Mutation of FOXC1 and PITX2 induces cerebral small-vessel disease

French, Curtis R.; Seshadri, Sudha; Destefano, Anita L.; Fornage, Myriam; Arnold, Corey R.; Gage, Philip J.; Skarie, Jonathan M.; Dobyns, William B.; Millen, Kathleen J.; Liu, Ting

Published in:
CLIN Journal

DOI:
10.1172/JCI75109

IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.

Document Version
Publisher's PDF, also known as Version of record

Publication date:
2014

Link to publication in University of Groningen/UMCG research database

Citation for published version (APA):

Copyright
Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

Take-down policy
If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Downloaded from the University of Groningen/UMCG research database (Pure): http://www.rug.nl/research/portal. For technical reasons the number of authors shown on this cover page is limited to 10 maximum.

Download date: 11-03-2019
Mutation of FOXC1 and PITX2 induces cerebral small-vessel disease


1Department of Ophthalmology, University of Alberta, Edmonton, Alberta, Canada. 2Department of Neurology and School of Public Health, Boston University, Boston, Massachusetts, USA. 3Institute of Molecular Medicine and School of Public Health, University of Texas Health Sciences Center, Houston, Texas, USA. 4Department of Biochemistry and Molecular Biology, University of Calgary, Calgary, Alberta, Canada. 5Department of Ophthalmology and Visual Sciences, University of Michigan Medical School, Ann Arbor, Michigan, USA. 6Department of Cell Biology, Neurobiology and Anatomy, Medical College of Wisconsin, Milwaukee, Wisconsin, USA. 7Department of Pediatrics and the Center for Integrative Brain Research, University of Washington, Seattle, Washington, USA. 8Department of Genetics, University of Groningen, Groningen, The Netherlands. 9Feinberg Cardiovascular Research Institute, Feinberg School of Medicine, Northwestern University, Chicago, Illinois, USA. 10Department of Genetics, University of Groningen, Groningen, The Netherlands. 11Department of Radiology and Diagnostic Imaging, Department of Biological Sciences, and 12Department of Medical Genetics, University of Alberta, Edmonton, Alberta, Canada.

Patients with cerebral small-vessel disease (CSVD) exhibit perturbed end-artery function and have an increased risk for stroke and age-related cognitive decline. Here, we used targeted genome-wide association (GWA) analysis and defined a CSVD locus adjacent to the forkhead transcription factor FOXC1. Moreover, we determined that the linked SNPs influence FOXC1 transcript levels and demonstrated that patients as young as 1 year of age with altered FOXC1 function exhibit CSVD. MRI analysis of patients with missense and nonsense mutations as well as FOXC1-encompassing segmental duplication and deletion revealed white matter hyperintensities, dilated perivascular spaces, and lacunar infarction. In a zebrafish model, overexpression or morpholino-induced suppression of foxc1 induced cerebral hemorrhage. Inhibition of foxc1 perturbed platelet-derived growth factor (Pdgf) signaling, impairing neural crest migration and the recruitment of mural cells, which are essential for vascular stability. GWA analysis also linked the FOXC1-interacting transcription factor PITX2 to CSVD, and both patients with PITX2 mutations and murine Pitx2-/- mutants displayed brain vascular phenotypes. Together, these results extend the genetic etiology of stroke and demonstrate an increasing developmental basis for human cerebrovascular disease.

Introduction

Stroke is a leading cause of morbidity and mortality, whose prevalence increases dramatically with age. Despite its substantial heritable basis, only a small number of causative genes have so far been identified, generally for severe early-onset phenotypes (cerebral autosomal dominant or recessive arteriopathy with subcortical infarcts and leukoencephalopathy: CADASIL [NOTCH3], CARASIL [HTRA1], and porencephaly [COL4A1]) (1–3). Such cases have revealed important pathways that contribute to stroke, including the roles of Notch and TGF-β signaling. In the same way, the vascular basement membrane’s contribution (COL4A1 and COL4A2) (3, 4) to juvenile stroke phenotypes further stimulated investigation of the cellular components (endothelial and mural cells) upon which brain vascular integrity depends. The demonstration that Notch signaling regulates pericyte numbers (5, 6) has in turn provided a mechanistic explanation for disorders such as CADASIL. These examples of juvenile stroke resulting from severe alterations in brain vascular development raise the intriguing possibility that milder changes contribute to late-onset disease and that a larger proportion of strokes have embryonic origins. It is therefore notable that the same genes regulate cerebral structural development and angiogenesis (7) and that the cell populations essential for cerebral vascular homeostasis (pericytes and vascular smooth muscle) are predominantly derived from the neural crest (8, 9).

The increasing prevalence of stroke exerts disproportionately severe effects on the quality of life of affected individuals and their families. Consequently, phenotypes predictive of future stroke merit investigation, with the goal of developing treatments targeting causative pathways and preventing a frequently preterminal disease. One such phenotype is cerebral small-vessel disease (CSVD), which represents a major risk factor for both ischemic and hemorrhagic stroke (10–13). Characterized by perturbed perforating end-artery function, CSVD results in lesions apparent on MRI that encompass white matter hyperintensities (WMHs), dilated perivascular spaces, microbleeds, and lacunar infarcts. These markers of cerebrovascular pathology provide opportunities for gene discovery and for defining the mechanisms that contribute to subsequent stroke.

Our study evaluated the hypothesis that the forkhead box transcription factor FOXC1, which patterns multiple organs including the CNS, contributes to CSVD. It was prompted by a higher incidence of self-reported stroke in some of our local pedigrees with FOXC1 mutations and supported experimentally by: (a) blood-stained hydrocephalus in murine Foxc1-/- mutants, (b) related zebrafish foxc1 morphant phenotypes, and (c) the exten-
sive involvement of Foxc1 in vascular development (14–16). The latter encompasses essential roles in arterial specification, angiogenesis regulation, endothelial lymphatic cell sprouting (17), as well as a requirement for Foxc1 in brain pericytes (18). Here, we demonstrate a role for FOXC1 in cerebrovascular disease through targeted genome-wide association (GWA) analysis, MRI of glaucoma patients with FOXC1-attributable Axenfeld-Rieger syndrome (ARS), and detailed zebrafish analyses.

Results and Discussion
We first performed a meta-analysis of GWA data for 500 kb encompassing FOXC1 on 6p25 in 9,361 individuals with brain MRI data from the Cohorts for Heart and Aging Research in Genomic Epidemiology (CHARGE) consortium. This identified 10 WMH-associated SNPs ($P = 0.0031–0.048$, Bonferroni-corrected) located in an intron of GMDS, which catalyzes GDP-mannose metabolism and lies adjacent to FOXC1 (Figure 1A and Supplemental Table 1; supplemental material available online with this article; doi:10.1172/JCI75109DS1). Analysis of 2 independent expression quantitative trait loci (eQTL) datasets (19, 20) demonstrated that 3 of these WMH-associated SNPs strongly influence FOXC1 transcript levels (study 1: $P = 2.96 \times 10^{-8}$–$5.82 \times 10^{-11}$; study 2: $P = 0.01$–0.008; Supplemental Table 1).

We next assessed whether patients with FOXC1-attributable ARS exhibited CSVD. Eighteen patients with either a FOXC1 mutation or copy number variation (CNV) (comprising missense
Consistent with these data, overexpression of zebrafish foxc1a or dual suppression of both paralogs (foxc1a and foxc1b) results in cerebral hemorrhage (overexpression 32 of 111 [28.8%], suppression 111 of 352 [31.5%]; Figure 1, I–K, and Supplemental Table 3). Notably, acellular perivascular spaces were evident on electron microscopy of foxc1 morpholino–treated embryos (Supplemental Figure 2). Foxc1 was expressed in the neural crest, and in morphants, there was aberrant migration of the cerebral neural crest from which most mural cells are derived (Supplemental Figures 3 and 4). Since platelet-derived growth factor (PDGF) signaling regulates neural crest recruitment to the developing vasculature, we evaluated this candidate pathway in foxc1 morphants. We observed reduced expression of both receptor tyrosine kinases (pdgfra and pdgfrb) in foxc1 morphants (Figure 2, A–H), positioning Pdgf signaling genetically downstream of Foxc1. Importantly, the prevalence of cerebral hemorrhage induced by morpholino inhibition of pdgfra alone, or pharmacological inhibition of pdgfra and pdgfrb, synergized with foxc1 morpholino inhibition (Figure 2, I–L). This is consistent with a model in which Foxc1 regulates vascular stability through the Pdgfra homodimer and Pdgfrβ heterodimer, either by the control of pdgfra and pdgfrb expression or indirectly as a consequence of aberrant neural crest migration and/or survival.

Since loss of neural crest–specific Pdgfα induces murine cerebral hemorrhage and irregular vascular smooth muscle cell coverage (25, 26), we predicted loss of such cells in foxc1 morphants. In keeping with the aberrant neural crest migration and increased cell death (Supplemental Figure 4), foxc1 morphants exhibited reduced numbers of neural crest cells associating with the cerebral vasculature at 32 hours post fertilization (hpf) (foxc1MO 48 ± 10, WT 61 ± 9; \( P = 0.01 \)) (Figure 3, A–C). Consistently, at 4 days post fertilization (dpf), this manifested as reduced numbers of vascular smooth muscle cells (foxc1MO 48 ± 10, WT 61 ± 9; \( P = 0.01 \)) (Figure 3, D–F). In contrast, we found that expression of markers for other vascular components (col4a1 and claudin5b) and endothelial cell numbers in both morphants and murine endothelial–specific Foxc1(+) mutants was unaltered (Supplemental Figure 5). Together, these data support a model of reduced neural crest–derived smooth
The contribution from altered neural crest function and substantially increase the proportion of strokes known to have developmental origins. Furthermore, the evidence presented here from Foxc1 and Pitx2 implicates other transcription factors with neural crest roles as candidates and thus provides practical opportunities for accelerating the identification of the molecular basis for stroke through integrated human genetic and zebrafish analyses. Our observation of a predominantly mural role for Foxc1 in the cerebral vasculature, which contrasts with an endothelial cell contribution systemically (16), also merits investigation and may correlate with the unique endothelial barrier properties of the cerebral circulation. From a clinical perspective, the substantial interval that elapses between the onset of MRI-detectable features of CSVD and the occurrence of stroke provides a therapeutic window for intervention, and potentially, patients with mutations involving neural crest genes may benefit from common stroke-prevention strategies. Finally, our findings have direct implications for Axenfeld-Rieger syndrome, a glaucoma subtype frequently associated with progressive visual decline despite surgical control of intraocular pressure (30). Evidence of a cerebral vasculopathy raises the possibility that perturbed vascular function contributes to the visual loss that has previously been attributed to optic nerve disease.

Figure 3. **foxc1** and **PITX2** regulate vascular smooth muscle cell numbers. Number of **sox10**-positive neural crest cells that associated with the cerebral vasculature in WT embryos at 32 hpf (**A**) was significantly reduced in **foxc1** morphants (**B** and **C**). Cerebral vascular mural cells expressed smooth muscle actin by 4 dpf (**D**), with fewer smooth muscle actin-positive cells observed in **foxc1** morphants (**E** and **F**). Patients with **PITX2**-attributable ARS exhibited CSVD. (**G**) WMH, (arrows) and (**H**) dilated perivascular spaces (arrowheads). Compared with WT embryos (**I**), murine **Pitx2**−/− mutants demonstrated reduced and discontinuous smooth muscle actin staining of large and small cerebral vessels (**J**). Original magnification, ×200 (**A** and **B**), ×100 (**D** and **E**), and ×400 (**I** and **J**).
Methods
Statistics. GWA data are presented as the −log10 value, with Bonferroni’s correction for the number of independent comparisons determined via the number of linkage disequilibrium blocks. For analysis of cell numbers and genetic interactions, unpaired 2-tailed Student’s t tests were used to assess significance. Analyses are displayed graphically as the mean ± SEM. All experiments were conducted in triplicate. A P value of less than 0.05 was considered statistically significant.

Study approval. Ethical approval was provided by the University of Alberta Health Research Ethics Board, with written informed consent received from all participants prior to their inclusion in the study. Animal experiments were approved by the IACUC of the University of Alberta, the University of Michigan, and Northwestern University.

Further details regarding the methods are available in the Supplementary Methods.

Acknowledgments
We are grateful to the patients who participated in this study. We thank Brian Link (Medical College of Wisconsin), Michael Walter (University of Alberta), PeterCarlsson (University of Gothenburg), and Alison Hardcastle (Institute of Ophthalmology, United Kingdom) for very helpful advice and reagents; Peter Seres and colleagues at the Peter S. Allen MR Research Centre for MR imaging; Aleah McCorry for zebrafish husbandry; and Wei Dong and the University of Calgary Microscopy and Imaging Facility for help with transmission electron microscopy. Funding was provided by National Eye Institute/NIH grants EY014126 and EY007003 (to P.J. Gage) and R01 EY019484 (to T. Kume); the Canadian Institutes of Health Research (CIHR) (MOP-133658, to O.J. Lehmann and MOP-114902, to S.J. Childs); the Natural Sciences and Engineering Research Council (to A.J. Waskiewicz); and a Heart and Stroke Foundation of Canada postdoctoral fellowship (to C.R. French).

C.R. French is the recipient of an American Society of Human Genetics Charles Epstein Trainee Award. C.R. Arnold is the recipient of a CIHR Training Grant in Genetics, Child Health and Development studentship.

Address correspondence to: Ordan J. Lehmann, Departments of Ophthalmology and Medical Genetics, 829 Medical Sciences Building, University of Alberta, Edmonton, AB T6G 2H7, Canada. Phone: 001.780.492.8550; E-mail: olehmann@ualberta.ca.