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Cadmium isotope variations in the Southern Ocean

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Cadmium concentrations and isotope compositions were determined for 47 seawater samples from the high nutrient low chlorophyll (HNLC) zone of the Atlantic sector of the Southern Ocean. The samples include 13 surface waters from a transect of the Weddell Gyre and 3 depth profiles from the Weddell Sea and Drake Passage. The Southern Ocean mixed layer samples from this study and Abouchami et al. (2011) define a clear but broad ‘HNLC trend’ in a plot of $\varepsilon^{114/110}$Cd versus [Cd], which is primarily a consequence of isotopic fractionation associated with biological uptake ($\varepsilon^{114/110}$Cd is the deviation of the $^{114}$Cd/$^{110}$Cd ratio of a sample from NIST SRM 3108 Cd in parts per 10,000). The trend is especially apparent in comparison to the large range of values shown by a global set of seawater Cd data for shallow depths. The Southern Ocean samples are also distinguished by their relatively high Cd concentrations (typically 0.2 to 0.6 nmol/kg) and moderately fractionated $\varepsilon^{114/110}$Cd (generally between +4 and +8) that reflect the limited biological productivity of this region. Detailed assessment reveals fine structure within the ‘HNLC trend’, which may record differences in the biological fractionation factor, different scenarios of closed and open system isotope fractionation, and/or distinct source water compositions. Southern Ocean seawater from depths $\geq 1000$ m has an average $\varepsilon^{114/110}$Cd of $+2.5 \pm 0.2$ ($2\sigma$, $n = 16$), and together with previous results this establishes a relatively constant $\varepsilon^{114/110}$Cd value of $+3.0 \pm 0.3$ ($2\sigma$, $n = 27$) for global deep waters. Significant isotopic variability was observed at intermediate depths in the Southern Ocean. Seawater from 200 m to 400 m in Weddell Sea has high Cd concentrations and $\varepsilon^{114/110}$Cd as low as +1, presumably due to remineralization of Cd from biomass that records incomplete nutrient utilization. Antarctic Intermediate Water, which was sampled at 150 to 750 m depth in the Drake Passage, features a distinct Cd isotope signature of $\varepsilon^{114/110}$Cd $\approx$ +4, which reflects biological isotope fractionation at the surface and subsequent mixing into the ocean interior. Taken together, our results demonstrate that coupled Cd isotope and concentration data provide valuable insights into the distribution and biological cycling of Cd in the water column. The highly systematic nature of Cd isotope signatures may furthermore prove to be of utility for future research in marine geochemistry and paleoceanography.

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1. Introduction

Cadmium is a trace element with a marine distribution similar to that of dissolved phosphate, thereby suggesting that seawater Cd concentrations are also controlled by phytoplankton uptake in surface waters and remineralization of organic matter at depth (Boyle et al., 1976; Bruland, 1980; Nolting and de Baar, 1994). A biochemical function for Cd has, furthermore, been demonstrated in some marine diatoms (Price and Morel, 1990), where Cd can replace Zn in the metalloenzyme carbonic anhydrase (Lane et al., 2005; Xu et al., 2008). The extent to which this substitution can account for the marine distribution of Cd has, however, recently been challenged (Horner et al., 2013).

Considerable Cd isotope fractionations were revealed in recent seawater studies and applied to investigate the marine cycling of this transition metal. Nutrient and Cd depleted surface waters can exhibit $\varepsilon^{114/110}$Cd values as high as +20 to +40, as a result of isotope fractionation during biological utilization ($\varepsilon^{114/110}$Cd represents the deviation of $^{114}$Cd/$^{110}$Cd for a sample relative to the
NIST 3108 Cd reference material in parts per 10,000). In contrast, the deep ocean appears to be characterized by relatively constant $\varepsilon_{114/110}\text{Cd}$ values of about $+2.5$ to $+3.0$ (e.g., Lacan et al., 2006; Ripperger et al., 2007; Gault-Ringold et al., 2012; Xue et al., 2012).

The study of Abouchami et al. (2011) was the first to investigate the Cd isotope composition of Southern Ocean surface seawater and this region is of particular interest, because the Southern Ocean constitutes by far the largest of the three major HNLC (high nutrient low chlorophyll) regions of the world ocean. The HNLC regions are characterized by relatively low amounts of biomass even though the major nutrients nitrate, phosphate and silicate are sufficiently abundant to support phytoplankton growth (Nelson and Smith, 1991). Previous studies have suggested that nutrient utilization in the Southern Ocean HNLC area is limited by the availability of Fe, a key marine trace nutrient, and light (de Baar et al., 1990, 1995; Martin et al., 1990; Sunda and Huntsman, 1997).

The present investigation follows up on the study of Abouchami et al. (2011) with coupled Cd isotope and concentration analyses for almost 50 seawater samples that were collected during the same sampling campaign. Our results for both a surface transect and three depth profiles thus significantly enlarge the limited Cd isotope dataset currently available for the Southern Ocean. Combining both datasets, we systematically investigate the processes that govern the distribution and cycling of Cd in this HNLC region.

2. Samples and sampling region

Cadmium concentrations and isotope compositions were determined for 47 seawater samples that were collected between February and March 2008 during the GEOTRACES expedition ANT XXIV/3 aboard the RV Polarstern. This includes 34 samples from three vertical profiles in the Weddell Sea (Station 198), near the Antarctic Peninsula (Station 216) and the northern Drake Passage (Station 249) and 13 surface water samples from the Zero Meridian between $62^\circ$ and $69^\circ$ S in the Weddell Gyre (Fig. 1, Table 1; Supplement Section 1). The latter samples continue and complete the Cd isotope surface water transect that was recently published by Abouchami et al. (2011) on samples from the same cruise (Supplement Section 1).

The vertical profile samples were collected with an ultraclean CTD system (de Baar et al., 2008), whilst the surface waters were obtained at 2–5 m depth with an ultraclean IFISH torpedo and pumpline system. After collection, the samples were immediately filtered using 0.2 μm nominal size cutoff filter cartridges into precleaned bottles and acidified to about pH 2 using distilled 6 M HCl. Further details of the sampling techniques are available in Middag et al. (2011).

Together, the water samples analyzed here and by Abouchami et al. (2011) comprehensively characterize the HNLC conditions of the Southern Ocean, encompassing (from north to south; Fig. 1): (i) the Subantarctic Zone (SAZ) north of the Subantarctic Front (SAF) at $\sim45^\circ$ S; (ii) the Antarctic Circumpolar Current (ACC) comprising the Polar Frontal Zone (PFZ; $\sim45^\circ$ S to $51^\circ$ S) that extends from the SAF to the Antarctic Polar Front (APF) and the Antarctic Zone (AAZ; $\sim51–56^\circ$ S) from the APF to the Southern Boundary of the ACC (SB-ACC); and (iii) the Weddell Gyre south of $\sim56^\circ$ S.

Numerous water masses are encountered in this region but only aspects significant for the present study are mentioned below. The upwelling of Circumpolar Deep Water (CDW) in the Weddell Gyre and AAZ generates Antarctic Surface Water (AASW), which spreads northward, subducting north of the APF to contribute to the formation of Antarctic Intermediate Water (AAIW). Subantarctic Mode Water (SAMW), formed from deep winter mixed layers, overlies AAIW north of the SAF and export of both water masses occurs in northward direction. More detailed summaries of the circulation system and water masses present in the Atlantic sector of the Southern Ocean are provided elsewhere (e.g., Whitworth and Nowlin, 1987; Orsi et al., 1993, 1995; relevant studies from the ANT XXIV/3 expedition are listed in Supplement Section 2).

3. Analytical methods

The instruments and techniques that were employed for the acquisition of data for salinity and potential temperature, as well as oxygen and nutrient concentrations, are described elsewhere (Middag et al., 2011, 2012). The coupled Cd isotope composition and concentration analyses were carried out at the Imperial College MAGIC Laboratories, using recently published methods (Ripperger and Rehtkämper, 2007; Xue et al., 2012). In brief, about 0.3 to 1.1 L of seawater were weighed to obtain at least 30 ng of natural Cd and a suitable volume of a $^{111}$Cd–$^{113}$Cd double spike was added to achieve a spike to sample ratio of $\sim$1. A three-stage column chemistry was then employed for the separation of Cd at yields of better than 90% and with a total procedural Cd blank of $\sim$20 pg, equivalent to less than 0.1% of the indigenous Cd present in the samples. The isotopic analyses were carried out using a Nu Plasma HR MC-ICP-MS (multiple collector inductively coupled plasma mass spectrometer), with data being processed offline to calculate the Cd isotope compositions of the samples. In the following, these results are expressed as $\varepsilon_{114/110}\text{Cd}$ values, which are given relative to the NIST SRM 3108 Cd isotope standard (std):

$$\varepsilon_{114/110}\text{Cd} = \left(\frac{\left[114\text{Cd}/110\text{Cd}\right]_{\text{sample}}}{\left[114\text{Cd}/110\text{Cd}\right]_{\text{std}}} - 1\right) \times 10,000$$

Additional details of the secondary Cd isotope reference materials BAM-1012 Cd ($\varepsilon_{114/110}\text{Cd} = -13.57 \pm 0.60$, 25d; $n = 17$), JMC Cd Münster ($-0.96 \pm 0.21$, $n = 25$), and Alfa Cd Zurich ($-0.61 \pm 0.29$, $n = 17$) were carried out during measurement sessions, to monitor data quality and all results showed excellent agreement with consensus values ($-13.32 \pm 0.43$, $-0.89 \pm 0.42$, and $-0.20 \pm 0.51$, respectively; Abouchami et al., 2012). Cadmium isotope data from the literature that were acquired relative to other zero-epsilon reference materials are reported here relative to NIST SRM 3108 Cd, based either on the community-consensus isotopic offsets between the standards (Abouchami et al., 2012) or the conversions given in the original publication.
Table 1
Cd concentrations and isotope compositions and other key data for the seawater samples analyzed in this study.

<table>
<thead>
<tr>
<th>Sampling Station</th>
<th>Latitude</th>
<th>Longitude</th>
<th>Depth (m)</th>
<th>Salinity</th>
<th>$\theta$ (°C)</th>
<th>$[O_2]$ (μmol/kg)</th>
<th>$[PO_4^{3-}]$ (μmol/kg)</th>
<th>$[Si]$ (μmol/kg)</th>
<th>$[NO_3^-]$ (μmol/kg)</th>
<th>$[Cd]$ (nmol/kg)</th>
<th>$\epsilon^{114/110}$Cd ±2sd</th>
</tr>
</thead>
<tbody>
<tr>
<td>142</td>
<td>62.23°S</td>
<td>0.00</td>
<td>2–6</td>
<td>33.95</td>
<td>0.44</td>
<td>1.69</td>
<td>57.76</td>
<td>23.53</td>
<td>0.270</td>
<td>6.763</td>
<td>0.50</td>
</tr>
<tr>
<td>145</td>
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<td>33.91</td>
<td>0.59</td>
<td>1.61</td>
<td>53.83</td>
<td>23.15</td>
<td>0.257</td>
<td>7.313</td>
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<tr>
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<td>1.60</td>
<td>46.87</td>
<td>22.76</td>
<td>0.247</td>
<td>7.382</td>
<td>0.50</td>
</tr>
<tr>
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<td>65.24°S</td>
<td>0.00</td>
<td>2–5</td>
<td>33.94</td>
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<td>1.62</td>
<td>62.49</td>
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<td>5.319</td>
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<tr>
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<td>0.01°E</td>
<td>2–6</td>
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<td>1.61</td>
<td>61.97</td>
<td>25.52</td>
<td>0.373</td>
<td>6.340</td>
<td>0.50</td>
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<td>1.64</td>
<td>59.23</td>
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<td>5.760</td>
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<td>0.41</td>
<td>1.63</td>
<td>58.60</td>
<td>25.41</td>
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<td>7.483</td>
<td>0.68</td>
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<tr>
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<td>0.76</td>
<td>1.67</td>
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<tr>
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<td>0.73</td>
<td>1.77</td>
<td>65.49</td>
<td>25.77</td>
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<tr>
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<td>0.74</td>
<td>1.78</td>
<td>62.87</td>
<td>26.40</td>
<td>0.413</td>
<td>4.830</td>
<td>0.70</td>
</tr>
<tr>
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<td>2–6</td>
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<td>0.78</td>
<td>1.74</td>
<td>60.58</td>
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<td>0.384</td>
<td>4.997</td>
<td>0.70</td>
</tr>
<tr>
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<td>2–6</td>
<td>33.92</td>
<td>1.02</td>
<td>1.75</td>
<td>60.50</td>
<td>25.66</td>
<td>0.390</td>
<td>5.000</td>
<td>0.50</td>
</tr>
<tr>
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<td>1.45</td>
<td>1.88</td>
<td>73.33</td>
<td>27.59</td>
<td>0.549</td>
<td>4.048</td>
<td>0.63</td>
</tr>
<tr>
<td>216</td>
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<td>50.84°W</td>
<td>25</td>
<td>34.14</td>
<td>1.85</td>
<td>2.05</td>
<td>73.04</td>
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<td>0.645</td>
<td>4.653</td>
<td>0.82</td>
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<tr>
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<td>1.81</td>
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<td>79.17</td>
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<td>0.773</td>
<td>2.014</td>
<td>0.497</td>
<td>0.49</td>
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<tr>
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<td>1.48</td>
<td>2.26</td>
<td>99.37</td>
<td>32.36</td>
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<tr>
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<td>35.78</td>
<td>1.81</td>
<td>2.30</td>
<td>104.25</td>
<td>32.37</td>
<td>0.789</td>
<td>1.970</td>
<td>0.35</td>
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<tr>
<td>249</td>
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<td>63.76°W</td>
<td>3.33</td>
<td>4.56</td>
<td>1.66</td>
<td>5.23</td>
<td>23.87</td>
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<td>6.680</td>
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<tr>
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<td>35.33</td>
<td>4.56</td>
<td>1.67</td>
<td>5.24</td>
<td>24.08</td>
<td>0.395</td>
<td>7.023</td>
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<tr>
<td>254</td>
<td>56.88°S</td>
<td>62.52°W</td>
<td>2–5</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>0.82</td>
</tr>
</tbody>
</table>

$\theta$ = potential temperature. The $\epsilon^{114/110}$Cd values are reported relative to NIST SRM 3108 Cd (Abouchami et al., 2012). The quoted uncertainties for $\epsilon^{114/110}$Cd are based on the ±2sd uncertainties determined for multiple analyses of matching standard solutions that were analyzed before and after each sample measurement. The Cd concentrations have an estimated uncertainty of about 1–2% (Xue et al., 2012). No hydrographic data (salinity, $\theta$, $[O_2]$) are available for the sample from Station 245.

4. Results

4.1. Hydrography and nutrient concentrations for Zero Meridian surface seawater

In Fig. 2, the hydrographic and nutrient concentration data for the 13 Zero Meridian surface waters of this study (from 62.2°S to 69.0°S in the Weddell Gyre) are plotted together with the samples of Abouchami et al. (2011), which were collected primarily north of 61.2°S. Large gradients in nutrient content are clearly evident (Fig. 2), in agreement with previous observations (e.g., Sarmiento et al., 2004). North of the SAF, the SAZ features low $[NO_3^-]$, $[PO_4^{3-}]$ and $[Si]$. To the south, the ACC comprises the high $[NO_3^-]$, $[PO_4^{3-}]$ and low $[Si]$ regime of the PFZ as well as the high $[NO_3^-]$, $[PO_4^{3-}]$, $[Si]$ surface waters of the AAZ. The more southern Weddell Gyre is uniformly high in $[NO_3^-]$, $[PO_4^{3-}]$, $[Si]$. Hence only the AAZ and Weddell Gyre are truly HNLC for all three major nutrients.

Focusing on the Weddell Gyre data of the present study, the potential temperatures reveal two main groups of samples. Between 62.2°S and 64.2°S, the potential temperature of the surface water is nearly constant at ~0.5°C and the samples are in accord with the temperature trend established by surface seawater collected further to the north. However, at 64.2°S there is a small but well-resolved drop in potential temperature. It is likely that these two groups of samples (north and south of 64.2°S; Fig. 2) reflect the eastward and westward flowing limb of the Weddell Gyre, respectively (Klunder et al., 2011). This interpretation is corroborated by nutrient ($[NO_3^-]$, $[PO_4^{3-}]$, $[Si]$) data, which also fall in two groups that encompass the same latitudes. In detail, $[NO_3^-]$, $[PO_4^{3-}]$, and $[Si]$ decrease southwards in the Weddell Gyre until about 65°S, but even further to the south, the parameters first increase and then decrease in southward direction (Fig. 2). The decrease south of ~67.5°S is probably related to an earlier (December 2007) algae bloom, which followed the retreating ice edge with a time lag (Rutgers van der Loeff et al., 2011).
Fig. 2. The Cd concentrations and isotopic compositions, as well as hydrographic and nutrient concentration data are shown as a function of latitude for the Zero Meridian surface water samples of this study and Abouchami et al. (2011). Most noteworthy is how well the Cd data of Abouchami et al. (2011) for the surface water sample from Station 163 at ∼66°S matches with the Cd results of the present study for neighboring locations. SAZ = Subantarctic Zone, PFZ = Polar Frontal Zone, AAZ = Antarctic Zone, ACC = Antarctic Circumpolar Current, SAF = Subantarctic Front, APF = Antarctic Polar Front. Note the strong gradient versus latitude for [Si], which is strongly depleted in the PFZ whilst [PO₄³⁻] and [NO₃⁻] are high. Only the AAZ and Weddell Gyre are thus truly HNLC for all three major nutrients.

4.2. Cd concentrations and isotope compositions of Zero Meridian surface seawater

The Cd concentrations and isotope compositions of the Zero Meridian surface seawater from this investigation and Abouchami et al. (2011) are also shown in Fig. 2. A key observation is that the data of the two studies match perfectly, with no apparent offsets in the results. This conclusion is in accord with previous work by the two laboratories, which demonstrated excellent agreement in the Cd data for both isotope reference materials (Abouchami et al., 2012) and GEOTRACES seawater intercomparison samples (Boyle et al., 2012). These observations are important, because they allow the Cd datasets of both studies to be interpreted in unison, with no need to correct for systematic differences in the results.

Focusing on the surface waters analyzed here, the Weddell Gyre transect displays Cd concentrations of 0.25 nmol/kg to 0.46 nmol/kg and ε¹¹⁴/¹¹⁰Cd values between about +4.5 and +7.5 (Fig. 2, Table 1). Importantly, these data can be used to divide the samples into the same two regional groups (north and south of ∼65°S) that were previously identified based on potential temperature and major nutrients.

4.3. Hydrography and nutrient concentrations for the depth profiles

The hydrographic and nutrient data for the three depth profiles are summarized in Fig. 3. The most apparent feature is the similarity of the results obtained for Stations 198 and 216, whilst Station 249 from the Drake Passage differs in its physical and chemical properties. This result is not surprising given that the two former stations are within the Weddell Gyre (Fig. 1), whilst the latter is situated in the PFZ of the ACC system (Middag et al., 2012) with its distinct hydrographic and chemical characteristics (e.g., Löscher et al., 1998; Sarmiento et al., 2004; Middag et al., 2011; Abouchami et al., 2011). Stations 198 and 216 show temperature minima near the surface due to the presence of AASW, whilst Station 249 records a surface water temperature maximum from inflow of warmer waters into the PFZ. Notably, Station 249 exhibits much lower [Si] than the Weddell Gyre (Stations 198, 216), particularly at or near the surface (Fig. 3). In contrast, the depth profiles of all three stations reveal that [PO₄³⁻] and [NO₃⁻] are modestly depleted in the surface waters, whereby this depletion is slightly more pronounced in the Drake Passage (Station 249). Also conspicuous are the data for the deepest waters of Station 216 (Fig. 3; Table 1). These samples were collected just above the shelf of the Antarctic Peninsula and they feature temperatures more akin to the surface than the overlying waters, high O₂ contents, as well as low concentrations of [Si] and to a lesser extent other nutrients.

4.4. Cd concentrations and isotope compositions for the depth profiles

The depth profiles of Cd concentrations and isotope compositions are relatively similar for all three stations (Fig. 4, Ta-
Table 1, whereby Cd is modestly depleted in surface waters with concentrations ranging from ~0.4 nmol/kg in the Drake Passage to ~0.7 nmol/kg in the Weddell Gyre. These results are in accord with the only moderately fractionated $\varepsilon^{114/110}$Cd values of between +4.0 to +6.7. In addition, all depth profile samples from below 1 km feature nearly identical Cd concentrations and isotope compositions, whereby the largest deviation (with relatively low [Cd] and high $\varepsilon^{114/110}$Cd) is seen for 2450 m at Station 216. Nonetheless, these data (and one additional published result; Ripperger et al., 2007) provide well-defined Southern Ocean deep water averages of $[\text{Cd}] = 0.77 \pm 0.01$ nmol/kg and $\varepsilon^{114/110}$Cd = +2.5 ± 0.2 (2σ, n = 16) for samples from depths >1000 m (see Supplement Section 5).

At depths of between about 200 m and 1 km there are clear differences in the Cd data for Station 249 versus Stations 198 and 216 (Fig. 4). Most notably, Station 249 reveals $\varepsilon^{114/110}$Cd values
the largest variability of Cd concentrations and isotope compositions. The new HNLC data are hence in accord with earlier studies, which concluded that biological cycling of Cd is the main process responsible for generating variations in seawater $\varepsilon^{114/110}\mathrm{Cd}$ (Lacan et al., 2006; Ripperger et al., 2007; Abouchami et al., 2011; Gault-Ringold et al., 2012; Horner et al., 2013).

Fig. 5 furthermore confirms that surface and near-surface seawater from the HNLC ocean features relatively high Cd contents and, therefore, can be readily distinguished from non-HNLC locations. To a lesser extent, such a distinction is also possible based on Cd isotope compositions. The shallow non-HNLC ocean shows significant isotopic variability, particularly at low [Cd] with $\varepsilon^{114/110}\mathrm{Cd}$ values from about $-5$ to $+40$. In contrast, the HNLC samples feature much less isotopic variability with Cd isotope effects that rarely exceed $\varepsilon^{114/110}\mathrm{Cd} \approx +8$. Assuming that Cd isotope fractionation results from biological cycling, the modest range of $\varepsilon^{114/110}\mathrm{Cd}$ in the HNLC ocean is consistent with the low productivity of these regions and the limited uptake of Cd by phytoplankton.

Fig. 5 also highlights that the HNLC surface and subsurface samples ($\lesssim 100$ m depth; orange symbols) define a comparatively tight trend, in comparison to the large global variability of Cd data in similar samples from other locations. This observation holds even when our results for a single growing season are compared with data from the only previous regional study of marine Cd isotope variations (Gault-Ringold et al., 2012), which investigated Subantarctic surface waters that were collected on a $\sim 100$ km long transect, albeit over a two year period (Fig. 5; green symbols). The variability of the global data reflects that the samples are from a range of biogeochemical regimes, such that they record both different conditions of Cd uptake and a myriad of other processes that can impact Cd systematics, including mixing of isotopically distinct water masses, external inputs of Cd (e.g., from aerosol), and remineralization of organic matter. The isotopic trends that are generated by biological uptake can also differ in response to a number of variables. For example, correlations with distinct slopes can be generated in Fig. 5 for different values of the biological fractionation factor $\alpha$, where $\alpha = R_{\text{biomass}}/R_{\text{seawater}}$ and $R = 114\mathrm{Cd}/110\mathrm{Cd}$ (Fig. 5a). Further variability can be produced even when $\alpha$ is constant, if Cd uptake occurs at conditions that fluctuate between the endmember cases of closed system Rayleigh fractionation ($\alpha_{\text{end}}$) and steady state isotope fractionation in an open system ($\alpha_{\text{os}}$; Fig. 5a).

Given the multiple factors that can impact marine Cd systematics it is not surprising that the global data for surface seawater collected at different times and locations exhibits such large variability in Fig. 5. In contrast, the comparatively tight ‘HNLC trend’ of the Southern Ocean surface and subsurface seawater thus reflects a comparatively uniform history of marine conditions and biogeochemical processes that impact Cd cycling, even though the samples were collected over a large region during a period of nearly 2 months (Fig. 1).

5.2. Biological Cd isotope fractionation in the Southern Ocean

Fig. 6 focuses only on the Cd data available for samples from the upper mixed layer of the Southern Ocean. This comprises the surface waters from the ‘Zero Meridian’ transect and samples from the surface mixed layer (Rutgers van der Loeff et al., 2011) of Stations 198 (25 m depth), 216 (25 m), 245 (2–5 m) and 249 (25, 50, 75 m). These graphs confirm the previously inferred Southern Ocean ‘HNLC trend’, but also reveal significant fine structure (and/or scatter) within the correlation. In particular, it is apparent that the ACC samples typically feature slightly higher $\varepsilon^{114/110}\mathrm{Cd}$ values at a given [Cd] compared to the Weddell Gyre surface waters. The upper mixed layer samples from the Drake Passage (Stations 245, 249) and Station 198 support this conclusion, with Cd systematics akin to those of the ACC and Weddell Gyre surface
samples, respectively (Fig. 6). In the following, we investigate the systematic nature of the Cd data within the HNLC trend and, using simple modeling, assess possible causes for its features. In particular, the discussion focuses on (i) variable fractionation factors, (ii) changes in the fractionation conditions, and (iii) differences in source water compositions.

5.2.1 Variable apparent fractionation factors

Previously, Abouchami et al. (2011) concluded that their Southern Ocean samples defined two linear correlations in a plot of $\varepsilon^{114/110}$Cd versus ln[Cd], implying two distinct values of $\alpha$ (Fig. 6a). In detail, they found that samples from ACC locations between 46°S and 56°S define a correlation with a slope equivalent to a fractionation factor of $\alpha_{ACC} = 0.99960$, whilst most samples from 56°S to 61°S in the Weddell Gyre are in accord with a less steep trend indicative of $\alpha_{WG} = 0.99975$ (Fig. 6a, red and blue dashed lines). Abouchami et al. (2011) inferred that the Cd systematics of these samples are in accord with biological isotope fractionation in two regimes that are associated with distinct values of $\alpha$, as a consequence of differences in phytoplankton biomass, community composition, and/or physiological mechanisms of Cd uptake (see Supplement Section 3).

Combination of the present data with those of Abouchami et al. (2011) reveals that 7 of the new mixed layer samples from south of 56°S are in accord with the Weddell Gyre trend, whilst the Drake Passage samples support the previous ACC correlation. However, Abouchami et al. (2011) also observed that two of their Weddell Gyre samples did not conform to the expected trend (open dark blue symbols; Fig. 6a) and even more "scatter" is added by samples from the present study (open dark blue symbols; Fig. 6a). These deviations from the predicted closed system Rayleigh fractionation trends may reflect atypical conditions at the respective locations, which are associated with distinct or intermediate fractionation factors. For example, three of the present samples from the Weddell Gyre, located between 60° and 65°S fall along the ACC correlation. Such a shift to heavier $\varepsilon^{114/110}$Cd values was also noted by Abouchami et al. (2011) for one of their samples from this region and thought to be related to an earlier phytoplankton bloom in December 2007 (see Supplement Section 3). The Weddell Gyre samples from around 66°S plot between the ACC and Weddell Gyre trends of Fig. 6a and they also show unusual contrasting changes in $\varepsilon^{114/110}$Cd and [Cd] (Fig. 2). It is conceivable that these features reflect the influence of melt water from icebergs, an explanation that has also been put forward for the variable Fe concentrations in this region (Klunder et al., 2011).

5.2.2 Closed versus open system fractionation – variable ‘conditions’ of Cd uptake and isotope fractionation

Figs. 6a/b show that the Southern Ocean mixed layer samples are in accord with linear $\varepsilon^{114/110}$Cd versus [Cd] trends regardless
of whether the concentrations are plotted in ln([Cd]) or [Cd] space. This indicates that the correlations are, in principle, compatible with both closed system Rayleigh fractionation (Fig. 6a) and open system isotope fractionation in a steady state system (Fig. 6b). The compatibility of the data with both endmember models of isotope fractionation reflects, at least in part, the comparatively modest depletion of Cd in HNLC surface waters compared with the global ocean dataset (Fig. 5). A clear isotope distinction between the models is only seen at low Cd contents, but such conditions are not observed in the Southern Ocean.

A key difference between the two scenarios is that an open, steady state system requires a larger isotope fractionation factor (i.e., smaller value of $\alpha$) to account for the correlations seen in Fig. 6b, in comparison to the closed system model of Fig. 6a. Using similar and realistic source compositions (see caption of Fig. 6), steady state fractionation can span the Cd data with $\alpha_{ss} = 0.9996$ to 0.9999 (hashed lines; Fig. 6b), whilst $\alpha_{ss}$ values between 0.9998 and 0.9995 are required to completely cover the results for closed system Rayleigh fractionation (black dashed lines, Fig. 6a). The modeling of Fig. 6c demonstrates that the total variability of the system Rayleigh fractionation (black dashed lines, Fig. 6a). The heavier Cd isotope compositions seen in the ACC compared to the ACC (see Section 5.3.2) and upwelling of nutrient-rich CDW is limited to regions south of the APF. Hence, the gentler slope of the Weddell Gyre correlation compared to the ACC (Fig. 6) may reflect conditions that are more favorable for open system fractionation. In contrast, transient phytoplankton blooms will generate high rates of Cd removal and can therefore act to generate temporary shifts in the apparent ‘openness’ of a system toward more Rayleigh-like (closed system) conditions. The bloom observed between 60° to 65°S in December 2007 is, therefore, consistent with the unusually high $^{114/110}$Cd values for the Weddell Gyre samples from this area.

5.2.3. Differences in source water compositions

It is also possible that the variability of the mixed layer Cd data (Fig. 6) is a consequence of differences in source water compositions. This scenario is discussed in the following based on statistical tests (Supplement Section 4) that were applied to all mixed layer results with the exception of Station 216, which may be affected by the proximal shelf. Seawater from the Drake Passage and transect samples from north of 56°S were hereby assigned to the ACC regime ($n = 15$) and more southern samples to the Weddell Gyre ($n = 22$), in accord with the division adopted in earlier studies of samples from the same cruise (e.g. Abouchami et al., 2011; Middag et al., 2011; Klunder et al., 2011). The statistical assessment pertains to data plotted in $^{114/110}$Cd versus ln([Cd]) space, which assumes Rayleigh distillation (Fig. 6d), but very similar results were obtained for a linear plot and a steady state system (Supplement Section 4). The complete dataset defines a slope of $\alpha_{BE} = 0.99959 \pm 0.00008$ (all uncertainties are ±2sd), but separating the data into an ACC and Weddell Gyre trend, based on the division outlined above, provides a statistically significant improvement in fit ($p = 1 \times 10^{-7}$). These two trends (Fig. 6d, dashed lines) have identical slopes, corresponding to fractionation factors of $\alpha_{ACC} = 0.99959 \pm 0.00005$ and $\alpha_{WG} = 0.99959 \pm 0.00009$, but they are offset by $^{114/110}$Cd = 1.1. This suggests that the Cd data are in accord with assignment of the samples into two different groups – as in Abouchami et al. (2011) – albeit with identical slopes (or apparent $\alpha$’s). The small but significant offset between the two trends can be explained by a difference in the Cd isotope composition of the source water masses and this may reflect larger fluxes of isotopically light Cd into Weddell Gyre source waters from the remineralization of biomass or perhaps upwelling of water with more positive $^{114/110}$Cd in the ACC region.

5.2.4. Synthesis

Biological cycling of Cd in the mixed layer of the Southern Ocean results in a co-variation of dissolved $^{114/110}$Cd with [Cd]. Detailed assessment reveals significant fine structure within the ‘HNLC trend’, which is most likely caused by either changes in the biological fractionation factor (Abouchami et al., 2011), variable ‘fractionation conditions’ (closed versus open system) and/or distinct compositions for the source water masses. A better understanding of the relative importance of these factors in determining the Cd systematics seen in Fig. 6 is desirable, as this would provide new insights into the processes that govern the dynamic cycling of Cd in the upper water column.

In principle, the relative impact of changes induced by variable fractionation factors and/or conditions could be assessed based on independent estimates of $\alpha$ from culturing experiments, but only scant data are currently available. Experiments with freshwater chlorophytes provide a fractionation factor of 0.9986 ± 0.0006 (Lacan et al., 2006), whereas the ‘accidental’ growth of phytoplankton in unfertilized seawater (Xue et al., 2012) and experiments with genetically modified bacteria (Horner et al., 2013) yielded estimates of 0.9995 ± 0.003 and ~0.9997, respectively. Perhaps most relevant for the Southern Ocean are the data of Gault-Ringold (2011) for cultured diatoms grown under Zn-limited and Zn-replete conditions, and these give $\alpha$ values of 0.9996 ± 0.0001 and 0.9991 ± 0.0001, respectively. Notably, the results of all studies are in good agreement with the fractionation factors that can be inferred from the Southern Ocean seawater trends (Fig. 6). Given that culturing experiments can never reproduce, but only approximate, actual marine conditions, the current data provide insufficient constraints to rule out any of the scenarios for the origin of the variable Cd isotope systematics seen in Fig. 6. Hence, it is possible that either of these variables dominates the Cd systematics or they could be determined by a combination of factors.

The different Cd fractionation patterns seen in the ACC and Weddell Gyre by Abouchami et al. (2011) were inferred to reflect two distinct marine biogeochemical provinces, whereby atypical conditions in some regions are responsible for samples with unusual compositions (Fig. 6). Our results suggest an alternative interpretation. When considered in a global marine context, the mixed layer samples from the Southern Ocean display a tightly clustered trend (Fig. 5), and the Southern Ocean as a whole hence appears to offer comparatively homogeneous conditions of Cd cycling. Within this domain, the trends shown by the majority of the ACC and Weddell Gyre samples denote the overall variability of the processes and conditions that impact the cycling of Cd (Fig. 6). The heavier Cd isotope compositions seen in the ACC compared to the Weddell Gyre may represent more positive $^{114/110}$Cd source waters in the ACC, the expression of smaller $\alpha$ values (that generate larger fractionations) during Cd uptake and/or ‘fractionation conditions’ that are closer to idealized closed system Rayleigh dis-
tillation. Samples that are intermediate to these two endmembers, particularly from the Weddell Gyre, can thus be explained in terms of intermediate marine conditions that produce transitional fractionation signatures.

Based on our assessment, it will be useful for future studies to characterize the biological fractionation factor for Cd at conditions that are relevant for the surface ocean. Also important are investigations that constrain the effect of nutrients on marine Cd fractionation systems. Such an impact is expected, given that a number of studies have reported that the uptake of Cd by phytoplankton is affected by ambient carbon dioxide concentrations, and the dissolved seawater contents of Cd, Fe, Zn and Mn (Lee et al., 1995; Sunda and Huntsman, 1998, 2000; Cullen et al., 1995, 2003; Cullen and Sherrell, 2005; Cullen, 2006; Lane et al., 2009).

5.3. Depth profiles

5.3.1. Deep water

The increasing Cd concentrations seen along the deepwater pathways of the meridional overturning circulation from the North Atlantic/Arctic Ocean, to the Southern Ocean and the Pacific are discernible in Fig. 5b. Our new data also supports previous work, which concluded that deep waters show only limited variability in Cd isotope compositions (Ripperger et al., 2007). Including the results of the present study, the new global dataset for seawater in Cd isotope compositions (Ripperger et al., 2007). Including the which concluded that deep waters show only limited variability discernible in Fig. 5b. Our new data also supports previous work, accounting for the nearly constant

ε_{114/110}Cd of +3.0 ± 0.3 (2σ, n = 27; see Supplement Section 5).

Fig. 5b highlights that the highest ε_{114/110}Cd values for deep water, of up to about +5, are found in samples from the Arctic and North Atlantic Ocean. Excluding these values, the compilation of Southern and Pacific Ocean deep waters have a marginally lower mean ε_{114/110}Cd = 2.6 ± 0.2 (2σ, n = 21; Supplement Section 5). Further data are, however, required to ascertain if there is a small systematic ε_{114/110}Cd offset between Arctic/North Atlantic and Southern/Pacific Ocean deep water masses. If confirmed, the more positive ε_{114/110}Cd values found in the former may represent a residual fractionated surface signal that is retained, along with low preformed [Cd], due to the young ventilation age of these water masses.

5.3.2. Stations 198 and 216 in the Weddell Gyre

Seawater from 200 m to 400 m depth at Station 198 in the Weddell Gyre demonstrates ε_{114/110}Cd values as low as about +1 (Fig. 4). A similar signature also appears to be present at Station 216 but this is less well resolved as the lowest ε_{114/110}Cd value is +2. It is unlikely that these light Cd isotope compositions are related to upwelling of CDW, the deep water source of Cd to the study sites, because all deep waters analyzed here display ε_{114/110}Cd values of higher than +2. The upwelling can, however, account for the nearly constant ε_{114/110}Cd ≈ 2.5, which is observed for depths >500 m at Station 198. Significantly, the lightest Cd isotope compositions for Stations 198 and 216 are recorded at approximately the same depths that show the highest [Cd]. This suggests that the low ε_{114/110}Cd values reflect addition of isotopically light Cd, most likely from remineralization of organic matter. Mass balance modeling (Fig. 7a) is used in the following to support this conclusion.

The green lines of Fig. 7a indicate the calculated isotopic evolution for Cd that is accumulated in biomass by biological uptake. These calculations are for both closed system Rayleigh fractionation and for an open, steady state system using small but reasonable values for α to maximize the extent of fractionation (α_{max} = 0.9996 and α_{calc} = 0.9994) and a realistic initial seawater composition (ε_{114/110}Cd = +2.5, [Cd] = 0.8 nmol/kg). The modeling shows that biomass with isotopically light Cd is likely to be common in HNLC regions where Cd uptake is incomplete. In detail, the accumulated biomass features ε_{114/110}Cd ≈ 0, if the residual dissolved seawater Cd concentration (following biological removal) exceeds 0.3 nmol/kg, in accord with the majority of the Weddell Gyre surface waters (Fig. 7a).

Fig. 7a also shows the mixing curves (in red and blue) that are generated by addition of Cd released from biomass to typical Southern Ocean surface waters, whereby the latter are the starting points of the mixing trends at low [Cd]. The biomass is assigned a Cd abundance of 2 × 10^4 nmol/kg, in accord with published elemental data for diatoms and other phytoplankton (Price, 2005; Ho et al., 2003; Supplement Section 6), but the exact value of this parameter is of little consequence because essentially identical results are obtained for Cd contents of 10^4 to 10^6 nmol/kg.
For biomass characterized by $\varepsilon^{114/110}\text{Cd} = -1.0$, a value that is in accord with both Rayleigh and steady state fractionation, remineralization can produce $\varepsilon^{114/110}\text{Cd}$ values as low as $+1.5$ for seawater with [Cd] $\approx 0.9$ nmol/kg, as observed for Stations 198 and 216 at depths of 200 to 400 m (full red lines, Fig. 7a).

The model described above cannot, however, account for $\varepsilon^{114/110}\text{Cd}$ values of less than $+1.4$, as observed for two samples from Station 198. For remineralization to produce seawater with $\varepsilon^{114/110}\text{Cd} \approx +1.1$ [Cd] $\approx 0.9$ nmol/kg, the biomass must feature $\varepsilon^{114/110}\text{Cd}$ of about $-2$ or less (dashed blue lines, Fig. 7a). Given the constraints of the seawater data on the fractionation factor $\alpha$ (Fig. 6), such biomass can only be produced by isotope fractionation in an open or at least partially open system, rather than by Rayleigh distillation (Fig. 7a). This conclusion is in accord with the observation that the majority of the Weddell Gyre surface waters are offset to lower $\varepsilon^{114/110}\text{Cd}$ values in comparison to the ACC trend (Fig. 6).

The possible role of remineralization can be further evaluated by incorporating the C budget into the modeling. Assuming that the biomass is characterized by a molar C/Cd ratio of $3.3 \times 10^5$ (Price, 2005; Ho et al., 2003; Supplement Section 6), the extent of remineralization that is needed to account for the Cd-rich water samples with $\varepsilon^{114/110}\text{Cd} \approx +1$ at Station 198 would increase the concentration of dissolved inorganic carbon (DIC) by about 150 to 250 $\mu$mol/kg, equivalent to an increase of about 5 to 10%. Given the uncertainties of the mass balance, this prediction is in accord with the observation that Station 198 samples from 200 to 400 m depth feature DIC contents, which exceed the surface concentrations by about 90 $\mu$mol/kg (Supplement Section 1). In summary, the modeling demonstrates that the addition of remineralized Cd, which records incomplete nutrient utilization at the surface, provides a reasonable explanation for the low $\varepsilon^{114/110}\text{Cd}$ values and high Cd contents seen at 200 to 400 m depth in the Weddell Sea.

Of further interest is the deepest sample from Station 216, obtained just above the shelf sediments of the Antarctic Peninsula at 2450 m, which features conspicuous hydrographic and nutrient data (Fig. 3). Based on these and other observations (e.g., elevated [Mn] and [Al]), Middag et al. (2013) concluded that freshly ventilated deep waters are being sampled at this location, which formed close to the shelf by cooling, brine rejection and subsequent subduction. The low Cd concentration and hint of higher $\varepsilon^{114/110}\text{Cd}$ for the 2450 m sample (Fig. 4, Table 1) are in accord with the conclusion of Middag et al. (2013) and hence indicate that the fractionated Cd isotope signatures of surface waters can be subducted into and retained within the deep ocean.

5.3.3. Station 249 in the Drake Passage

Nearly constant $\varepsilon^{114/110}\text{Cd}$ values of about $+4$ were found for seawater from depths of 150 to 750 m at Station 249 in the Drake Passage (Fig. 4). This result is of interest because such positive and relatively constant $\varepsilon^{114/110}\text{Cd}$ values were not detected at similar depths in any previous study and it is conceivable that they are also a residual signal from isotope fractionation at the surface.

Based on the hydrographic data of the cruise (Table 1), Station 249 is clearly within the PFZ (Middag et al., 2012). These results are also in accord with studies of physical oceanography, which have shown that AAIW is present within the PFZ below the mixed layer, centered on a depth of about 500 m (Santoso and England, 2004; Carter et al., 2009; Naveira Garabato et al., 2009; Sudre et al., 2011). Whilst some nutrient data (particularly negative Si$^+$; Supplement Section 1) may also hint at the presence of SAMW in the upper 150 to 250 m of Station 249, the water mass studies generally conclude that SAMW overlies AAIW only to the north of the SAF. Based on these considerations, the main water mass for Station 249 at 150 to 750 m depth is thus freshly ventilated AAIW, which is characterized by $\varepsilon^{114/110}\text{Cd} \approx +4$ and [Cd] $\approx 0.65$ nmol/kg. This conclusion is corroborated by previous investigations, which have observed fractionated N and Si isotope signatures for AAIW (and SAMW; Rafter et al., 2012; de Souza et al., 2012) and further supported by simple modeling (Fig. 7b), as discussed below.

A number of previous studies have shown that AAIW is formed by subduction of surface/mixed layer waters, with a chemical signature that is subsequently modified by mixing processes (e.g., Sievers and Nowlin, 1984; Santos and England, 2004). The slightly fractionated Cd isotope composition of AAIW is thus most likely a consequence of biological fractionation near the surface and moderation of this signal by mixing. Such a scenario is confirmed by calculations, which demonstrate (Fig. 7b) that the Cd composition inferred for AAIW can be obtained as a mixture of Weddell Gyre or ACC surface waters (although AAIW at Station 249 is sourced eastward of the Drake Passage; Naveira Garabato, 2009) and Southern Ocean deep water (from $\geq 1$ km depth). Given that our results show that a fractionated Cd isotope surface signature can be preserved at depth to a sufficient extent to be still recognizable in AAIW, it will be of interest for future studies to determine how far northward this isotopic signal is retained along the water mass flowpath.

6. Conclusions

The new Cd concentration and stable isotope data for surface seawater and three depth profiles from the Southern Ocean provide a number of important observations and interpretations:

(1) The data of this study and Abouchami et al. (2011) for seawater from the upper mixed layer define a broad but clear ‘HNLC trend’ in a plot of $\varepsilon^{114/110}\text{Cd}$ versus [Cd], which is primarily a consequence of isotopic fractionation associated with biological uptake. In comparison to surface water samples from non-HNLC locations, Southern Ocean surface water features relatively high Cd concentrations (typically from 0.2 to 0.6 nmol/kg) and only moderately fractionated $\varepsilon^{114/110}\text{Cd}$, generally between $+4$ and $+8$. These data reflect the limited biological productivity of the HNLC ocean. Detailed assessment reveals fine structure within the ‘HNLC trend’, which records differences in the biological fractionation factor, variable fractionation conditions (closed versus open system) or distinct compositions for the source water masses, or a combination of these factors.

(2) Southern Ocean seawater from depths $\geq 1$ km has an average $\varepsilon^{114/110}\text{Cd} = \pm 2.5 \pm 0.2$ (2σ, $n = 16$). This agrees well with published Cd data and the combined global dataset supports a relatively uniform deep water Cd isotope composition, with an average $\varepsilon^{114/110}\text{Cd}$ of $+3.0 \pm 0.3$ (2σ, $n = 27$) for samples from depths $\geq 1$ km.

(3) Significant differences in Cd isotope compositions were observed for seawater from intermediate depths. Samples from 200 m to 400 m depth in the Weddell Gyre have $\varepsilon^{114/110}\text{Cd}$ values as low as $+1$. These light isotope compositions most likely reflect the addition of remineralized Cd from biomass that records incomplete nutrient utilization at the surface. Samples from depths of 150 m to 750 m in the Drake Passage where AAIW is the dominant water mass, display relatively low [Cd] of $\sim 0.65$ nmol/kg and moderately fractionated $\varepsilon^{114/110}\text{Cd}$ of about $+4$. This Cd signature of AAIW is a consequence of biological Cd isotope fractionation near the surface and moderation of the biological signal by mixing, in accord with the formation history of this water mass.

In summary, our results demonstrate that coupled Cd isotope and concentration data provide valuable new insights into the dis-
tribution and cycling of Cd in the ocean. The highly systematic nature of the Cd isotope signatures may furthermore prove to be of utility for future research in marine geochemistry and paleoceanography.

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Appendix A. Supplementary material

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References


