerical simulation of Scheme 1A and 1B as described. Apparent steady-state dissociation constants (Kₐ) for halide binding were determined as described by Kennes et al. (1995). Apparent dissociation constants were also calculated from the kinetic and equilibrium constants indicated in Scheme 1A and 1B (Schanstra & Janssen, 1996a).

Numerical simulation of Scheme 2, which is a simplified version of Scheme 1, was used to derive rate constants for substrate conversion. The steady-state kinetic parameters under initial velocity steady-state conditions for Scheme 2 are

\[
\frac{k_{cat}}{K_m} = \frac{k_1 k_2}{k_{cat} (k_{-1} + k_2)}
\]

which were derived using the method described by Huang (1979).

For clarity, the overall rate of halide release as determined from halide binding under pseudo-first-order conditions is depicted by kₐₐₙ, whereas the rate of halide release as determined with pre-steady-state analysis of substrate conversion is depicted by kₙ. The kₐₐₙ was extracted from the rate and equilibrium constants for Scheme 1A and 1B by simulation with Gepasi. The simulation was started with all enzyme present in the form of E₁,X and halide concentration zero.

**Crystallization.** Four mutants were subjected to crystallization experiments, Phe172Met, Phe172Trp, Phe172Tyr, and Phe172Val. Protein was dissolved in 5 mM phosphate buffer (pH 6.0), with 1 mM EDTA, 1 mM 2-mercaptopoethanol, and 3 mM sodium azide. Prior to the crystallization trials, an extra purification step on a Mono Q ion-exchange column was performed, since this was shown to be essential to obtain good quality crystals of the wild type enzyme (Verschueren et al., 1993a). All experiments were performed at room temperature using the vapor diffusion method in hanging drop setups. The drops consisted of 6 μL of equal amounts of reservoir and protein solution (±5 mg of protein/mL in 100 mM MES [2-(N-morpholino)ethanesulfonic acid] buffer, pH 5.8). Ammonium sulfate was used as a precipitant in the concentration range of 44–58% saturation (at 0 °C), with and without 5% (v/v) PEG (polyethylene glycol) 400 as an additive, in 100 mM MES buffer, pH 5.6–6.2.

Only crystals of the Phe172Trp mutant could be grown, from 52 to 56% (v/v) saturated (at 0 °C) (NH₄)₂SO₄ in 100 mM MES buffer, pH 5.8. The crystals had the monoclinic space group P2₁, with cell parameters a = 95.7 Å, b = 73.5 Å, c = 41.6 Å, and β = 91.3°. These are just slightly different from the conditions and parameters for the wild type orthorhombic crystal form (Rozeboom et al., 1988). The asymmetric unit contains two molecules.

**Diffraction Data Collection.** A data-set was collected from one single crystal at room temperature on an Enraf Nonius FAST area detector system (Enraf Nonius, Delft, The Netherlands). The detector was equipped with a CAD4 kgonostat with graphite monochromatized Cu Kα radiation (λ = 1.5418 Å) from an Elliot GX21 rotating anode X-ray generator. Data were collected and processed using the MADNES package (Messerschmidt & Pflugrath, 1987). Profile fitting and local scaling of the data was done according to Kabsch (1988) and afterward merged with the in-house BIOMOL crystallographic package. Further information on the data processing is given in Table 1.
Molecular Replacement and Crystallographic Refinement. The structure was solved by molecular replacement. The structure of the wild type haloalkane dehalogenase at pH 6.2 (Verschueren et al., 1993a) was used as a search model with Phe172 replaced by Ala. Self-rotation and cross-rotation functions were calculated according to Crowther (1972) with data ranging from 10 to 3.0 Å. The translation function (6.0–3.0 Å data) used was that according to Crowther and Blow (1967). The relative position of the second molecule with respect to the first molecule was found with BRUTE (Fujinaga & Read, 1987).

The model was refined with X-PLOR version 3.1 (Brünger, 1992a). A random set of five percent of the unique reflections was set apart to calculate Rmerge values (Brünger, 1992b) as an independent validation of the refinement progress. First, a rigid body refinement was done to position the two independent molecules more precisely in the asymmetric unit. In all subsequent steps, strong non-crystallographic symmetry restraints were imposed on the two molecules and only strong reflections [F ≥ 2σ(F)] were used. Then two rounds of simulated annealing (4000–300 K) plus positional refinement were applied trying several values for the relative weight of the X-ray term (8.0–3.0 Å data). This cycle was repeated once with data to the resolution maximum. An OMIT Fo – Fc electron density map (Bhat, 1988) calculated at this stage clearly showed density for the side chain of residues 172 in both molecules. The side chains of residues 172 were then extended to tryptophans, followed by two cycles of alternating positional and restrained individual B-factor refinement. Another cycle of positional and B-factor refinement was used to make some final corrections. At all stages OMIT 2Fo – Fc and Fo – Fc electron density maps were calculated and inspected with O (Jones et al., 1991) to check the agreement of the model with the data. PROCHECK (Laskowski et al., 1993) was used to assess the stereochemical quality. Whenever necessary the model was manually adjusted with O. No water molecules were included. When the refinement gave no further decrease in Rfree nor any improvement in stereochemistry, it was stopped.

**RESULTS**

*Mutants of Phenylalanine 172.* Using random site-directed mutagenesis, 15 different mutants of Phe172 were obtained (Table 2). The mutants were constructed in two mutation rounds. In the first round, inactive mutants were obtained by Kunkel mutagenesis using oligonucleotide pF172X and wild type haloalkane dehalogenase as single-stranded DNA-template. Screening for inactive mutants was done by incubating *E. coli* BL21(DE3) grown on LBi plates in 1,2-dibromoethane vapour. LBi plates are LB plates that contain bromothymol blue as a pH indicator that gives yellow instead of green/blue colonies upon production of HCl or HBr by the active dehalogenase (Schanstra et al., 1993). Inactive colonies were analyzed for the occurrence of the additional PsI restriction site that was present in the mutant oligonucleotide. In the second mutation round, mutant DNA-template (Phe172Gly) obtained in the previous mutation round and encoding an inactive haloalkane dehalogenase was used to obtain mutants with activity for 1,2-dibromoethane. Screening was again done on LBi plates in *E. coli* BL21(DE3).

Only mutant enzymes in which Phe172 was substituted by other aromatic amino acids retained an activity for 1,2-dichloroethane which was significantly above the detection limit of the assay. Furthermore, only substitution of Phe172 by Trp, Tyr, His, and Met yielded an enzyme that was active with 1,2-dibromoethane. Mutation to Trp and Tyr resulted in an increased catalytic activity with 1,2-dibromoethane. A Phe172Cys dehalogenase was constructed to see whether this mutant also retained activity with 1,2-dibromoethane. This
equilibrium constants in Table 4.

The alkyl
2-Chloroethanol production (O
(0.25
-enzyme. There is a 2 H 2 O solvent kinetic isotope effect on both mutants, resulting in a lower k
-1,2-dichloroethane and 1,2-dibromoethane decreased in ethane was decreased in both mutants, however. The affinity
-1,2-dichloroethane and 650 µ
(250 µM). 2-Bromoethanol production (A) Rapid-quench experiment with 10 mM 1,2-dibromoethane and 500 µM enzyme. The steady-state rate of alcohol production was 6 ± 0.3 s
, as determined from the slope of the plot. (B) Rapid-quench-flow experiment with enzyme (500 µM) in excess over 1,2-dibromoethane (250 µM). 2-Bromoethanol production (C) and 1,2-dibromoethane consumption (●) in time. (C) Rapid-quench experiment with 40 mM 1,2-dichloroethane and 650 µM enzyme. The rate of steady-state alcohol production was 2.8 ± 0.4 s
 as determined from the last part (0.25–2 s) of the graph. (D) Rapid-quench-flow experiment with enzyme (770 µM) in excess over 1,2-dichloroethane (750 µM). 2-Chloroethanol production (○) and 1,2-dichloroethane consumption (●). In B and D, the dashed lines are the simulated concentrations of the alkyl–enzyme in time and the solid lines indicate the best fits to the data obtained by simulation of Scheme 2 using the rate and equilibrium constants in Table 4.

Phe172Cys enzyme, constructed by Kunkel mutagenesis using the Phe172Gly template, indeed showed activity with 1,2-dibromoethane (0.54 units/mg of protein in crude extract) and no detectable activity with 1,2-dichloroethane.

Steady-State Kinetic Parameters of Phe172Trp and Phe172Tyr Haloalkane Dehalogenases. The two mutants that displayed the highest activities in crude extracts, Phe172Trp and Phe172Tyr, were characterized in more detail (Table 3). The k
 for 1,2-dibromoethane was increased 1.8 and 2-fold over the wild type for the Phe172Tyr and Phe172Trp enzyme, respectively. The k
 for 1,2-dichloroethane was decreased in both mutants, however. The affinity for 1,2-dichloroethane and 1,2-dibromoethane decreased in both mutants, resulting in a lower k
/K
 than the wild type enzyme. There is a 2 H 2 O solvent kinetic isotope effect on the k
 of 1,2-dichloroethane conversion in the wild type enzyme. This originates both from an effect on halide release (Schanstra & Janssen, 1996a) and from an effect on hydrolysis of the alkyl–enzyme intermediate (Schanstra et al., 1996b). This 2 H 2 O kinetic isotope effect was not observed with the two mutant enzymes, as shown by k
/k
 values of 0.9 and 1 (Table 3). This indicated that in the mutants the rate-limiting step had shifted to a step before hydrolysis of the alkyl–enzyme intermediate with this substrate. The slow step in conversion of 1,2-dibromoethane was most likely still located after formation of the covalent intermediate since the k
/k
 was 0.4 and 0.5 for the Phe172Tyr and Phe172Trp enzyme, respectively, which was similar to the value found in the wild type (Table 3). The k
 for conversion of 1-bromo-2-chloroethane, which was exclusively converted to 2-chloroethanol, was also elevated to rates close to the k
 for 1,2-dibromoethane conversion.

The mutant enzymes showed a notable increase in the specificity constant (k
/K
) with 1-chlorohexane to a value which was 10-fold higher for the Phe172Trp mutant and 5-fold higher for the Phe172Tyr mutant than for the wild type enzyme. The increase in k
/K
 was caused both by a higher k
 and a lower K
. For 1-chlorohexane the main

Table 3: Kinetic Properties of Purified Wild Type and Phe172Trp and Phe172Tyr Haloalkane Dehalogenase

<table>
<thead>
<tr>
<th>substrate</th>
<th>property</th>
<th>wild type</th>
<th>Phe172Trp</th>
<th>Phe172Tyr</th>
</tr>
</thead>
</table>
| 1,2-dibromoethane             | K
 | 0.010     | 0.025     | 0.012     |
|                               | k
 | 3.0       | 5.9       | 5.3       |
|                               | k
/k
 | 3.0 × 10
 | 2.4 × 10
 | 4.4 × 10
 |
| 1,2-dichloroethane            | K
 | 0.53      | 5.13      | 4.96      |
|                               | k
 | 3.3       | 2.9       | 1.5       |
|                               | k
/k
 | 6.2 × 10
 | 5.6 × 10
 | 3.0 × 10
 |
| 1-bromo-2-chloroethane        | K
 | <0.07\textsuperscript{a} | 0.1       | 0.1       |
|                               | k
 | 3.6       | 5.5       | 4.8       |
|                               | k
/k
 | 5.5 × 10
 | 4.8 × 10
 | 4.8 × 10
 |
| 1-chlorohexane                | K
 | 1.4       | 0.57      | 0.90      |
|                               | k
 | 0.088     | 0.38      | 0.31      |
|                               | k
/k
 | 6.7 × 10
 | 3.4 × 10
 | 3.4 × 10
 |
\textsuperscript{a}Lower values could not be measured due to the detection limit of 2-chloroethanol by gas chromatography.

FIGURE 2: Pre-steady-state kinetic analysis of 1,2-dibromoethane and 1,2-dichloroethane conversion by Phe172Trp haloalkane dehalogenase. (A) Rapid-quench experiment with 10 mM 1,2-dibromoethane and 500 µM enzyme. The steady-state rate of alcohol production was 6 ± 0.3 s
, as determined from the slope of the plot. (B) Rapid-quench-flow experiment with enzyme (500 µM) in excess over 1,2-dibromoethane (250 µM). 2-Bromoethanol production (○) and 1,2-dibromoethane consumption (●) in time. (C) Rapid-quench experiment with 40 mM 1,2-dichloroethane and 650 µM enzyme. The rate of steady-state alcohol production was 2.8 ± 0.4 s
 as determined from the last part (0.25–2 s) of the graph. (D) Rapid-quench-flow experiment with enzyme (770 µM) in excess over 1,2-dichloroethane (750 µM). 2-Chloroethanol production (○) and 1,2-dichloroethane consumption (●). In B and D, the dashed lines are the simulated concentrations of the alkyl–enzyme in time and the solid lines indicate the best fits to the data obtained by simulation of Scheme 2 using the rate and equilibrium constants in Table 4.
rate-limiting step occurs before hydrolysis of the alkyl–enzyme intermediate (Schanstra et al., 1996b).

**Pre-Steady-State Kinetic Analysis of 1,2-Dibromoethane Conversion.** Substrate conversion by the Phe172Trp mutant was studied with pre-steady-state kinetic techniques to establish which steps in the catalytic cycle determined the altered kinetics of this mutant. A rapid-quench-flow experiment with 1,2-dibromoethane in excess over enzyme showed a nearly linear production of 2-bromoethanol in time (Figure 2A). This indicates that the rate of a step before hydrolysis of the alkyl–enzyme intermediate or the hydrolysis step itself was close to the steady-state $k_{\text{cat}}$ (Schanstra et al., 1996b). Formation of the alkyl–enzyme intermediate was faster than its breakdown since the substrate decrease and product increase curves crossed at about 60 $\mu$M in a single-turnover experiment with 250 $\mu$M 1,2-dibromoethane and 500 $\mu$M enzyme (Figure 2B). This indicates that alkyl–enzyme accumulated and that the slowest step in the catalytic mechanism must be hydrolysis of the alkyl–enzyme intermediate.

An estimate for the rate of 1,2-dibromoethane binding ($k_1$) was obtained from numerical simulation of the initial 20 ms of a stopped-flow fluorescence transients obtained from a single-turnover experiment (15 $\mu$M enzyme and 15 $\mu$M 1,2-dibromoethane) and of the initial 50 ms of the transient from an experiment in which enzyme (15 $\mu$M) was mixed with 100 $\mu$M 1,2-dibromoethane (not shown). This first part of the transient is related to substrate binding (Schanstra et al., 1996b). For the simulations, the relative fluorescence of the intermediates for the quenched Phe172Trp enzyme were set 13% lower than those of the wild type enzyme (Schanstra & Janssen, 1996a), based on the assumption that the fluorescence of the additional tryptophan was also quenched. The combined experiments resulted in a $k_1$ of 0.4 ± 0.1 $\mu$M$^{-1}$ s$^{-1}$. A similar value for $k_1$ (0.3 $\mu$M$^{-1}$ s$^{-1}$) was obtained from analysis of the observed rate of 1,2-dibromoethane binding (initial 50 ms) under pseudo-first-order conditions (not shown). The rate of substrate dissociation ($k_{-1}$) was set at the beginning of the simulation and fitting procedure to a value that was similar to or larger than that of the wild type enzyme (≥20 s$^{-1}$). This value was fine-tuned during the fitting procedure with the experimentally determined steady-state kinetic parameters as additional constraints.

Combining these data with the rapid-quench-flow data resulted in rates that can explain the kinetics of this mutant with 1,2-dibromoethane (Table 4, Scheme 2, Figure 2A and B). The slowest step in the sequence was found to be hydrolysis of the alkyl–enzyme, which had a rate constant of 9 $s^{-1}$ and could explain the effect of $H_2O$ found on the steady-state $k_{\text{cat}}$ (Table 3). The most important conclusion was that the rate of bromide release ($k_X$) was clearly higher with the Phe172Trp enzyme than with the wild type enzyme. The increase in the steady-state $k_{\text{cat}}$ with 1,2-dibromoethane of the Phe172Trp enzyme was mainly caused by this elevated rate of halide release, which could be set at values between 50 and 100 s$^{-1}$ without affecting the fit. The rate of carbon–bromine bond cleavage was reduced 4-fold in the Phe172Trp enzyme.

**Bromide Release.** The increased rate of bromide release with the Phe172Trp enzyme was confirmed by stopped-flow fluorescence experiments of bromide binding under pseudo-first-order conditions. All stopped-flow fluorescence transients observed from mixing Phe172Trp enzyme (5 $\mu$M) with 3 to 750 mM KBr could be fitted by single exponentials (Figure 3A). The resulting dependence of the observed bromide binding rate ($k_{\text{obs}}$) on the bromide concentration showed that this rate decreased from 48 s$^{-1}$ at 3 mM to around 31 s$^{-1}$ at 30 mM KBr, followed by a hyperbolic increase of the $k_{\text{obs}}$ at higher bromide concentrations (Figure 3B). The same dependence was found when the ionic

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**Table 4: Phe172Trp and Wild Type Haloalkane Dehalogenase Kinetic Constants** (Fits in Figure 2)

<table>
<thead>
<tr>
<th></th>
<th>$k_1$ ($\mu$M$^{-1}$ s$^{-1}$)</th>
<th>$k_{-1}$ (s$^{-1}$)</th>
<th>$k_2$ (s$^{-1}$)</th>
<th>$k_3$ (s$^{-1}$)</th>
<th>$k_X$ (s$^{-1}$)</th>
<th>$k_{\text{cat(calc)}}$ (s$^{-1}$)</th>
<th>$K_m$ (calc) ($\mu$M)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1,2-dibromoethane</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Wild type</td>
<td>0.75 ± 0.1</td>
<td>&gt;20</td>
<td>&gt;130</td>
<td>10 ± 2</td>
<td>4 ± 1.5</td>
<td>2.8</td>
<td>4.3</td>
</tr>
<tr>
<td>Phe172Trp</td>
<td>0.4 ± 0.1</td>
<td>25 ± 5</td>
<td>30 ± 5</td>
<td>9 ± 1.5</td>
<td>75 ± 25</td>
<td>6.3</td>
<td>29</td>
</tr>
<tr>
<td>1,2-dichloroethane</td>
<td>(9 ± 1) $\times$ 10$^{-3}$</td>
<td>20 ± 5</td>
<td>50 ± 10</td>
<td>14 ± 3</td>
<td>8 ± 2</td>
<td>4.6</td>
<td>0.72 ± 10$^4$</td>
</tr>
<tr>
<td>Wild type</td>
<td>(3 ± 1) $\times$ 10$^{-3}$</td>
<td>30 ± 5</td>
<td>4.5 ± 1</td>
<td>9.5 ± 1</td>
<td>&gt;75</td>
<td>2.9</td>
<td>7.5 ± 10$^3$</td>
</tr>
<tr>
<td>Phe172Trp</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

* Wild-type data are from Schanstra et al. (1996b). * Steady-state kinetic parameters calculated from the obtained rate constants for a four-step reaction mechanism (see Materials and Methods).

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Conversion of 1,2-Dichloroethane. The absence of a solvent kinetic isotope effect on the steady-state rate of 1,2-dichloroethane conversion suggested that neither hydrolysis of the alkyl−enzyme intermediate nor halide release was rate-limiting in the Phe172Trp mutant (Table 3). Stopped-flow fluorescence analysis of chloride binding under pseudo-first-order conditions showed that the overall rate of chloride release ($k_{off}$) had increased to 108 s$^{-1}$, which was 7-fold faster than with the wild type enzyme (Table 5). This observation confirmed that chloride release cannot be rate-limiting in this mutant. The dependence of the $k_{obs}$ on the chloride concentration, which over the complete concentration range could be fitted with single exponentials (12.5−75 mM NaCl), was similar to that of the wild type enzyme (Scheme 1B, Figure 3C).

Further information about the rates of the separate steps of 1,2-dichloroethane conversion was obtained from rapid-quench-flow experiments. An experiment with substrate in excess over enzyme showed a clear lag in 2-chloroethanol formation (Figure 2C). Thus, a step before hydrolysis of the alkyl−enzyme or hydrolysis itself was rate-limiting. In an experiment with substrate in excess over enzyme, one cannot distinguish the rates of $k_2$ and $k_3$. A single-turnover experiment with enzyme (770 µM) in excess over substrate (750 µM) showed that the substrate decrease and product increase curves crossed at 370 µM (Figure 2D), indicating that hydrolysis of the alkyl−enzyme ($k_3$, Scheme 2) was at least as fast as its formation. Extraction of rate constants for the reaction needed, like with 1,2-dibromoethane conversion, additional input from stopped-flow fluorescence experiments of substrate conversion (not shown). Again, an estimate of the rate of substrate binding ($k_1$) could be extracted from the initial part stopped-flow fluorescence transients (3 ± 1 mM$^{-1}$ s$^{-1}$).

Using numerical simulation and fitting, a complete set of rate constants could be derived for 1,2-dichloroethane conversion by the Phe172Trp mutant (Table 4). The slowest step in the sequence was cleavage of the carbon-halogen bond, which was 4.5 s$^{-1}$. The rate of halide release ($k_3$) could be fitted with values larger than 75 s$^{-1}$, which was in agreement with the rates found in chloride binding experiments under pseudo-first-order conditions. The fact that carbon−halogen bond cleavage had become rate-limiting can explain the loss of the solvent kinetic isotope effect on 1,2-dichloroethane conversion and the decrease in steady-state $k_{cat}$, although the rate of chloride release, which was rate-limiting in wild type enzyme, had increased.

Structure Determination: Molecular Replacement and Crystallographic Refinement. To establish the structural

<table>
<thead>
<tr>
<th>X</th>
<th>$K_{d}$ (mM)</th>
<th>$k_{cat}$ (s$^{-1}$)</th>
<th>$k_{cat}^{E_{0.5}}$ (mM$^{-1}$ s$^{-1}$)</th>
</tr>
</thead>
<tbody>
<tr>
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<td>10.0</td>
<td>10.0</td>
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<tr>
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<td>14.0</td>
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<tr>
<td>Phe172Trp</td>
<td>14.0</td>
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Wild-type values were taken from Schanstra and Janssen (1996a). Apparent $K_d$ obtained from steady-state fluorescence measurements. Calculated apparent steady-state $k_{cat}$, from obtained rate and equilibrium constants when assuming that the equilibrium $E_{0.5}^X$ and $E_{0.5}^X$−$E_{0.5}^X$ or scheme used in fit.
basis for the observed kinetic effects we determined the X-ray structure of the Phe172Trp dehalogenase. A self-rotation function showed two peaks: the origin peak and a peak at 74% height of the origin, representing a non-crystallographic 2-fold (screw) axis in the a,c-plane, 1.3° off the a-axis. The position of the axis corresponds to the position of a 21-axis in the wild type crystals. A cross-rotation function yielded δ respectively. In molecule A, Asp184O η bonded to N-ø structural reason was apparent, however. Asp184 had only five contacts. Nevertheless, the orientation of the molecules in the mutant crystal lattice and the total number of intermolecular contacts were very similar to the wild type, although some detailed interactions differed. Both monomers A and B had lost the interactions between Asn14 and symmetry-related Gln167 and between Glu41 and symmetry-related Arg193. Molecule A showed new contacts between Arg112 and a related Glu228 and from Asp212 to Lys304 and Glu308. Molecule B displayed new or extra crystal contacts between Pro19 and Tyr177, Phe20 and Tyr177, Asn43 and Arg186, and Glu72 and Ser183. These changed intermolecular contacts have been caused by the conformational change in the 165–166 region (see below).

The three-dimensional structure of the mutant was very similar to the wild type structure at pH 6.2 (Verschueren et al., 1993a). The catalytic triad, formed by residues Asp124, His289, and Asp260, was intact, and the amino acids lining the active site cavity were also in an identical orientation (Figure 4). The cavity volume as calculated with VOIDOO (Kleywegt & Jones, 1994) had not changed significantly. The tryptophan side chain introduced at position 172 was located in the same plane as the phenylalanine in wild type, with the six-membered aromatic rings in an identical position. The bigger size of the Trp172 side chain has caused the main chain of residues Val165 and Thr166 to move outward by about 1.1 Å for the Cα atoms (Figure 4). The hydrogen bond between Thr173Oy1 and Val165O was hereby disrupted in the mutant (interatomic distance 4.7 Å vs 3.3 Å in the wild type). Due to this outward displacement, the adjacent Phe164 has also moved from its position in the wild type enzyme by about 0.5 Å, thus weakening the quadrupole–quadrupole interaction between the aromatic rings at positions 164 and 172.

**DISCUSSION**

Phenylalanine 172 of haloalkane dehalogenase interacts with the Cl β and Cα of 1,2-dichloroethane in the Michaelis complex and alkyl–enzyme during catalysis (Verschueren et al., 1993b), and therefore it might affect the mode of binding and reaction rate of this substrate. Furthermore, inspection of the structure indicates that Phe172 may be involved in stabilization of the N-terminal helix—loop—helix region between residue 159 and 181 of the cap domain by the stabilizing interactions between Phe172 and Phe164. To obtain information about the role of phenylalanine 172 in haloalkane dehalogenase structure and function, we performed a mutational analysis of position 172, and studied the X-ray structure and steady-state and pre-steady-state kinetics of the Phe172Trp enzyme.
Inspection of the X-ray structure of the Phe172Trp dehalogenase showed that the two tryptophans that make up the halide binding site of the enzyme (Verschueren et al., 1993c) were at the same positions in the Phe172Trp mutant as in the wild type enzyme (Figure 4). The bigger size of the Trp side chain compared to wild type Phe had forced the main chain atoms of residues Val165 and Thr166 to move outward (Figure 4), causing the loss of the Thr173Oγ–Val165O hydrogen bond. Because of the displacement of the main chain at positions 165 and 166, the packing interactions made in wild type by the adjacent residues Gln167 and Pro168 are not present in the mutant crystals. This change has caused the mutant enzyme to crystallize in a different space group than the wild type, which forms crystals in space group P2₁2₁2₂ exclusively.

The catalytic properties of the Phe172Trp enzyme were significantly different from those of the wild type enzyme. The $k_{cat}$ for 1,2-dibromoethane was about 2-fold higher for the Phe172Trp dehalogenase than for the wild type enzyme. Pre-steady-state analysis showed that this could be attributed to the increased rate of bromide release. The rate of 1,2-dibromoethane binding and C–Br bond cleavage was lower in the Phe172Trp enzyme than in the wild type enzyme, however.

The helix–loop–helix region of the cap domain of the dehalogenase contributes to the hydrophobic substrate binding site of the enzyme (Verschueren et al., 1993a,b) and it is likely that the observed conformational difference might influence the binding energy of smaller substrates such as 1,2-dichloroethane and 1,2-dibromoethane. Furthermore, the lower rate of carbon–halogen bond cleavage found in the Phe172Trp enzyme indicates that this part of the cap could also be involved in transition state stabilization during carbon–halogen bond cleavage.

For the longer substrate 1-chlorohexane, however, this structural change in the mutant seems to be favorable. 1-Chlorohexane is a poor substrate for the wild type dehalogenase (Pries et al., 1994b), and the rate-limiting step for conversion of this substrate is before hydrolysis of the alkyl–enzyme intermediate (Schanstra et al., 1996b). How this substrate is bound is unknown since the active site cavity of both the wild type dehalogenase (Verschueren et al., 1993b) and the Phe172Trp enzyme is too small to bind 1-chlorohexane, but it is conceivable that a more flexible hydrophobic active site cavity in the Phe172Trp enzyme might facilitate a conformational change to accommodate a larger substrate. Dehalogenase mutants with improved conversion of 1-chlorohexane were also obtained using in vivo selection (Pries et al., 1994b). No X-ray structures have been obtained for these mutants so far, but most of the changes in these mutants were located in the helix–loop–helix region of the cap domain, indicating similar changes in the active sites of these mutants as in the Phe172Trp dehalogenase.

The fact that only the Phe172Trp and the Phe172Tyr enzymes retained their activity with 1,2-dichloroethane could indicate that besides a more nonspecific hydrophobic interaction, an aromatic (172)–chlorine interaction contributes to substrate binding and transition state stabilization during C–Cl bond cleavage. This is supported by the observation that the alkyl–enzyme of the wild type dehalogenase the Cl$_2$ of the 1,2-dichloroethane lies in the plane of the positively charged edge (C$_{11}$ and C$_{31}$) of Phe172 (Verschueren et al., 1993b). The interaction of negatively charged groups with aromatic rings has also been observed with sulfur–aromatic (Reid et al., 1985; Burley & Petsko, 1988) and oxygen-aromatic interactions (Thomas et al., 1982; Burley & Petsko, 1988), where the negatively charged oxygen or sulfur exhibits affinity toward the positively charged edge of the aromatic ring.

An interaction between the enzyme and the alkyl group of the substrate seems to be less important for the dehalogenase reaction with 1,2-dibromoethane, since the Phe172Met, the Phe172Cys, and the Phe172His enzymes which were inactive with 1,2-dichloroethane had a significant activity with 1,2-dibromoethane. The difference between 1,2-dichloroethane and 1,2-dibromoethane conversion can originate from the fact that bromine is a better leaving group in $S_N$2 type reactions. Furthermore, the larger van der Waals volume of 1,2-dibromoethane compared to 1,2-dichloroethane could also lead to a more “tight” fit of this substrate in the active site cavity.

Both for 1,2-dichloroethane and 1,2-dibromoethane conversion, the rate of hydrolysis of the alkyl–enzyme intermediate in the Phe172Trp enzyme did not change significantly compared to the wild type enzyme. This is not surprising, since residues involved in this step, i.e., the Asp260–His289 couple which activates the nucleophilic water molecule and residues that form the oxyanion hole (Verschueren et al., 1993b; Pries et al., 1995a), are located in the main domain of the dehalogenase and were at nearly identical positions in the Phe172Trp and wild type enzyme.

A possible explanation for the lack of activity displayed by the other mutants in Table 2 is that the structural integrity of the active site is lost by their mutations. There was, for example, a very low affinity for bromide and chloride in the Phe172Val enzyme (J.P. Schanstra, unpublished results), while the apparent affinity constant ($K_a$) increased only 2-fold for the Phe172Trp enzyme (Table 5). The origin of this may be located in the loss or weakening of the interaction between residues 164 (helix 4) and 172 (helix 5) upon mutation of residue 172. In the wild type enzyme, the positively charged edge of Phe164 approaches the negative face of Phe172 (ring centroids are 5.8 Å apart and oriented perpendicular). These two residues might serve as an hydrophobic anchor for the helix–loop–helix structure, and such aromatic interactions in the interior of proteins are known to contribute to protein stability (Burley & Petsko, 1985, 1988; Hunter et al., 1991; Anderson et al., 1993). In the Phe172Trp enzyme, an interaction between these two residues is still present, and also in the Phe172Tyr, Phe172Met, Phe172Cys, and Phe172His enzymes an interaction between position 172 and 164 can be envisaged. It is known that sulfur-containing residues can interact with aromatic residues (Reid et al., 1985; Burley & Petsko, 1988). Histidine is also capable of interacting favorably with aromatic rings (Loewenthal et al., 1992; Armstrong et al., 1993; Jasanoff & Weiss, 1993). The other amino acids introduced at position 172 could either be not capable of a specific interaction with aromatic residues such as Phe164 (Gly/Ala/Val/Leu/Ile) or are likely to be too hydrophilic (Glu/Asn/Ser/Thr), disturbing the hydrophobic character of the active site cavity.

Both in the wild type enzyme (Schanstra & Janssen, 1996a) and in the Phe172Trp enzyme, we have found two routes for release of the charged halide ion out of the buried active site cavity (Scheme 1A and 1B). For the route which
is kinetically most important, actual release was preceded by a slow enzyme isomerization. Pre-steady-state kinetic analysis of halide binding under pseudo-first-order conditions showed that the overall increase in the rate of halide release in the Phe172Trp enzyme was caused by an increased rate of this unimolecular enzyme isomerization. For the wild type enzyme we have hypothesized that this enzyme isomerization could be a conformational change which is necessary to allow water to enter the active site and solvate the halide ion (Schanstra & Janssen, 1996a). The X-ray structure of the Phe172Trp enzyme showed that the two tryptophan residues between which the halogen moiety of the substrate and halide is bound were at identical positions in mutant and wild type enzyme (Figure 4), indicating that altered positions of these tryptophans were not responsible for the large difference in rate of halide release. However, the structural changes in the helix-loop-helix region that covers the active site of the dehalogenase and the observation that the rate of halide release was increased dramatically in the Phe172Trp enzyme indicates that the proposed slow conformational change necessary for solvation of the halide is located in this region. Other mutations that influence the stability of the helix-loop-helix region without affecting active site residues should indicate whether this is indeed the site of the conformational change that was identified by kinetic measurements.

REFERENCES


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