Short communication

Placental histopathology after *Coxiella burnetii* infection during pregnancy

J.M. Munster a,*, A.C.A.P. Leenders b, C.J.C.M. Hamilton c, E. Hak d, J.G. Aarnoudse a, A. Timmer e

aUniversity of Groningen, University Medical Centre Groningen, Department of Obstetrics and Gynaecology, Hanzeplein 1, 9713 GZ, Groningen, The Netherlands
bJeroen Bosch Hospital, Department of Medical Microbiology and Infection Prevention, Henri Dunantstraat 1, 5223 GZ, ’s-Hertogenbosch, The Netherlands
cUniversity of Groningen, University Centre for Pharmacy, PharmacoEpidemiology & PharmacoEconomics, Antonius Deusinglaan 1, 9713 AV, Groningen, The Netherlands
dJeroen Bosch Hospital, Department of Obstetrics and Gynaecology, Henri Dunantstraat 1, 5223 GZ, ’s-Hertogenbosch, The Netherlands
eUniversity of Groningen, University Medical Centre Groningen, Department of Pathology and Medical Biology, Hanzeplein 1, 9713 GZ, Groningen, The Netherlands

**A R T I C L E   I N F O**

Article history:
Accepted 17 November 2011

Keywords:
Q fever
*Coxiella burnetii*
Asymptomatic
Placental histopathology

**A B S T R A C T**

Symptomatic and asymptomatic *Coxiella burnetii* infection during pregnancy have been associated with obstetric complications. We described placental histopathology and clinical outcome of five cases with asymptomatic *C. burnetii* infection during pregnancy and compared these cases with four symptomatic cases from the literature. In contrast with the symptomatic cases, we did not observe necrosis or active inflammation in the placentas of the asymptomatic women. Obstetrical outcome was more favourable in the asymptomatic cases than in the symptomatic cases. Asymptomatic and symptomatic *C. burnetii* infection during pregnancy are different entities with respect to placental histopathology and the risk of obstetric complications.

© 2011 Elsevier Ltd. All rights reserved.

1. Introduction

Several European countries notified increasing numbers of human Q fever since 2007 [1,2]. Q fever is a zoonosis caused by the intracellular bacterium *Coxiella burnetii*. It primarily infects ruminants and rodents, in which the infection is mainly associated with miscarriage and stillbirth [3]. Humans are predominantly infected by inhalation of contaminated aerosols [4].

Up to 90% of pregnant women with antibodies suggesting recent infection with *C. burnetii* remain asymptomatic [5]. However, symptomatic and asymptomatic *C. burnetii* infection during pregnancy have been associated with obstetric complications, including miscarriage, preterm delivery and fetal death [6,7]. Placental infection assessed by polymerase chain reaction (PCR) or culture can be strongly related to these complications [6]. However, information about placental histopathology, in particular in asymptomatic cases, is lacking. Therefore, we described placental histopathology from women with asymptomatic *C. burnetii* infection during pregnancy. Subsequently, we compared our results with symptomatic cases described in the literature.

2. Patients and methods

2.1. Setting and participants

This study was embedded in a clustered randomised controlled trial about the effectiveness of a screening program for *C. burnetii* infection during pregnancy. In that study pregnant women living in Q fever high-risk areas in The Netherlands were serologically screened for *C. burnetii* infection. Details about the screening study are described elsewhere [8]. The study protocol was approved by the Medical Ethical Review Board of the University Medical Centre Groningen (UMCG). All participants included in this study gave written informed consent to collect and analyse placental tissue and clinical outcome data.

2.2. Design

From women who participated in the intervention group of the screening trial and who had serological evidence for an acute infection, placentas were collected. An acute infection was defined as the presence (cut-off titre ≥ 1:32) of immunoglobulin (Ig)M accompanied with (rising) IgG during follow-up. Serology was performed with indirect immunofluorescence assay (IFA, Focus Diagnostics, Cypress, CA, USA). Placentas were histopathologically analysed by one pathologist (AT) from the UMCG. Furthermore, *C. burnetii* specific real-time PCR was performed. Primers and probes used have been described earlier [9], other technical details are available on request.

2.3. Systematic review

A systematic review of the literature was done by searching PubMed and the references of the included papers following the PRISMA-guidelines. Our search was limited to human studies in English or Dutch. The search strategy was: "Q fever OR..."
**Table 1**
Summary of patient characteristics and placental histopathology.

<table>
<thead>
<tr>
<th>Age (y)</th>
<th>Parity</th>
<th>Symptoms</th>
<th>Initial serological values</th>
<th>Treatment</th>
<th>Gestational age at delivery (wks + days)</th>
<th>Birth weight (g/percentile)</th>
<th>Clinical outcome</th>
<th>C. burnetii present in placenta tissue?</th>
<th>Placental weight (g/percentile)</th>
<th>Summery of placenta histology</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>lgM I lgM II lgG I lgG II</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Asymptomatic patients</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>31</td>
<td>0</td>
<td>None</td>
<td>1:1024 1:32 1:512 &lt;1:32</td>
<td>Erythromycin 42 ± 0</td>
<td>4030/50–80th</td>
<td>Arrest of second stage of labour; uncomplicated caesarean section at term</td>
<td>PCR negative</td>
<td>Unknown</td>
<td>No significant pathology</td>
</tr>
<tr>
<td>2</td>
<td>32</td>
<td>0</td>
<td>None</td>
<td>1:256 1:32 1:1024 1:128</td>
<td>Erythromycin 37 ± 0</td>
<td>2930/50–80th</td>
<td>Suspicion of solution placenta at term; emergency caesarean section</td>
<td>PCR negative</td>
<td>540/75–90th</td>
<td>No significant pathology</td>
</tr>
<tr>
<td>3</td>
<td>31</td>
<td>0</td>
<td>None</td>
<td>1:512 1:64 1:256 1:32</td>
<td>Erythromycin 34 ± 1</td>
<td>2170/50–80th</td>
<td>PPROM, retained placenta, postpartum haemorrhage</td>
<td>PCR negative</td>
<td>337/10–25th</td>
<td>Maternal vascular underperfusion; fibromuscular hyperplasia of stem villus vessels; scattered fibrotic villi, low grade chronic villitis (Fig. 1)</td>
</tr>
<tr>
<td>4</td>
<td>34</td>
<td>0</td>
<td>None</td>
<td>1:512 &lt;1:32 1:128 &lt;1:32</td>
<td>None 39 ± 6</td>
<td>3535/50–80th</td>
<td>Uncomplicated, at term Congenital hydropneumosis, meconium stained amniotic fluid at term</td>
<td>PCR negative</td>
<td>42S/10–10th</td>
<td>Low grade chronic villitis</td>
</tr>
<tr>
<td>5</td>
<td>33</td>
<td>1</td>
<td>None</td>
<td>1:256 &lt;1:32 1:128 &lt;1:32</td>
<td>None 40 ± 2</td>
<td>3535/20–50th</td>
<td>NA</td>
<td>PCR negative</td>
<td>52S/25–50th</td>
<td></td>
</tr>
<tr>
<td>Symptomatic cases from the literature</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>1 Reichman et al., 1988</td>
<td>29</td>
<td>2</td>
<td>Fever, headache, weakness, sweating, purpuric rash</td>
<td>1:400 1:1600 1:1600 1:400</td>
<td>Tetracycline 28</td>
<td>1000/unknown</td>
<td>Induced labour because of maternal illness</td>
<td>Immunofluorescent stain positive</td>
<td>Unknown</td>
<td>Areas of necrosis</td>
</tr>
<tr>
<td>2 Raoult et al., 1994</td>
<td>26</td>
<td>Unknown</td>
<td>Fever, cough, fatigue, dyspnea, arthritis</td>
<td>Unknown; seroconversion</td>
<td>Cotrimoxazole 24</td>
<td>Unknown; rising antibodies</td>
<td>Miscarriage</td>
<td>Immunofluorescent strain positive</td>
<td>Unknown</td>
<td>Multiple foci of necrosis</td>
</tr>
<tr>
<td>3 Friedland et al., 1994</td>
<td>26</td>
<td>Unknown</td>
<td>Fever, fatigue, dyspnea, arthritis</td>
<td>Unknown; rising antibodies</td>
<td>Erythromycin postpartum 25</td>
<td>Unknown; rising antibodies</td>
<td>Oligohydramnios, intrauterine fetal death</td>
<td>Immunocytotoxic strain positive</td>
<td>Unknown</td>
<td>Severe necrotising villitis in 40% of the placental tissue</td>
</tr>
<tr>
<td>4 Bental et al., 1995</td>
<td>28</td>
<td>Unknown</td>
<td>Fever, cough, arthritis</td>
<td>1:1600 1:200 1:800 1:25000</td>
<td>Erythromycin/ rifampicin 30</td>
<td>1300/unknown</td>
<td>Premature labour, caesarean section because of transverse lie of the fetus</td>
<td>PCR positive</td>
<td>Unknown</td>
<td>No areas of necrosis or other gross pathology</td>
</tr>
</tbody>
</table>

y, years; Ig, immunoglobulin; wks, weeks; g, grams; PCR, polymerase chain reaction; NA, not applicable due to PCR inhibition; PPROM, preterm premature rupture of membranes.

* A serology of the asymptomatic cases was performed with indirect immunofluorescence assay (IFA, Focus Diagnostics, Cypress, CA, USA), measuring both IgM and IgG against phase I and II antigens. Serology of the symptomatic cases was performed with in-house assays.
C. burnetii AND "placenta". First we pre-screened the titles and the abstracts; afterwards the eligibility of the studies was judged by reading the full-texts. Only studies describing human placental histopathology were included.

3. Results

Seven of the 536 women in the intervention group of the screening trial had serological profiles suggesting an acute C. burnetii infection and were treated with antibiotics. Overall, five placentas were stored and sent for re-evaluation to the UMCG, including two placentas from women with follow-up serology suggesting a previous infection. All cases were asymptomatic at the moment of screening. Clinical outcome and placental histopathology are summarised in the first part of Table 1.

The PubMed search resulted in 30 hits. Only 2 papers included data on human placental histopathology and were included. Two other reports were included based on the references. All included papers concerned case-reports of symptomatic acute or chronic Q fever cases [10–13]. Clinical outcome and placental histopathology of these cases are summarised in the second part of Table 1.

4. Discussion

We showed that asymptomatic and symptomatic C. burnetii infection during pregnancy are different entities with respect to placental pathology and the risk of obstetric complications. Placental histology in the asymptomatic cases showed, in contrast with the symptomatic cases, no foci of necrosis or active inflammation. We only observed a few scattered fibrotic villi, which could be a result of interruption of fetal blood flow or destruction of capillaries due to previous villitis [14]. The presence of low grade chronic villitis is a frequent finding in third trimester placentas and probably related to a maternal immune response directed against fetal antigens inherited from the father. Until present no microbiological pathogens have been linked to chronic villitis [15]. Whether placenta hypoplasia and pathology consistent with maternal vascular underperfusion are linked to C. burnetii infection is to our knowledge unknown.

In none of the placentas from asymptomatic cases C. burnetii could be detected with PCR. Previously this also has been shown for a larger cohort of 153 asymptomatic seropositive women [7], suggesting that the rate of placental infection during asymptomatic C. burnetii infection is very low.

Our findings are in line with animal studies. In cows, where Q fever is usually not clinically apparent, positive PCR on bulk tank milk is only rarely associated with histopathological inflammation of placentas [16]. On the other hand, in goats and sheep, in which C. burnetii infection is often associated with miscarriage and stillbirth, necrotising inflammation of placental tissue is a common finding [17,18].

Various factors, including host immune response, cytokines and different strains of C. burnetii, have been suggested to play a role in the clinical manifestation and outcome of C. burnetii infection in both animals and humans, but further research is needed to find target points for prevention and treatment [18–20].

In conclusion, after asymptomatic C. burnetii infection during pregnancy placental examination reveals no major pathology related to previous villitis, which is associated with a favourable clinical outcome. Symptomatic infection is a different entity. Obstetric complications in these cases can very well be explained by colonisation with C. burnetii and massive necrosis of the placenta.

Authors’ contributions

JMM: I declare that I participated in the design of the study, performed the study, analysed and interpreted the data and drafted the manuscript; ACAPL: I declare that I participated in the study as an expert in laboratory testing and performed the serology; CJCMH: I declare that I participated in the study as an expert on obstetric care; EH: I declare that I initiated and designed the screening trial, wrote the grant application, and supervised the data collection, analysis and report; JGA: I declare that I participated in the study as an expert on obstetric care and supervised the analysis and report; AT: I declare that I performed the histopathological re-evaluation of the placentas, and supervised the analysis and report. All authors commented on the manuscript and approved the final manuscript.

Funding

The clustered randomised controlled trial about the effectiveness of a screening program for C. burnetii infection during pregnancy was financed by ZonMw, The Netherlands Organization for Health Research and Development; programme "Effectiviteits- en Doelmatigheidsonderzoek" (grantnumber 125030014). The funder had no role in the study design; in the collection, analysis and interpretation of data; in the writing of the report; and in the decision to submit the article for publication.

Conflict of interests

JGA and EH are members of the Health Council of the Netherlands on a non-profit base.

Ethical approval

The study was conducted according to the principles of the Declaration of Helsinki. The study protocol of the randomised controlled trial about the effectiveness of a screening program for C. burnetii infection during pregnancy was approved by the Medical Ethical Review Board of the University Medical Centre Groningen (number 2009.323). Written informed consent was obtained from all participants before placentas were collected and analysed.
Acknowledgements

The authors gratefully thank all midwives, residents, obstetricians, medical microbiologists and pathologists of the participating centres for their help in patient recruitment, and data and placenta collection.

References


