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Common Variants in the Type 2 Diabetes KCNQ1 Gene Are Associated with Impairments in Insulin Secretion During Hyperglycaemic Glucose Clamp


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Abstract

Background: Genome-wide association studies in Japanese populations recently identified common variants in the KCNQ1 gene to be associated with type 2 diabetes. We examined the association of these variants within KCNQ1 with type 2 diabetes in a Dutch population, investigated their effects on insulin secretion and metabolic traits and on the risk of developing complications in type 2 diabetes patients.

Methodology: The KCNQ1 variants rs151290, rs2237892, and rs2237895 were genotyped in a total of 4620 type 2 diabetes patients and 5285 healthy controls from the Netherlands. Data on macrovascular complications, nephropathy and retinopathy were available in a subset of diabetic patients. Association between genotype and insulin secretion/action was assessed in the additional sample of 335 individuals who underwent a hyperglycaemic clamp.

Principal Findings: We found that all the genotyped KCNQ1 variants were significantly associated with type 2 diabetes in our Dutch population, and the association of rs151290 was the strongest (OR 1.20, 95% CI 1.07–1.35, p = 0.002). The risk C-allele of rs151290 was nominated associated with reduced first-phase glucose-stimulated insulin secretion, while the non-risk T-allele of rs2237892 was significantly correlated with increased second-phase glucose-stimulated insulin secretion (p = 0.025 and 0.0016, respectively). In addition, the risk C-allele of rs2237892 was associated with higher LDL and total cholesterol levels (p = 0.015 and 0.003, respectively). We found no evidence for an association of KCNQ1 with diabetic complications.

Conclusions: Common variants in the KCNQ1 gene are associated with type 2 diabetes in a Dutch population, which can be explained at least in part by an effect on insulin secretion. Furthermore, our data suggest that KCNQ1 is also associated with lipid metabolism.
Funding: This study was funded by SenterNovem (an IOP Genomics IGE05012 from Agentschap NL), and by the Dutch Diabetes Research Foundation (2006.00.060). In addition, the UDES study was supported by an unrestricted research grant from Novo Nordisk BV (Alphen aan de Rijn, the Netherlands), and the Scientific Institute of Dutch Pharmacists (WINAp, The Hague, the Netherlands). The EPIC-NL study was funded by the Dutch Ministry of Public Health, Welfare and Sports (VWS), the Netherlands Cancer Registry (NKI), the LK Research Funds, the Dutch Prevention Funds, the Dutch ZON (Zorg Onderzoek Nederland), and the World Cancer Research Fund (WCRF) (The Netherlands). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: Dr. O. H. Klungel has been consultant to Sanofi-Aventis on issues not related to this paper (2006.00.060). The UDES study was supported by an unrestricted research grant from Novo Nordisk BV (Alphen aan de Rijn, the Netherlands). This does not alter the authors’ adherence to all the PLoS ONE policies on sharing data and materials. The authors have declared that no competing interests exist.

Introduction

Recent genome-wide association (GWA) studies have provided a first significant insight into the genetic architecture of type 2 diabetes, and to date, around 40 loci have been identified to be robustly associated with the disease [1]. So far, the majority of GWA scans have been performed in populations of European descent [1]. The first GWA studies in East Asians have recently identified single nucleotide polymorphisms (SNPs) in a previously unreported gene, KCNQ1, which were associated with type 2 diabetes susceptibility [2,3]. The original studies also confirmed this finding in populations of European descent [2,3].

It is recognized, that in the pathophysiology of type 2 diabetes both disturbances in insulin action (liver and muscle) and in insulin secretion are early events [4] with additional factors such as age itself [5]. Therefore, it is important to examine the association between the KCNQ1 variants and metabolic traits to elucidate the underlying diabetes-causing mechanisms.

In this study we aimed to investigate (1) the relationship of specific KCNQ1 gene SNPs in the pathophysiology of type 2 diabetes by examining their association with metabolic traits and insulin secretion during hyperglycemic clamps, and (2) whether these SNPs relate to the risk of developing diabetes complications and to the risk of mortality among type 2 diabetes patients of Dutch origin.

Materials and Methods

Type 2 diabetes case-control sample description

We included 4,620 type 2 diabetes patients and 5,285 healthy controls of Dutch Caucasian origin ascertained from various study populations in the Netherlands: 1) the New Hoorn and Diabetes Care System (DCS) West-Friesland studies: 1,969 patients with type 2 diabetes and 1,951 controls with a normal glucose tolerance (n = 465); 2) the Breda study: 569 type 2 diabetes patients and 920 healthy blood bank donors (n = 9); 3) the Zwolle Outpatient Diabetes project Integrating Available Care (ZODIAC) study: 914 primary care patients with type 2 diabetes (n = 10); 4) the European Prospective Investigation Into Cancer and Nutrition-the Netherlands (EPIC-NL): 976 type 2 diabetes patients and 1,646 controls (n = 1,12); 5) the Vlagtwedde/Vlaardingen cohort: 768 controls from the general population [13]; 6) the Utrecht Diabetes Epidemiology Study (UDES) study: 192 Dutch white individuals with type 2 diabetes. The ancestry in all studies except the EPIC-NL sample was determined based on self-reported information. Detailed characteristics are shown in Table S1.

The UDES population has not been described before: it was collected from the population-based Pharmaco-Morbidity Record Linkage System (PHARMO, www.pharmo.nl) linking drug-dispensing histories from a representative sample of Dutch community pharmacies to the national register of hospital discharges (Landelijke Medische Registratie (LMR)) from 1985 onwards. A retrospective cohort study of new users of blood glucose - lowering drugs (either oral hypoglycaemic agents or insulin), who were 18 years or older, were selected, and 1,609 patients were recruited through community pharmacies participating in PHARMO. Diagnosis of type 2 diabetes was confirmed by self-reported information from the participants. We checked a small sample of 24 type 2 diabetes cases with 92% of these could be confirmed according to the World Health Organisation (WHO) criteria for diagnosing type 2 diabetes. From these 1,609 patients, 255 took part in the study, returned the questionnaire that had been sent to them, and donated blood for various assessments and DNA retrieval. Laboratory measurements included plasma total cholesterol, low-density lipoprotein (LDL) cholesterol, high-density lipoprotein (HDL) cholesterol, triglycerides, fasting blood glucose, non-fasting blood glucose, and HbA1c. Only Dutch white individuals were included in the present study (n = 192).

All patients from studies 1–4 were diagnosed according to the WHO criteria (2-hour plasma glucose levels >11.1 mmol l⁻¹ or fasting plasma glucose levels ≥7.0 mmol l⁻¹).

Data on diabetic complications and on mortality in type 2 diabetes patients

The presence of macrovascular complications (n = 264), nephropathy (n = 442), neuropathy (n = 333), and diabetic retinopathy (n = 465) was available in a subset of type 2 diabetes patients from DCS West-Friesland, the ZODIAC and the EPIC-NL studies (Table S2). A diagnosis of diabetic retinopathy in the DCS West-Friesland was based on the examination of fundus photography graded by an ophthalmologist according to the EURODIAB Study grading system as described elsewhere [8]. A diagnosis of diabetic retinopathy in ZODIAC was based on an ophthalmologist’s examination that demonstrated microaneurysms, macular edema, or pre-proliferative retinopathy. In the EPIC-NL study data on diabetic complications was collected during the ascertainment and verification of prevalent and incident diabetes cases in the Dutch contributor to the EPIC cohort. Diabetes was ascertained via self-report, linkage to registers of hospital discharge diagnoses and a urinary glucose strip test. Ascertained diabetes cases were verified against general practitioner (GP) or pharmacist information using mailed questionnaires [12]. The GP questionnaire contained 12 questions on diabetes. The questions on in what year and which type of diabetes had been diagnosed, on how the diagnosis was established, on treatment during the first year after diagnosis and current treatment (diet, oral glucose lowering medication, insulin), and whether the patient suffered from long-term complications were asked. When the GP was unknown, pharmacist information was used to verify the diagnosis of diabetes. The pharmacist questionnaire contained eight questions concerning use of diabetes medication [12].

Data from the ZODIAC study on mortality were collected after a follow-up period of 10 years in a prospective Zwolle cohort of
type 2 diabetes patients by retrieving life status and cause of death from records maintained by the hospital and the GPs [10].

**Hyperglycaemic clamp study**

Participants from three independent studies from the Netherlands were used: 138 impaired glucose tolerance (IGT) subjects from the Hoorn study, 74 subjects (63 normal glucose tolerance (NGT)/11 IGT) from the Utrecht clamp cohort and 123 twins and sibs (116 NGT/7 IGT) from the Netherlands Twin Register (NTR) [14,15]. The latter cohort is a family based twin study, which included 66 monozygotic, and 28 dizygotic twins as well as 29 of their nontwin sibs recruited from 50 families [15]. The clinical characteristics of the study groups are given in Table S3. Details of these study groups and the clamp procedure have previously been described elsewhere [14,15].

All participants (n = 335) underwent a hyperglycaemic clamp at 10 mmol/l glucose for at least 2 h. First-phase insulin secretion was determined as the sum of the insulin levels during the first 10 min of the clamp. Second-phase insulin secretion was determined as the mean of the insulin levels during the last 40 min of the second hour of the clamp (80–120 min). The insulin sensitivity index (SI) was calculated by relating the glucose infusion rate (M) to the plasma insulin concentration (I) during the last 40 min of the second hour of the clamp (M/I). The disposition index (DI) was calculated by multiplication of first-phase insulin secretion and SI in order to quantify insulin secretion in relation to the ambient insulin sensitivity [15].

All study protocols were approved by local institutional review boards or hospital medical ethics committees (the New Hoorn and DCS West-Friesland studies were approved by the medical ethics committee of the VU University Medical Center Amsterdam; the Breda, EPIC-NL, and UDES studies were approved by the Medical Ethics Committee of the University Medical Center Utrecht; the ZODIAC study was approved by the local Medical Ethical Committee of the Isala Clinics; the Vlagtwedde/Vlaardingen study was approved by the Medical Ethics Committee of the University Medical Center Groningen, the ‘clamp studies’ were approved by the medical ethics committees of the VU University Medical Center Amsterdam and the University Medical Center Utrecht). All participants gave their written informed consent.

**Genotyping**

Based on the original GWA scans and the replication studies in Europeans, we selected gene variants with a minor allele frequency (MAF) >5% - rs151290, rs2237892, and rs2237895 in the KCNQ1, which were reported to be strongly associated with type 2 diabetes in European population [2,3,16,17,18]. The two SNPs - rs2237895 and rs2237892 showed the strongest association with T2D and were replicated in European population in the original studies by Unoki et al. and Yasuda et al., respectively [2,3]. The variant rs151290 showed strong association in the third screening in the original study by Yasuda et al. [3], and was also reported to be associated with insulin secretion in a German population by Mussig et al. [18].

These variants were genotyped in all the samples except the EPIC-NL study samples using Taqman assays (Applied Biosystems, Applied Biosystems, Nieuwerkerk a/d IJssel, The Netherlands) and were analyzed using a TaqMan 7900HT (Applied Biosystems). The DNA samples were processed in 384-well plates. Each plate contained 16 genotyping controls (4 duplicates of 4 different Centre d’Etude du Polymorphisme Humain (CEPH) samples). There were no discordances in the genotypes of any of the CEPH samples and the CEU data available from HapMap. The genotype success rate was 95.6%, 97.6% and 98.2% for rs151290, rs2237892, and rs2237895, respectively. For the individuals from the EPIC-NL study, the genotypes data for the two SNPs - rs2237892 and rs2237895, were available. The EPIC-NL samples were genotyped using the Illumina IBC v.3 array (also referred to as the CardioChip or the Human Cardiovascular Disease [HumanCVD] BeadChip [Illumina] [19]). The genotyping information was not available for rs151290 because this SNP was not included on the CardioChip. The rs2237892 and rs2237895 SNPs were clustered into genotypes with the use of the Illumina Beadstudio software and were subjected to quality control filters at the sample (i.e. only samples with call rate >95% and only not related and individuals of European ancestry were included) and SNP levels (i.e. SNPs with a call rate <95% or HWE p<10^-6 were removed).

**Statistical analysis**

The genotype frequencies were tested for Hardy-Weinberg equilibrium (HWE) by χ² analysis. In the genotyped samples from the EPIC-NL study, p-hat, a measure of identity by descent, was calculated to exclude cryptic relatedness and duplicate samples (p-hat>0.2) via the method implemented in PLINK [20]: EIGEN-STRAT method was used to compute principal component with HapMap panels as reference standards to exclude the population outliers [21].

To test for association of genotypes and type 2 diabetes and its complications, genotype-based odds ratios (OR) with 95% confidence intervals (CI) were calculated in the combined sample of type 2 diabetes patients and controls using a logistic regression model, with individuals homozygous for the non-risk allele as the reference group. The risk and non-risk alleles were defined based on the previous reports [2,3,16,17,18]. The association between genotypes and metabolic traits (BMI, HbA1c, fasting glucose, HDL- and LDL cholesterol, total cholesterol, triglyceride) was determined using linear regression analysis. This analysis was restricted to control individuals to avoid diabetes status or treatment masking potential effects of the variants on these phenotypic traits. All analyses were done under the additive model and the presented p-values are adjusted for age, sex and study center, with the continuous traits log-transformed prior to statistical comparisons. The effect of the risk alleles on the responses during hyperglycaemic clamping was examined by calculating the β values for the risk allele with linear generalized estimating equations, which takes into account the family relatedness when computing the standard errors (i.e., in the twin sample from the Netherlands Twin Register study). Only for rs2237892 the non-risk allele was used for the latter calculations since our sample did not have any homozygotes for the risk allele. All outcome variables were log-transformed prior to analysis. Analyses of hyperglycaemic clamp data were also adjusted for age, sex, BMI, study center and glucose tolerance status. A Cox proportional hazard model was used to assess association between the SNPs and total mortality with correction for age and sex.

To account for the number of independent tests, a p-value of <0.0033 (α = 0.05/15) was considered statistically significant, given independent tests for 15 outcomes (type 2 diabetes status, its complications (i.e. retinopathy, nephropathy, neuropathy), metabolic traits (i.e. BMI, HbA1c, fasting glucose, HDL, LDL, total cholesterol, triglyceride) and the parameters related with insulin secretion). However, as this level is probably too stringent as a Bonferroni correction assumes independency of the tests, which is clearly not the case in this study, p-values between 0.05 and 0.0033 were considered nominally significant. All statistical analyses were performed using the SPSS program, version 14.0 for Windows (SPSS, Chicago, IL, USA).
Table 1. Clinical characteristics of the study participants.

<table>
<thead>
<tr>
<th>Trait</th>
<th>T2D patients (n = 4620)</th>
<th>Controls (n = 5285)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>n</td>
<td>Mean ± SD</td>
</tr>
<tr>
<td>Male (%)</td>
<td>4616</td>
<td>1999 (43.3)</td>
</tr>
<tr>
<td>Age-at-study (years)</td>
<td>4617</td>
<td>64.3±10.6</td>
</tr>
<tr>
<td>Age at diagnosis (years)</td>
<td>3497</td>
<td>58.7±11.6</td>
</tr>
<tr>
<td>BMI (kg/m2)</td>
<td>4550</td>
<td>29.4±4.9</td>
</tr>
<tr>
<td>HbA1c (%)</td>
<td>4370</td>
<td>7.1±1.3</td>
</tr>
<tr>
<td>Fasting glucose</td>
<td>2496</td>
<td>8.3±2.3</td>
</tr>
<tr>
<td>HDL-cholesterol (mmol/l)</td>
<td>4187</td>
<td>1.2±0.3</td>
</tr>
<tr>
<td>LDL-cholesterol (mmol/l)</td>
<td>2746</td>
<td>2.8±0.9</td>
</tr>
<tr>
<td>Total cholesterol (mmol/l)</td>
<td>4226</td>
<td>5.1±1.2</td>
</tr>
<tr>
<td>Triglyceride (mmol/l)</td>
<td>4232</td>
<td>2.0±1.3</td>
</tr>
</tbody>
</table>


Table 2. Association of the KCNQ1 variants with type 2 diabetes in the Dutch population.

<table>
<thead>
<tr>
<th>SNP</th>
<th>Major/minor allele</th>
<th>Genotype distribution (%)</th>
<th>p-value\a,b</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>RAF</td>
<td>OR (95% CI)</td>
<td>p-value</td>
</tr>
<tr>
<td>rs151290 T2D</td>
<td>C/A</td>
<td>1.20 (1.07–1.35)</td>
<td>0.002</td>
</tr>
<tr>
<td>Control</td>
<td>0.76</td>
<td>2081 (60.2)</td>
<td>1220 (35.3)</td>
</tr>
<tr>
<td>rs2237892</td>
<td>C/T</td>
<td>1.16 (0.97–1.40)</td>
<td>0.11</td>
</tr>
<tr>
<td>T2D</td>
<td>0.96</td>
<td>4149 (92.0)</td>
<td>348 (7.7)</td>
</tr>
<tr>
<td>Control</td>
<td>0.95</td>
<td>4638 (90.0)</td>
<td>507 (9.8)</td>
</tr>
<tr>
<td>rs2237895</td>
<td>A/C</td>
<td>1.09 (1.00–1.17)</td>
<td>0.035</td>
</tr>
<tr>
<td>T2D</td>
<td>0.43</td>
<td>1522 (33.5)</td>
<td>2158 (47.4)</td>
</tr>
<tr>
<td>Control</td>
<td>0.41</td>
<td>1803 (34.8)</td>
<td>2516 (48.6)</td>
</tr>
</tbody>
</table>

\aAdjusted for age, sex and study center.
\bP-value for the additive model.

For each variant the C-allele is the risk allele for type 2 diabetes as identified by previous studies. RAF: risk allele frequency. T2D: type 2 diabetes.

doi:10.1371/journal.pone.0032148.t001

Table 1. Clinical characteristics of the study participants.

Table 2. Association of the KCNQ1 variants with type 2 diabetes in the Dutch population.

Effect of KCNQ1 polymorphisms on insulin secretion as assessed with hyperglycaemic glucose clamps

To investigate potential mechanisms by which the variants in KCNQ1 may contribute to type 2 diabetes susceptibility, we used regression analysis to examine the effects of KCNQ1 genotypes on first- and second-phase of glucose stimulated insulin secretion, insulin sensitivity index (ISI) and disposition index (DI) in a sample of non-diabetic individuals in whom hyperglycaemic clamp was performed (Table 3). We found nominal association between the C-allele of rs151290 and decreased insulin secretion during first-phase and increased ISI (p = 0.025 and p = 0.006, adjusted for age, sex, study center, BMI and glucose tolerance status, respectively) as well as nominal relationship between the C-allele of rs2237895 and higher ISI values (p = 0.034, adjusted for age, sex, study center, BMI and glucose tolerance status). In addition, we observed that carriers of the non-risk allele for rs2237892 had significantly higher second phase insulin secretion and nominally significant lower ISI compared to the homozygotes for the C-risk allele (p = 0.0016 and p = 0.036, adjusted for age, sex, study center, BMI and glucose tolerance status, respectively). None of the KCNQ1 variants had an effect on the DI in our study.

Relationship of KCNQ1 polymorphisms with metabolic parameters

Next, we investigated whether the KCNQ1 polymorphisms influenced relevant clinical parameters, such as BMI, HbA1c, fasting glucose, HDL, LDL, total cholesterol and triglycerides, in
non-diabetic individuals (Table 4). Under an additive genetic model, the diabetes risk allele of rs2237892 was nominally associated with increased LDL level and significantly associated with higher level of total cholesterol ($p = 0.015$ and $p = 0.003$, adjusted for age, sex and study center, respectively). In addition, carriers of the C-risk allele of rs2237895 had nominally higher HbA1c levels ($p = 0.036$, adjusted for age, sex and study center).

Relationship of KCNQ1 polymorphisms with diabetes complications and mortality

We further investigated whether there was an association between the KCNQ1 gene SNPs and various type 2 diabetes complications and mortality. We did not observe any association of KCNQ1 SNPs with diabetic complications, although there was a trend of the homozygous carriers of the risk C-allele of rs2237895 being more frequent among the diabetic patients with retinopathy compared to the patients without it (22.5 vs. 19.2%, $p = 0.06$, adjusted for age, sex and study center).

Data on mortality were collected after a follow-up period of 10 years in the ZODIAC study; the characteristics of the study population are shown in Table S4. After a median follow-up period of 9.5 years, a total of 358 (39%) patients had died, of these 146 (41%) had died from cardiovascular disease and 82 (23%) deaths were cancer-related. The cause of death was known for 336 (97.6%) patients. All variables were log-transformed before analysis. 

**Discussion**

Two independent GWA studies recently performed in Japanese populations identified KCNQ1 as a type 2 diabetes susceptibility gene [2,3]. We here confirm the association of the KCNQ1 common variants with an increased risk of type 2 diabetes in a Dutch population. The individuals carrying the same at-risk alleles C, as reported in the Japanese studies [2,3], had a modestly increased risk of developing type 2 diabetes, with a population attributable risk from 0.6% to 4.3%. These results are also consistent with previous studies performed in Caucasian populations [2,3,16,17,22,23]. In the present study, we demonstrate in a large cohort of subjects having undergone hyperglycaemic glucose clamps that the risk allele of the KCNQ1 SNP is significantly associated with reduced glucose-stimulated second-phase insulin secretion. In addition, we report a significant association of KCNQ1 variants with impaired lipid parameters. We could not find any significant relationship of the risk alleles of the KCNQ1 gene with type 2 diabetes complications or mortality.

The variants rs151290, rs2237892, and rs2237895 are located in intron 15 of the KCNQ1 gene on chromosome 11p15, encoding the pore-forming alpha subunit of the I(Ks) channel, a voltage-gated potassium channel that is expressed in a number of tissues, notably, the heart, pancreas, kidneys and intestine [2,3]. The encoded protein plays a role in the electrical depolarisation of the cell membrane in the heart and presumably in pancreas beta cells. It is likely to be involved in insulin secretion, although there are other possibilities, such as secretory processes in incretin (GLP-1 and/or GIP) -producing cells [2,3]. Our results provide compelling evidence that the KCNQ1 rs2237892 variant is associated with impaired second-phase insulin secretion. Moreover, these data confirm the observations from the previous studies in which the relationship between the type 2 diabetes risk alleles in KCNQ1 and reduced levels of various measures of insulin secretion have been reported [17,18,22,24], and hence, supports the hypothesis on a
The potential link between KCNQ1 and impaired beta-cell function [2,3,17,18,22,23,24,25,26].

The importance of the investigation of the various aspects of insulin secretion was highlighted previously [27]. It is thought that first phase insulin secretion is reflecting the rapid recruitment and release of insulin granules from the readily releasable pool, while second phase is due to the release of granules located further away from release site (reserve pool) and due to (new) insulin synthesis [28]. In the present study, we used the hyperglycaemic clamp technique, generally considered to be the gold standard for quantification of first- and second-phase insulin secretion to measure beta-cell function [29]. We show, for the first time, an association between a variant in the KCNQ1 gene and reduced estimates of first-phase insulin release [17]. Both above-mentioned studies used data obtained during oral glucose tolerance test [17,18]. Our data derived from the hyperglycaemic clamp extends these finding and suggests that KCNQ1 may have an effect regarding second phase secretion and therefore may have additional effects apart from early, rapid recruitment and exocytosis of insulin granules after glucose stimulation. In addition, there was a surprising trend of the association between all three risk alleles and higher ISI values. Interestingly, Boini et al. have recently shown that insulin sensitivity is strikingly increased in the KCNQ1 knockout mouse, which agrees with our results [33]. Thus, although the KCNQ1 gene is primarily a candidate gene for impaired insulin secretion, its effect on tissue-specific insulin-sensitivity independent of changes in insulin secretion is plausible.

In addition, we found a significant association between the type 2 diabetes risk allele of rs2237892 and higher level of total cholesterol and a trend towards increased level of LDL among the carriers of the same allele. Also, we observed a lower level of HDL

| Genotype (N) | rs151290 | p-value
<table>
<thead>
<tr>
<th></th>
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</tr>
</thead>
<tbody>
<tr>
<td>AA (204)</td>
<td>26.5 ± 3.6</td>
<td>26.0 ± 3.8</td>
</tr>
<tr>
<td>AC (1252)</td>
<td>5.30 ± 0.23</td>
<td>5.33 ± 0.26</td>
</tr>
<tr>
<td>CC (2035)</td>
<td>5.31 ± 0.36</td>
<td>5.31 ± 0.38</td>
</tr>
<tr>
<td>HDL cholesterol (mmol/l)</td>
<td>1.62 ± 0.40</td>
<td>1.56 ± 0.44</td>
</tr>
<tr>
<td>LDL cholesterol (mmol/l)</td>
<td>3.24 ± 0.83</td>
<td>3.31 ± 0.87</td>
</tr>
<tr>
<td>Total cholesterol (mmol/l)</td>
<td>5.44 ± 0.93</td>
<td>5.46 ± 0.95</td>
</tr>
<tr>
<td>Triglyceride (mmol/l)</td>
<td>1.29 ± 0.60</td>
<td>1.30 ± 0.69</td>
</tr>
</tbody>
</table>

| Genotype (N) | rs2237892 | p-value
<table>
<thead>
<tr>
<th></th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>TT (7)</td>
<td>23.5 ± 3.2</td>
<td>26.1 ± 3.9</td>
</tr>
<tr>
<td>CT (507)</td>
<td>5.18 ± 0.14</td>
<td>5.40 ± 0.51</td>
</tr>
<tr>
<td>CC (4623)</td>
<td>5.07 ± 0.06</td>
<td>5.27 ± 0.40</td>
</tr>
<tr>
<td>HDL cholesterol (mmol/l)</td>
<td>1.33 ± 0.29</td>
<td>1.41 ± 0.40</td>
</tr>
<tr>
<td>LDL cholesterol (mmol/l)</td>
<td>3.00 ± 0.84</td>
<td>3.08 ± 0.80</td>
</tr>
<tr>
<td>Total cholesterol (mmol/l)</td>
<td>5.11 ± 0.92</td>
<td>5.22 ± 0.96</td>
</tr>
<tr>
<td>Triglyceride (mmol/l)</td>
<td>1.57 ± 0.70</td>
<td>1.39 ± 0.81</td>
</tr>
</tbody>
</table>

| Genotype (N) | rs2237895 | p-value
<table>
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<tbody>
<tr>
<td>AA (1797)</td>
<td>26.0 ± 3.8</td>
<td>25.9 ± 3.9</td>
</tr>
<tr>
<td>AC (2509)</td>
<td>5.30 ± 0.37</td>
<td>5.40 ± 0.40</td>
</tr>
<tr>
<td>CC (857)</td>
<td>5.30 ± 0.37</td>
<td>5.31 ± 0.38</td>
</tr>
<tr>
<td>HDL cholesterol (mmol/l)</td>
<td>1.44 ± 0.41</td>
<td>1.42 ± 0.42</td>
</tr>
<tr>
<td>LDL cholesterol (mmol/l)</td>
<td>3.20 ± 0.87</td>
<td>3.21 ± 0.87</td>
</tr>
<tr>
<td>Total cholesterol (mmol/l)</td>
<td>5.36 ± 1.00</td>
<td>5.38 ± 1.00</td>
</tr>
<tr>
<td>Triglyceride (mmol/l)</td>
<td>1.39 ± 0.86</td>
<td>1.41 ± 0.81</td>
</tr>
</tbody>
</table>

*Adjusted for age, sex and study center.
**P-value for the additive model.

The data are presented as mean ± SD. All variables were log-transformed before analysis. Alleles in bold are the risk alleles for type 2 diabetes identified by previous studies. BMI: Body Mass Index. HbA1c: haemoglobin A1c (glucose bound to haemoglobin). HDL: high density lipoprotein. LDL: low density lipoprotein.

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in the carriers of the type 2 diabetes risk allele of rs151290 variant than in non-carriers. Interestingly, Chen et al. have recently reported the relationship of KCNQ1 variants with higher level of triglycerides (i.e. also rs2237892) and lower levels of HDL [34]. Although the mechanisms behind the observed associations are not clear, taken together, these data suggest that KCNQ1 may have an effect on lipid metabolism. Further studies are needed to elucidate the underlying molecular mechanisms.

In the present study, we found no significant evidence for an association of KCNQ1 with diabetic complications or mortality in type 2 diabetes patients. Recently, Ohshima et al. reported that KCNQ1 might be a potential susceptibility gene for diabetic nephropathy in a Japanese population [35]. In our sample, we could not confirm that finding. These results could be due to the relatively small sample of the patients with nephropathy in our study (there were 442 patients in our study versus 1545 diabetic patients with overt nephropathy in the study by Ohshima et al. [35]). Next, the association with nephropathy was not easy to detect due to differences in risk-allele frequency (33% in Japanese versus 5% in European) of the rs2237897 variant associated with nephropathy ($r^2 = 0.6$ with rs2237892) [35]. Our study had 20% power at $p = 0.05$ to detect the OR of 1.22 reported by Ohshima et al. [35] assuming prevalence of nephropathy of 0.40 among type 2 diabetes patients [36] and an additive model. In addition, we cannot exclude a proportion of patients with diabetic nephropathy in the reference sample. Therefore, additional replication attempts in larger studies with detailed information on the complications status are warranted.

It needs to be noted that in the current study we used control subjects younger than the patients. Thus, we cannot exclude that some of these individuals may develop diabetes in later life. However, this would result in slight reduction in statistical power and also in the ORs and would lead to the underestimation of the association of the gene with type 2 diabetes in a Dutch population. We also found that the variants in the KCNQ1 gene may have an effect on lipid metabolism. These results provide new insight into the complex pathogenesis of diabetes.

**Supporting Information**

**Table S1** Detailed clinical characteristics of the study participants from different studies. The data are presented as mean±SD. BMI: Body Mass Index. HbA1c: haemoglobin A1c. HDL: high density lipoprotein. T2D: type 2 diabetes. NA: not applicable.

**Table S2** Data on diabetic complications in type 2 diabetes patients from the DCS West-Friesland, the ZODIAC and the EPIC-NL studies.

**Table S3** Clinical characteristics of the individual hyperglycaemic clamp study samples. Data are presented as means ± SD or median (interquartile range). NTR = Netherlands Twin Register.

**Table S4** The characteristics of the Zwolle study population. Data are presented as mean ± SD or median (interquartile range) for non-normally distributed data or %.

**Table S5** Effect of KCNQ1 variants rs151290, rs2237892 and rs2237895 on beta-cell function as assessed with hyperglycaemic clamp. Analysis was adjusted for glucose tolerance status (NGT/IGT), study center, age, gender and BMI. All variables were log-transformed before analysis. $p$-values were computed for different additive models using linear generalized estimating equations (GEE), which takes into account the family relatedness when computing the standard errors. Alleles in bold are the risk alleles for type 2 diabetes identified by previous studies. DI, disposition index; IGT, impaired glucose tolerance; ISI, insulin sensitivity index; ND, not determined; NGT, normal glucose tolerance.

**Acknowledgments**

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**Author Contributions**

Conceived and designed the experiments: JVV ER MHH LMTH. Performed the experiments: JVV ER LMTH. Analyzed the data: JVV ER LMTH. Contributed reagents/materials/analysis tools: TWVH GWDL ER NK HG GB OHK AD CCVD GW HMB JMD EVTR GN LMWC HW EJB CCE FB NCO YTVDS DEG AMWS DLVDA AMS EME MD MHK DB EGD GW PES. Wrote the paper: JVV. Contributed to the interpretation of the data and to the writing of the manuscript: TWVH ER MHJ LMTH. Provided critical comments on the manuscript: GWDL NK JMD EVTR LMICW HZ NCOM YTVDS AMWS AMS.
References


