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Eclampsia & preeclampsia

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Introduction



Introduction

Preeclampsia is a pregnancy induced disease, characterized by the occurrence of hypertension (systolic \geq 140 mmHg and/or diastolic \geq 90 mmHg) and proteinuria in the second half of pregnancy in a previously normotensive woman. It complicates 5 to 7% of pregnancies in the United States.¹ It is a systemic disease of which the exact pathophysiology remains to be elucidated.

Preeclampsia is a systemic disease that can affect several organs including the liver, the kidneys and the brain. Involvement of the brain can lead to one of the most feared complications of preeclampsia: eclampsia. This is the onset of seizures or coma in a preeclamptic woman not attributable to any other cause. Accompanying symptoms are headache, visual disturbances (cortical blindness, scotoma, photopsia), photophobia, sonophobia, nausea, vomiting and altered mental status.² It is a life threatening disease for both mother and child and in the Netherlands cerebral complications such as eclampsia are responsible for most maternal deaths.³

The incidence of eclampsia in the Netherlands is 6.2 per 10,000 deliveries. The number of maternal deaths due to eclampsia is 1 in 74, fatality rate 1.4%. Between 1993 and 2005 the maternal mortality rate of (pre)eclampsia in the Netherlands was 3.5 per 100,000 live births. The major mode of maternal death due to hypertensive diseases is a cerebrovascular complication in 71% of the cases. In other European countries the incidence of eclampsia is ranges from 2.4 to 5.0 per 10,000 deliveries and ranging from 71 to 173 per 10,000 deliveries in developing countries. The mortality rate from eclampsia in the UK is 1.8% in the USA 0.5% and ranging from 6.0-8.0% in developing countries.

Why women with preeclampsia are susceptible to brain involvement is not clear. In addition, how pregnancy affects the brain is just being elucidated. Therefore, in our labs we investigate the brain during pregnancy in animal models and in patient studies we investigate the brain several years after a pregnancy complicated by eclampsia or preeclampsia.

Pathophysiology of eclampsia

There are two major theories on the pathophysiology of eclampsia, both based on failure of the cerebral autoregulation. One is the 'vasculopathy' theory which suggests that there is an 'overregulation' of the cerebral blood flow because previously it was thought that the neurologic symptoms of eclampsia were caused by ischemia. The theory behind this was that in reaction to acute hypertension the cerebral vasculature constricted too severely (overregulation of the cerebral blood flow) causing hypoxia and ischemia.

Evidence favoring this theory was the vasospasm seen on cerebral angiograms in some women with eclampsia. 12,13 Later newer imaging techniques showed presence of vasogenic edema, making ischemia as the only cause unlikely. More recently, Bartynski developed a more subtle theory based on the vasculopathy theory. 14 He hypothesizes that systemic toxicity in patients with (pre)eclampsia such as immune system activation, endothelial cell activation and injury, vascular instability and organ hypoperfusion, are associated with increased vasoconstriction of cerebral vasculature. The presence of systemic hypertension poses an additional trigger for vasoconstriction, leading to cerebral hypoperfusion and hypoxia. Prolonged hypoxia stimulates vascular endothelial growth factor (VEGF) release and activation of endothelial cells. Together, this may result in increased permeability of endothelium followed by vasogenic edema. 14 In support of this theory is the fact that vasogenic edema is typically found in the watershed zones. However, the suggestion that the cerebral vasculature would overregulate and constrict to such an extent to cause ischemia, seems unlikely. Therefore, we consider the second and currently more popular theory that focuses on loss of cerebral autoregulation, more plausible. In this theory it is hypothesized that in the presence of endothelial dysfunction an acute elevation of blood pressure exceeds the upper limit of the cerebral autoregulation leading to forced dilatation of cerebral arteries (Figure 1). Increased cerebral blood flow subsequently results in disruption of the blood brain barrier followed by extravasation of water and plasma solutes and formation of vasogenic edema. 15,16 Once the blood pressure decreases within normal limits of cerebral autoregulation, the vasogenic edema resolves and the neurological symptoms disappear. 17,18 Loss of cerebral autoregulation based on acute hypertension is called hypertensive encephalopathy. Eclampsia may be a form of hypertensive encephalopathy 19-21, however, not all patients with eclampsia have blood pressures high enough to reach the upper limit of cerebral autoregulation and 10% - 16% of eclamptic patients do not even become hypertensive. 10,11 In addition, a clinical and radiological condition similar to eclampsia has been recognized and is thought to be identical to eclampsia although the underlying triggering factor differs. This syndrome was first recognized by Hinchey et al. in 1996 and named 'reversible posterior leucoencephalopathy syndrome'. 17 Later, different names were introduced of which 'posterior reversible encephalopathy syndrome' (PRES) is currently most popular. 22,23

Several aspects of the cerebral vasculature and its regulating mechanisms play an important role in the pathophysiology of eclampsia or PRES. In the following paragraphs the most important mechanisms of the cerebral vasculature will be discussed. These are cerebral autoregulation, the blood-brain barrier and perivascular innervation. The current knowledge about how these aspects are influenced by pregnancy will be discussed as well.

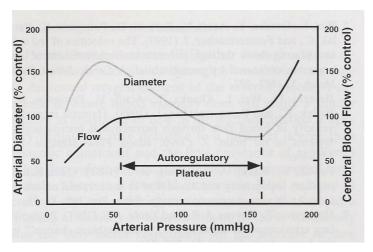


Figure 1 The cerebral autoregulation curve, adapted from Chillion and Baumbach 1997 and used with the permission of Elsevier Limited for Academic Press. 25

Cerebral autoregulation

The term 'autoregulation' in relation to the cerebral circulation, was first introduced by Lassen in 1959.²⁴ Autoregulation of blood flow ensures that the blood flow through an organ is maintained at a relatively constant level despite changes in perfusion pressure or arterial blood pressure. Within the limits of cerebral autoregulation this is called the autoregulatory plateau (Fig 1), which ranges from approximately 50-60 to 150-160 mmHg in the healthy human.²⁵ This plateau is brought about by changes in the arterial diameter with vasoconstriction when blood pressure increases and vasodilatation when blood pressure decreases. When the mean arterial pressure is not within this range, autoregulation may be lost. In case of acute severe hypertension, such as in most cases of eclampsia, the cerebral arterial pressure exceeds the upper limit of autoregulation and cerebral blood flow increases linearly with increase in blood pressure.^{26,27} Subsequently, the blood-brain barrier is disrupted and vasogenic edema develops.^{15,16} When the blood pressure decreases, the cerebral blood flow normalizes and the edema resolves.¹⁷⁻¹⁹

Cerebral autoregulation is regulated by several control mechanisms.^{28,29} One of those is the myogenic mechanism by which the small vessels react with constriction or vasodilation in response to changes in transmural pressure.^{30,31} Secondly, there are metabolic mechanisms by which changes in metabolic demand of local brain areas during neuronal activity alters the cerebral blood flow in this area.³² Thirdly, endothelium derived factors can regulate cerebral blood flow by exerting a constrictive or relaxing effect on the vascular smooth muscle. Vasodilatory factors are nitric oxide (NO), endothelium derived hyperpolarization factor (EDHF) and prostacylin, constrictor factors are thromboxane A₂,

prostaglandin $F_{2\alpha}$ and endothelin-1.³³ Finally, perivascular nerve activity may control cerebral autoregulation, which will be described below.

Chronic hypertension causes a right-shift of the cerebral autoregulation curve.³⁴ This is a protective effect, since loss of cerebral autoregulation on the upper end of the curve now occurs at higher pressures.^{35,36} This right shift is caused by hypertensive remodeling and vascular hypertrophy, which normalize wall stress by increasing wall thickness and decreasing vascular diameter.³⁷⁻³⁹ Stimulation of sympathetic innervation also results in a shift of the upper limit of the autoregulation plateau to higher pressures and has a protective effect on the blood-brain barrier.⁴⁰⁻⁴² In contrast, acute sympathetic denervation causes lowering of the upper limit of cerebral autoregulation.⁴³ During normal conditions the effect of sympathetic innervation on resting cerebral blood flow is minimal.⁴⁴

Although progression is being made in this field, whether and how pregnancy influences the cerebral autoregulation has not exactly been elucidated. Some eclamptic women never reach blood pressures considered to be in the hypertensive range and therefore they do not necessarily reach the upper limit of cerebral autoregulation as this has been established in nonpregnant individuals. Thus, it seems likely that pregnancy shifts the upper limit or even the entire curve to the left and that hyperperfusion may occur at lower blood pressures compared to nonpregnant women. In vitro studies in late-pregnant rats showed that myogenic reactivity of posterior cerebral arteries was decreased and that the pressure at which forced dilatation occurs was lower in late-pregnant animals. As An in vivo study did not show a decrease of the upper limit of autoregulation in pregnant rats; there was no difference between late-pregnant and nonpregnant rats. However, cerebral edema was more extensive in the late-pregnant animals after autoregulation breakthrough, suggesting that the blood-brain barrier is more vulnerable to disruption during pregnancy.

Blood-brain barrier

In the cerebral circulation, one of the most obvious and significant specializations is the formation of the blood-brain barrier by cerebral capillary endothelial cells. The endothelial cells in the brain's vasculature form high-resistance tight junctions through which there is no paracellular transport of water or solutes. Also, endocytosis (transcellular transport) is minimal in capillaries of the central nervous system. These features of the cerebral endothelium are considered the blood-brain barrier and protect the brain from formation of vasogenic edema. However, acute increased hydrostatic pressure may disrupt the blood-brain barrier with subsequent vasogenic edema formation.

It is important to understand how pregnancy influences the blood-brain barrier considering the fact that cerebral vasogenic edema is present in eclamptic and some preeclamptic patients. ^{49,50} In isolated cerebral arteries from rats, the vessels from late pregnant animals show increased permeability of the blood-brain barrier by increased endocytosis and paracellular transport when hydrostatic pressure is increased and forced dilatation occurs. ^{51,52} In an in vivo study, permeability to Evan's blue was increased after autoregulation breakthrough in late pregnant rats but there was no increase in permeability to sodium fluorescein. ⁵³ Evan's blue permeability was greater in the posterior versus the anterior cerebrum suggesting regional differences in vulnerability of the blood-brain barrier, which may be an explanation for the regional distribution of vasogenic edema in eclampsia. Together, these findings suggest that pregnancy alters the permeability of the blood-brain barrier which may be important in case of acute hypertension.

Perivascular innervation

Cerebral vessels are associated with parasympathetic, sympathetic and sensory or trigeminal fibers.²⁸ They originate in cranial ganglia and distribute mainly to extraparenchymal vessels.²⁹ The autonomic neurons run through the adventitial layer of cerebral arteries in pial vessels ending in preterminal axons and terminals proper, which come in close contact with the outer smooth muscle layer of the vessel media.²⁹ This is the vasomotor innervation of both extracerebral and intracerebral small arteries and arterioles. Under normal resting conditions, perivascular innervation has little or no effect on cerebral autoregulation.^{26,54,55} When the steady state of the cerebral autoregulation is altered such as in acute hypertension^{55,56} or ischemia/reperfusion^{57,58}, perivascular innervation may influence the cerebral blood flow. However, the exact role on the regulation of cerebral autoregulation is controversial because of its complex nature.⁵⁸

Cerebral vessels are innervated by an extensive sympathetic nerve supply.⁵⁹ In general, anterior vessels in the circle of Willis receive a denser sympathetic nerve supply than those in the posterior circulation.⁵⁹ Most of the pial arterioles are extrinsically innervated with sympathetic nerves originating from the cervical sympathetic ganglia.⁶⁰⁻⁶² Small pial vessels may receive intrinsic noradrenergic nerve fibers from the pons (locus ceruleus).⁶³ Under resting conditions, experimental manipulation of sympathetic input generally has little or no effect on cerebral blood flow in all the species examined, including humans.²⁹ However, during acute hypertension sympathetic nerves play a role in protecting the brain from autoregulation breakthrough and loss of integrity of the blood-brain barrier.^{64,65} Stimulation of sympathetic nerves during acute hypertension attenuates the increase in cerebral blood flow.^{42,55} In chronic hypertension sympathetic nerves exert

a trophic effect on cerebral vessels and contribute to hypertrophy of vascular muscle hereby protecting the downstream vessels.⁶⁶ The posterior brain is the primary site where autoregulation is overcome during acute hypertension and thus the primary site of vasogenic edema formation in eclampsia. One hypothesis is that this is because of the lesser sympathetic innervation posteriorly compared to the anterior cerebral vasculature, however, this hypothesis has not been proven.

Parasympathetic innervation plays a less well defined role in cerebral autoregulation. Parasympathetic fibers exert a dilator role, for example in ischemia/reperfusion. In addition, cholinergic nerves interact with presynaptic noradrenergic nerve terminals and modify neurotransmitter release, for example by reducing the constrictor effects of sympathetic stimulation. Se

Trigeminal innervation is mostly important in nociception, but also plays a role in cerebral vasodilation during cortical spreading depression ^{67,68}, post ischemia/reperfusion ⁵⁷ and on the lower end of the cerebral autoregulation curve. ⁶⁹ Because of its role in migraine ⁷⁰⁻⁷⁴ — a disease that shares features with eclampsia ⁷⁵⁻⁷⁷ — and in vascular adaptation during pregnancy, the sensitivity and perivascular innervation of the trigeminal neuropeptide calcitonin gene-related peptide (CGRP) in cerebral arteries during pregnancy are a subject of this thesis.

Calcitonin Gene-Related Peptide

Calcitonin gene-related peptide (CGRP) is a neuropeptide of the sensory or trigeminal nervous system. ^{29,58,78} It is involved in modulation of peripheral vascular resistance ⁷⁹⁻⁸¹, attenuation of gastric acid production 82 and nociception.83 There are two isoforms of CGRP present, which are the α - and β -CGRP. The α -CGRP is a 37-amino-acid peptide produced by tissue-specific alternative splicing of the calcitonin/αCGRP RNA transcript. 78 It is located mostly at the central nervous system.⁸⁴ β-CGRP is produced from a different gene exclusive of alternative splicing and is mainly located in the enteric nervous system. 84 CGRP is co-located with substance P and neurokinin A in perivascular nerves where it transmits sensory information to the central nervous system, for example noxious stimuli, and where it serves a regulatory function of the local environment through the release of neurotransmitters. 29 CGRP containing perivascular nerves are found in many vascular beds throughout the entire body. CGRP is the strongest endogenous vasodilator known, in which the α -isoform has a stronger effect compared to β -CGRP. The receptor that is identified as the CGRP receptor, is the calcitonin receptor-like receptor (CRLR). It is a Gprotein coupled receptor and it requires binding to receptor activity modifying protein 1 (RAMP1) in order to become receptive to CGRP. 84,85 The signaling pathway can be either endothelium-dependent, endothelium-independent or both, differing per tissue bed and

species.⁸⁷ In the endothelium-dependent manner, CGRP can activate nitric oxide synthase (NOS), thereby releasing nitric oxide, causing vascular smooth muscle cell (VSMC) relaxation.^{73,88} In the endothelium-independent pathway, CGRP binds directly on the CRLR/RAMP1 complex on the VSMC.^{73,89} This activates adenyl cyclase, which in turn increases cAMP levels, causing VSMC relaxation.⁷³

During pregnancy, levels of plasma CGRP increase to term and drop after delivery 90,91, suggesting that CGRP plays a role in the vascular adaptations that occur during pregnancy, when the plasma volume expands with 40% and blood pressure remains normal. In addition, plasma CGRP levels appear to be lower in the plasma of women whose pregnancy was complicated by preeclampsia compared to normotensive control women. 91 Plasma CGRP levels are also higher in pregnant rats compared to nonpregnant rats and the levels drop after delivery. 92 The vascular sensitivity to CGRP is greater in pregnant rats and ovariectomized rats treated with female sex steroid hormones compared to ovariectomized rats and male rats.^{89,93} Administration of CGRP in pregnant rats treated with the NOS inhibitor L-NAME to cause preeclampsia-like features resulted in lowering of blood pressure, less fetal death and increased birth weight. 4 When the CGRP antagonist CGRP₈₋₃₇ was administered in L-NAME treated pregnant animals, blood pressure raised even further while there was no effect on blood pressure of untreated animals.95 This effect was not associated with an increase of CGRP mRNA in dorsal root ganglia, suggesting that the underlying mechanism is an increased vascular sensitivity to CGRP during pregnancy. Together these findings suggest an important role of CGRP in maternal cardiovascular adaptations during pregnancy and that possibly CGRP or vascular sensitivity to CGRP is involved in the pathogenesis of preeclampsia.

Posterior Reversible Encephalopathy Syndrome

The occurrence of PRES is associated with several different conditions and progressively recognized in more and more different underlying pathologies. Most common patient categories are preeclampsia/eclampsia, patients treated with immunosuppressants or chemotherapy (e.g. cyclosporine and tacrolimus), malignant hypertension, infection/sepsis, solid organ or bone marrow transplantation and several autoimmune diseases. The syndrome occurs at all ages, including the pediatric population. There are no strict criteria, but diagnosis is made based on clinical findings and imaging. Symptoms of this syndrome are similar to eclampsia and include headache, nausea and vomiting, decreased alertness, altered mental functioning, seizures and visual disturbances such as blurred vision or cortical blindness. These symptoms occur usually in conjunction with an acute elevation of the blood pressure. Computed tomography (CT) shows localized hypodense lesions at the gray white matter junction or widespread

diffuse edema. However, occasionally focal areas of edema are beyond the resolution of CT scan, in which case magnetic resonance imaging (MRI) is necessary. ¹⁰¹ Findings on MRI are consistent with cerebral edema and include hypointensities on T1 sequence and hyperintensities on T2 and fluid attenuation inversion recovery (FLAIR) sequences. ^{17,98} The areas that are hyperintense on T2/FLAIR imaging are iso- or hypointense on diffusion weighted imaging (DWI) and hyperintense on apparent diffusion coefficient (ADC). This pattern of MRI abnormalities is consistent with vasogenic edema. The distribution of the edema on MRI is typically in the subcortical white matter of the parieto-occipital lobes and appears symmetrically. Although the name of the syndrome suggests indeed this kind of distribution, there is a wide variety. Atypical findings include edema in the frontal lobes, the inferior temporal-occipital junction, the cerebellum, the basal ganglia and brainstem. 97,98,102,103 Also, in some cases, the grey matter can be involved, the edema may be asymmetric or accompanied by hemorrhage. Moreover, DWI may demonstrate small areas of cytotoxic edema within lesions of vasogenic edema in patients with eclampsia. 104,105 In those instances it has been suggested that vasogenic edema in PRES can progress to such an extent that regional perfusion pressure decreases and blood flow decreases to ischemic levels leading to cytotoxic edema and infarction. ^{104,105} Imaging of obstetric patients with PRES (eclampsia and some cases of preeclampsia) is not distinct from other causes of PRES. In one study 106 the obstetric patients demonstrated more often involvement of the basal ganglia, however, this was not found in a larger study by Fugate et al. 98 Except for patients with hypertensive encephalopathy, hypertension is not always clearly present in all patients with PRES; 6 - 16% of PRES patients do not reach blood pressures commonly referred to as being in the hypertensive range. 11,97

An abrupt rise in blood pressure undoubtedly contributes to the development of PRES but the exact underlying mechanism that causes disruption of the blood-brain barrier in this wide variety of patient categories, has not been elucidated. The conditions associated with PRES are typically systemic processes, which have some degree of endothelial dysfunction and an inflammatory response. Possibly, these conditions (and other unidentified processes) predispose to an increased vulnerability of the cerebral vasculature to loss of cerebral autoregulation. This may be because increased permeability of the blood-brain barrier, a shift of the cerebral autoregulation curve to the left or both.

Animal models of (pre)eclampsia.

Preeclampsia and eclampsia are diseases that do not naturally occur in animals other than primates. ¹⁰⁷ The ideal model of preeclampsia should include preferentially as many of the clinical and laboratory features of the disease as possible and progress to eclampsia-like symptoms if severe. However, such a model does not exist and therefore different animal

models of preeclampsia have been designed, some expressing the preeclamptic features more than others. Podjarny et al. gave a nice overview of animal models for preeclampsia including reduction of arterial blood flow to the uterus and placenta by aorta or ovarian artery clipping, chronic NOS inhibition with L-NAME administration or in an endothelial-NOS knock-out model, sympathetic nervous and/or renin-angiotensin system overactivation, inflammatory models with pro-inflammatory cytokine injection or injection of low-dose lipopolysaccharide, insulin resistance and models of angiogenesis antagonism with soluble fms-like tyrosine kinase 1 (sFlt1).¹⁰⁷

Only a few studies focused on the brain when investigating preeclampsia models. Several models of hypertensive encephalopathy exist and have been used during pregnancy. 45,47 The systemic blood pressure can be increased acutely by administration of a pressor agent which causes loss of cerebral autoregulation. This has been done in our lab while cerebral blood flow was measured indirectly by laser Doppler⁴⁷ and with measuring microsphere content in different brain areas after autoregulation breakthrough. 19,108 Kanayama et al. described a rat model in which the celiac ganglion is stimulated with lipopolysaccharide in pregnant animals after which a preeclamptic and HELLP syndrome-like condition develops (Hemolysis, Elevated Liver enzymes, Low Platelets). 109 In addition, seizures occur, cerebral blood flow increases and cerebral edema develops. 110 Another model used in our lab is that of the Dahl salt-sensitive rat. 45 This rat becomes hypertensive when fed with a diet containing a high salt percentage (8%). At high blood pressures the Dahl salt-sensitive rat demonstrates symptoms similar to seizures of hypertensive encephalopathy: rhythmic, abrupt movements of the head in an up-an-down motion often associated with a lateral deflection or repetitive forearm flexion unilaterally.¹¹¹ There is evidence for disruption of the blood-brain barrier and edema formation in the brains of these rats as well as linearly decreasing myogenic reactivity with the duration of the high salt diet. 111 These rats also become hypertensive when fed a high salt diet during pregnancy¹¹² and demonstrate elevated markers of oxidative stress^{113,114}, both features of human preeclampsia. Furthermore, Dahl salt-sensitive rats suffer endothelial dysfunction, likely due to the salt-induced hypertension. ¹¹⁵ This model has one disadvantage: the Dahl salt-sensitive rats become symptomatic after 2.5 weeks of high salt diet, while the duration of pregnancy in rats is 3 weeks. However, it seems a good model for eclampsia and is used in Chapter 5.

Pressurized arteriograph system

The method that was used to investigate the effect of pregnancy, hypertension and different neurotransmitters on cerebral arteries *in vitro* in Chapters 5 and 6 is the pressurized arteriograph system. This system was developed at the University of Vermont,

United States, and is used extensively throughout the world to investigate vascular structure and function of small arteries and arterioles. In this experimental system, it is possible to investigate the effect of different pharmacological agents and physical forces such as pressure on the vascular smooth muscle and endothelium. The arteriograph that was used was a dual chambered system with two 20mL baths containing physiologic salt solution that is circulated to maintain temperature, oxygen, carbon dioxide and pH at physiologic levels. The carefully dissected vessels are mounted on two glass cannulas with nylon ties. The distal end of both of the cannulas is closed off to maintain pressure that is generated by a servo system that is connected to the proximal cannula. The servo system consists of a miniature peristaltic pump, an in-line pressure transducer and a controller. Through an optical window in the bath, the vessels are imaged by an inverted microscope that is connected to a video camera and a monitor. The video dimension analyzer (VDA) is used to analyze the signal obtained from the video image and to continuously register lumen diameter and wall thickness. The dynamic responses of vessel diameter and the intraluminal pressure are visualized on a computer by a serial data acquisition system that registers the VDA and pressure controller output, similar to a chart recorder. The cerebral autoregulation is subject to many different influences that cannot all be mimicked in an in vitro experiment. However, the use of the arteriograph system gives detailed insight into myogenic activity and passive structural properties of small cerebral arteries in response to some of the control mechanisms of autoregulation such as neurotransmitters and transmural pressure.

The maternal brain following eclampsia

It has long been thought that when a woman with eclampsia survives this condition without the occurrence of cerebral haemorrhage, she will fully recover. This is plausible, because when the blood pressure decreases, cerebral vasogenic edema resolves. However, in addition to vasogenic edema also cytotoxic edema has been found during the acute phase of eclampsia. Six to eight weeks post partum approximately one fourth of these formerly eclamptic women showed white matter lesions consistent with gliosis. How these lesions in formerly eclamptic women develop over life and their clinical relevance are unknown and are subject of this thesis. Moreover, some women with severe preeclampsia also demonstrated evidence of PRES on cerebral imaging even without experiencing eclamptic seizures and an increased risk of stroke in formerly preeclamptic women has been reported. Therefore, also formerly preeclamptic women are subject of neuroimaging and cognitive testing in this thesis.

Aims of this thesis

Part I (patient studies)

- To assess the long term consequences of preeclampsia and eclampsia on daily life with regard to cognitive function (**Chapter 2**)
- To provide insight into the long term consequences of eclampsia on the maternal brain with MR imaging (**Chapter 3**)
- To investigate the prevalence and severity of cerebral white matter lesions several years after preeclampsia and to find factors that are associated to these lesions (Chapter 4)

Part II (animal studies)

- To investigate whether decreased myogenic reactivity and the lack of hypertensive remodelling in cerebral arteries that occurs during pregnancy is due to the type of hypertension or to pregnancy (Chapter 5)
- To investigate the changes in cerebral perivascular innervation in different pregnancy-related states and the effect of gender on perivascular innervation (Chapters 5 and 6)
- To investigate the effect of pregnancy and gender on the sensitivity of cerebral arteries to sympathetic and trigeminal neurotransmitters (**Chapter 6**)

References

- Wagner LK. Diagnosis and management of preeclampsia. Am Fam Physician 2004;70:2317-2324
- 2. Shah AK, Rajamani K, Witty JE. Eclampsia: a neurological perspective. J Neurol Sci 2008;271:158-167
- Schutte JM, Schuitemakers NWE, van Roosmalen J, Steegers EAP on behalf of the Dutch Maternal Mortality Committee. Substandard care in maternal mortality due to hypertensive disease in pregnancy in the Netherlands. BJOG 2008;115:732-736
- 4. Zwart JJ, Richters A, Öry F, de Vries JIP, Bloemenkamp KWM, van Roosmalen J. Eclampsia in the Netherlands. Obstet Gynecol 2008;112:820-827
- Schutte JM, Steegers EA, Schuitemakers NWE, Santema JG, de Boer K, Pel M, Vermeulen G, Visser W, van Roosmalen J. Netherlands Maternal Morgality Committee. Rise in maternal mortality in the Netherlands. BJOG 2010;117:399-406
- 6. Andersgaard AB, Herbst A, Johansen M, Ivarsson A, Ingemarsson I, Langhoff-Roos J, Henriksen T, Straume B, Oian P. Eclampsia in Scandinavia: incidence, substandard care and potentially preventable cases. Acta Obstet Gynecol Scand 2006;85:929-936
- 7. Ekholm E, Salmi MM, Erkkola R. Eclampsia in Finland in 1990-1994. Acta Obstet Gynecol Scand 1999;78:877-882
- 8. Urassa DP, Cardstedt A, Nyström L, Massawe SN, Lindmark G. Eclampsia in Dar es Salaam, Tanzenia Incidence, outcome and the role of antenatal care. Acta Obstet Gynecol Scand 2006;85:571-578
- 9. Miguil M, Chekairi A. Eclampsia, study of 342 case. Hypertens Pregnancy 2008;27:103-111
- 10. Douglas KA, Redman CWG. Eclampsia in the United Kingdom. BMJ 1994;309:1395-1400
- 11. Mattar F, Sibai BM. Eclampsia VIII. Risk factors for maternal morbidity. Am J Obstet Gynecol 2000;182:307-312
- Trommer BL, Homer D, Mikhael MA. Cerebral vasospasm and eclampsia. Stroke 1988;19:326-329
- 13. Sengar AR, Gupta RK, Dhankuna AK, Roy R, Das K. MR imaging, MR angiography, and MR spectroscopy of the brain im eclampsia. AJMR 1997;18:1485-1490
- 14. Bartynski WS. Posterior reversible encephalopathy syndrome, part 2: controversies surrounding pathophysiology of vasogenic edema. Am J Neuroradiol 2008;29:1043-1049
- 15. Sokrab TEO, Johansson BB, Kalimo H, Olsson Y. A transient hypertensive opening of the blood-brain barrier can lead to brain damage. Acta Neuropathol. (Berl.) 1988:75:557-565.
- 16. Westergaard E, van Deurs B, Bronsted HE. Increased vesicular transfer of horseradish peroxidase across cerebral endothelium, evoked by acute hypertension. Acta Neuropathol 1977:37:141-15
- Hinchey J, Chaves C, Appignani B, Breen J, Pao L, Wang A, et al. A reversible posterior leukoencephalopathy syndrome. N Engl J Med 1996;334:494-500
- 18. Striano P, Striano S, Tortora F, De Robertis E, Palumbo D, Elefante A, Servillo G. Clinical spectrum and critical care management of posterior reversible encephalopathy syndrome (PRES) Med Sci Mont 2005;11:CR549-553

- Schwartz RB, Jones KM, Kalina P, Bajakian RL, Mantello MR, Garada , Holman BL.
 Hypertensive encephalopathy: findings on CT, MR imaging and SPECT imaging in 14 cases.
 Am J Radiol 1992;159:379-383
- 20. Donaldson JO. Eclamptic hypertensive encephalopathy. Sem Neurol 1988;8:230-33
- 21. Cipolla MJ. Cerebrovascular function in pregnancy and eclampsia. Hypertension 2007;50(1):14-24
- 22. Stott VL, Hurrell MA, Anderson TJ. Reversible posterior leukoencephalopathy syndrome: a misnomer reviewed. Intern Med J 2005;35:83-90
- Casey SO, Sampaio RC, Michel E, Truwit CL. Posterior reversible encephalopathy syndrome: utility of fluid-attenuated inversion recovery MR imaging in the detection of cortical and subcortical lesions. Am J Neuroradiol 2000;21:1199-206
- Lassen NA. Cerebral blood flow and oxygen consumption in man. Physiol Rev 1959;39:183 238
- Chillon JM, Baumbach GL. Autoregulation of Cerebral Blood Flow. In: Primer on Cerebrovascular Disease. Ch. 13, pp. 5154. Welch KMA, Reis DJ, Caplan LR, Seijo BK, and Weir B, editors. Academic Press, 1997
- 26. Strandgaard S, MacKenzie ET, Sengupta D, Rowan JO, Lassen NA, Harper AM. Upper limit of autoregulation of cerebral blood flow in the baboon. Circ Res 1974;34:435-440
- 27. MacKenzie ET, Strandgaard S, Graham DI, Jones JV, Harper AM. Effects of acutely induced hypertension in cats on pial arteriolar caliber, local cerebral blood flow, and the blood-brain barrier. Circ Res 1976:39:33-41
- Paulson, OB, Strandgaard S, Edvinsson L. Cerebral Autoregulation. Cerebrovascular and Brain Metabolism Reviews 1990;2:161-192
- Edvinsson L, Krause DN. Cerebral blood flow and metabolism. Second edition, 2002,
 Lippincott Williams & Wilkins, Philadelphia PA, USA
- 30. Halpern W, Osol G. Influence of transmural pressure on myogenic responses of isolated cerebral arteries of the rat. Ann Biomed Eng 1985;15:287-293
- 31. Osol G, Halpern W. Myogenic properties of cerebral blood vessels from normotensive and hypertensive rat. Am J Physiol 1985;249:H914-H921
- 32. Kuchinsky W, Wahl M. Lochal chemical and neurogenic regulation of cerebral vascular resistance. Physiol Rev 1978;58:656-689
- 33. Anderesen J, Shafi NI, Bryan RM. Endothelial influences on cerebrovascular tone. J Appl Physiol 2006;100:318-327
- 34. Strandgaard S, Olesen J, Skinhøj E, Lassen NA. Autoregulation of brain circulation in severe arterial hypertension. Br Med J 1973;1: 507–510
- 35. Mueller SM, Heistad DD. Effect of chronic hypertension on the blood-brain barrier. Hypertension 1980;2:809-812
- 36. Heistad DD. Protection of the blood-brain barrier during acute and chronic hypertension. Federation Proc 1984;43:205-209
- Baumbach GL, Heistad DD. Cerebral circulation in chronic arterial hypertension.
 Hypertension 1988;12:89-95
- 38. Baumbach GL, Heistad DD. Remodeling of cerebral arteries in chronic hypertension. Hypertension 1989:13:968-972
- Heistad DD, Baumbach GL. Cerebral vascular changes during chronic hypertension: good guys and bad guys. J Hypertens 1992;10:S71-S75

- 40. Bill A, Linder J. Sympathetic control of cerebral blood flow in acute arterial hypertension. Acta Physiol Scan 1976;96:114-121
- 41. Gross PM, Heistad DD, Strait MR, Marcus ML, Brody MJ. Cerebral vascular responses to physiological stimulation of sympathetic pathways in cats. Circ Res 1979;44:288-294
- 42. Heistad DD, Marcus ML. Effect of sympathetic stimulation on permeability of the blood-brain barrier to albumin during acute hypertension in cats. Circ Res 1979;45:331-338
- 43. Sadoshima S, Fujishima M, Yoshida F, Ibayashi S, Shiokawa O, Omae T. Cerebral autoregulation in young spontaneously hypertensive rats: Effect of sympathetic denervation. Hypertension 1985;7:392-397
- 44. Edvinsson L, MacKenzie ET. Amine mechanisms in the cerebral circulation. Pharmacol Rev 1977:28:275-348
- 45. Aukes AM, Vitullo L, Zeeman GG, Cipolla MJ. Pregnancy prevents hypertensive remodeling and decreases myogenic reactivity in posterior cerebral arteries in Dahl salt-sensitive rats: a role in eclampsia? Am J Physiol Heart Circ Physiol 2007;292:H1071-H1076
- 46. Cipolla MJ, Vitullo L, and McKinnon J. Cerebral artery reactivity changes during pregnancy and the postpartum period: a role in eclampsia? Am J Physiol Heart Circ Physiol 2004;286: H2127-2132
- 47. Euser AG, Cipolla MJ. Cerebral blood flow autoregulation and edema formation during pregnancy in anesthetized rats. Hypertension 2007;49:334-340
- 48. Reese TS, Karnovsky MJ. Fine structural localization of a blood-brain barrier to exogenous peroxidase. J Cell Biol 1967;34:207-217
- Digre KB, Varner MW, Osborn AG, Crawford S. Cranial magnetic resonance imaging in severe preeclampsia vs eclampsia. Arch Neurol 1993;50:399-406
- 50. Matsuda H, Sakaguchi K, Shibasaki T, Kawakami Y, Furuya K, Kikuchi Yoshihiro. Cerebral edema on MRI in severe pre-eclamptic women developing eclampsia. J Perinat Med 2005;33(3):199-205
- 51. Cipolla MJ, Vitullo L, DeLance N, Hammer E. The cerebral endothelium during pregnancy: a potential role in the development of eclampsia. Endothelium 2005;12:5-9
- 52. Wiegman MJ, Cipolla MJ. Pregnancy increases blood-brain barrier permeability coefficient (Lp) to Lucifer Yellow: role of estrgen. Reprod Sci 2008:15;S89A
- Euser AG, Bullinger L, Cipolla MJ. Magnesium sulphate treatment decreases blood-brain barrier permeability during acute hypertension in pregnant rats. Exp Physiol 2008;93:254-261
- 54. Harper, AM, Deshmukh VD, Rowan JO, Jennett WB. The influence of sympathetic nervous activity on cerebral blood flow. Arch Neurol 1972;27:1-6
- 55. Busija DW, Heistad DD, Marcus ML. Effects of sympathetic nerves on cerebral vessels during acute, moderate increases in arterial pressure in dogs and cats. Circ Res 1980;46:696-702
- 56. Bill A, Linder J. Sympathetic control of cerebral blood flow in arterial hypertension. Acta Physiol Scand 1976;96:114-121
- Moskowitz MA, Sakas DE, Wei EP, Kano M, Buzzi MG, Ogilvy C, Kontos HA. Postocclusive cerebral hyperemia is markedly attenuated by chronic trigeminal ganglionectomy. Am J Physiol 1989;26:H1736-H1739
- 58. Sándor P. Nervous control of the cerebrovascular system: doubts and facts. Neurochem Int 1999;35:237-259

- Edvinsson L, Owman C, Sjöberg NO. Autonomic nerves, mast cells, and amine receptors in human brain vessels. A histochemical and pharmacological study. Brain Res 1976;115:377-203
- Falck B, Nielsen KC, Owman C. Adrenergic innervation of the pial circulation. Scan J Clin Lab Invest Suppl 1968;102:VI:B
- 61. Arbab MA, Wiklund L, Svendgaard NA. Origin and distribution of cerebral vascular innervation from superior cervica, trigeminal and spinal ganglia investigated with retrograde and anterograde WGA-HRP tracing in the rat. Neuroscience 1986;19:695-708
- 62. Handa Y, Caner H, Hayashi M, Tamamaki N, Nojyo Y. The distribution pattern of the sympathetic nerve fibers to the cerebral arterial system in rats as revealed by anterograde labeling with WGA-HRP. Exp Brain Res. 1990;82:493-498
- 63. Raichle ME, Hartman BK, Eichling JO, Sharpe LG. Central noradrenergic regulation of cerebral blood flow and vascular permeability. Proc Natl Acad Sci USA 1975;72:3726-2730
- 64. Sadoshima S, Heistad DD. Sympathetic nerves protect eh blood-brain barrier in stroke-pone spontaeously hypertensive rats. Hypertension 1982;4:904-907
- 65. Faraci FM, Mayhan WG, Werber AH, Heistad DD. Cerebral circulation: effects of sympathetic nerves and protective mechanism during hypertension. Circ Res 1987;61:II-102-II-106
- 66. Hart MN, Heistad DD, Brody MJ. Effect of chronic hypertension and sympathetic denervation on wall:lumen ratio of cerebral blood vessels. Hypertension 1980;2:419-423
- 67. Colonna DM, Meng W, Deal DD, Busija DW. Calcitonin gene-related peptide promotes cerebrovascular dilation during cortical spreading depression in rabbits. Am J Physiol 1994;266:H1095-H1102
- 68. Bari F, Paprika D, Jancsó G, Domoki F. Capsaicin-sensitive mechanisms are involved in cortical spreading depression-associated cerebral blood flow changes in rats. Neurosci Let 2000:393:17-20
- 69. Hong KW, Pyo KM, Lee WS, Yu SS, Rhim BY. Pharmacological evidence that calcitonin generelated peptide is implicated in cerebral autoregulation. Am J Physiol 1994;266:H11-H16
- Lassen LH, Haderslev PA, Jacobsen VB, Iversen HK, Sperling B, Olesen J. CGRP may play a causative role in migraine. Cephalalgia 2001;22:54-61
- 71. Bolay H, Reuter U, Dunn AK, Huang Z, Boas DA, Moskowitz MA. Intrinsic brain activity triggers trigeminal meningeal afferents in a migraine model. Nat Med 2002;8:136-142
- 72. Juhasz G, Zsombok T, Modos EA, Olajos S, Jakab B, Nemeth J, Szolcsanyi J, Vitrai J, Badgy G. NO-induced migraine attack: strong increase in plasma calcitonin gene-related peptide (CGRP) concentration and negative correlation with platelet serotonin release. Pain 2003:106:461-470
- 73. Geppetti P, Capone JG, Trevisani M, Nicoletti P, Zagli G, Tola MR. CGRP and migraine: neurogenic inflammation revisited. J Headache Pain 2005;6:61-70
- 74. Hargreaves R. New migraine and pain research. Headache 2007;47:S26-S43
- 75. Rotton WN, Sachtleben MR, Friedman EA. Migraine and eclampsia. Obstet Gynecol 1959;14:322-330
- 76. Facchinetti F, Allais G, Nappi RE, d'Amico R, Marozio L, Bertozzi L, Ornati A, Benedetto C. Migraine is a risk factor for hypertensive disorders in pregnancy: a prospective cohort study. Cephalalgia 2008;29:286-292
- 77. Adeney KL, Williams MA. Migraine headaches and preeclampsia: an epidemiologic review. Headache 2006;46:794-803

- 78. Amara SG, Jonas V, Rosenfeld MG, Ong ES, Evans RM. Alternative RNA processing in calcitonin gene expression generates mRNA encoding different polypeptide products. Nature 1982;298:204-244
- Brain SD, Williams TJ, Tippins JR, Morris HR, MacIntyre I. Calcitonin gene-related peptide is a potent vasodilator. Nature 1985;313:54-56
- 80. Fujimori A, Saito A, Kimura S, Watanabe T, Uchiyama Y, Kawasaki H, Goto K. Neurogenic vasodilation and release of calcitonin gene-related peptide (CGRP) from perivascular nerves in the rat mesenteric artery. Biochem Biophys Res Commun 1989;165:1391-1398
- 81. Prieto D, Benedito S, Nyborg NC. Heterogenous involvement of endothelium in calcitonin gene-related peptide-induced relaxation in coronary arteries from rat. Br J Pharmacol 1991;103:1764-1768
- 82. Taché Y, Gunion M, Lauffenberger M, Goto Y. Inhibition of gastric acid secretion by intracerebral injection of calcitonin gene related peptide in rats. .Life Sci 1984;35:871-878
- 83. May A, Goadsby PJ. The trigeminovascular system in humans: pathophysiologic implications for primary headache syndromes of the neural influences on the cerebral circulation. J Cereb Blood Flow Metab 1999;19:115-127
- 84. Mulderry PK, Ghatei MA, Spokes RA et al. Differential expression of α -CGRP and β -CGRP CGRP by primary sensory neurons and enteric autonomic neurons of the rat. Neuroscience 1988;25:195-205
- 85. Poyner DR, Sexton PM, Marshall I et al. The mammalian calcitonin gene-related peptides, adrenomedullin, amylin and calcitonin receptors. Pharmacol Rev 2002:54:233-246
- 86. Hay DL. What makes a CGRP2 receptor? Clin Exp Phramacol Physiol 2007:34:963-971
- 87. Bell D, McDermott BJ. Calcitonin gene-related peptide in the cardiovascular system: characterization of receptor populations and their (patho)physiological significance. Pharmacol Rev 1996;48:253-288
- 88. Wimalawansa SJ. Calcitonin gene-related peptide and its receptors: molecular genetics, physiology, pathophysiology, and therapeutic potentials. Endocr Rev. 1996;17:533-585.
- 89. Gangula PRR, Lanlua P, Bukoski RD, Wimalawansa SJ, Yallampally C. Mesenteric arterial relaxation to calcitonin gene-related peptide is increased during pregnancy and by sex steroid hormones. Biol Reprod 2004;71:1739-1745.
- Stevenson JC, MacDonald DW, Warren RC, Booker MW, Whitehead MI. Increased concentration of circulating calcitonin gene-related peptide during normal human pregnancy. Br Med J 1986;293:1329-1320
- 91. Dong YL, Chauhan M, Green KE, Vegiraju S, Wang HQ, Hankins GD, Yallampalli C. Circulating calcitonin gene-related peptide and its placental origins in normotensive and preeclamptic pregnancies. Am J Obstet Gynecol 2006;195:1657-1667
- Gangula PRR, Wimalawansa SJ, Yallampalli C. Pregnancy and sex steroid hormones enhance circulating calcitonin gene-related peptide concentrations in rats. Hum Reprod 2000;15:949-53
- 93. Grewal M, Cuevas J, Chaudhuri G, Nathan L. Effects of calcitonin gene-related peptide on vascular resistance in rats: role of sex steroids. Am J Physiol 1999;276:H2063-H2068
- 94. Yallampalli C, Dong Y.L. Wimalawansa SJ. Calcitonin gene-related peptide reverses the hypertension and significantly decreases the fetal mortality in preeclampsia ratds induced by NG-nitro-L-arginine methyl ester. Hum Reprod 1996;11;895-899

- Gangula PRR, Supowit SC, Wimalawansa SJ, Zhao H, Hallman M, DiPette DJ, Yallampalli C.
 Calcitonin gene-related peptide is a depressor in NG-nitro-L-arginine methyl ester-induced hypertension during pregnancy. Hypertension 1997;29(2):248-253
- 96. Covarrubias DJ, Luetmer PH, Campeau NG. Posterior reversible encephalopathy syndrome: prognostic utility of quantitative diffusion-weighted MR images. AJNR 2002;23:1038-1048
- 97. McKinney AM, Short J, Truwit CL, McKinney ZJ, Kozak OS, SantaCruz KS, Teksam M. Posterior reversible encephalopathy syndrome: incidence of atypical regions of involvement and imaging findings. AJR 2007;189;904-912
- 98. Fugate JE, Claassen DO, Cloft HJ, Kallmes DF, Kozak OS, Rabinstein AA. Posterior reversible encephalopathy syndrome: associated clinical and radiologic findings. Mayo Clin Proc 2010;85:427-432
- Onder AM, Lopez R, Teomete U, Francoeur D, Bhatia R, Knowbi O, Hizaji R, Chandar J, Abitbol
 C, Zilleruelo G. Posterior reversible encephalopathy syndrome in the pediatric renal population. Pediatr Nephrol 2007;22:1921-1929
- 100. Morris EB, Laningham FH, Sandlund JT, Khan RB. Posterior reversible encephalopathy syndrome in children with cancer. Pediatr Blood Cancer 2007;48:152-159
- 101. Mirza A. Posterior reversible encephalopathy syndrome: a variant of hypertensive encephalopathy. J Clin Neurosci 2006;13:590-595
- 102. Bartynski WS, Boardman JF. Distinct imaging patterns and lesion distribution in posterior reversible encephalopathy syndrome. Am J Neuroradiol 2007;28:1320-1327
- 103. Bartynski WS. Posterior reversible encephalopathy syndrome, part 1: fundamental imaging and clinical features. Am J Neuroradiol 2008;29:1036-1042
- 104. Koch S, Rabinstein A, Falcone S, Forteza A. Diffusion-weighted imaging shows cytotoxic and vasogenic edema in eclampsia. AJNR 2001;22:1068-1070
- 105. Zeeman GG, Fleckenstein JL, Twickler DM, Cunningham FG. Am J Obstet Gynecol 2004;190:714-20
- 106. Mueller-Mang C, Mang T, Pirker A, Klein K, Prchla C, Prayer D. Posterior reversible encephalopathy syndrome: do predisposing risk factors make a difference in MRI appearance? Neuroradiology 2009;51:373-383
- 107. Podjarny E, Losonczy G, Baylis C. Animal models of preeclampsia. Semin Nephrol 2004:24:596-606
- 108. Cipolla MJ, Sweet JG, Chan SL. Cerebral vascular adaptation to pregnancy and its role in the neurological complications of eclampsia. J Appl Physiol 2010;11:[Epub ahead of print]
- 109. Kanayama N, She L, Maehara K, Kajiwara Y, Terao T. Induction of HELLP syndrome-like biochemical parameters by stimulation of the celiac ganglion in rats. J Hypertens 1996;14:453-459
- 110. Kanayama N, Khatun S, Belayet HM, She L, Terao T. Induction of eclampticlike changes by stimulation of the celiac ganglion in rats. Hypertens Pregnancy 1999;18:249-260
- 111. Smeda JS, Payne GW. Alterations in autoregulatory and myogenic function in the cerebrovasculature of Dahl salt-sensitive rats. Stroke 2003;34:1484-1490
- 112. Dobesova Z, Zicha J, and Kunes J. The influence of prenatal exposure to different salt diets on body and organ weights in newborn Dahl rats. J Dev Physiol 19: 17-21, 1993
- 113. Bayorh MA, Ganafa AA, Socci RR, Silvestrov N, and Abukhalaf IK. The role of oxidative stress in salt-induced hypertension. Am J Hypertens 17: 31-36, 2004

- 114. Kushiro T, Fujita H, Hisaki R, Asai T, Ichiyama I, Kitahara Y, Koike M, Sugiura H, Saito F, Otsuka Y, and Kanmatsuse K. Oxidative stress in the Dahl salt-sensitive hypertensive rat. Clin Exp Hypertens 27: 9-15, 2005
- 115. Cosentino F, Bonetti S, Rehorik R, Eto M, Werner-Felmayer, Volpe M, Lüscher TF. Nitric-oxide-mediated relaxations in salt-induced hypertension: effect of chronic beta1-selective receptor blockade. J Hypertens. 2002;20(3):421-8
- 116. Witlin AG. Prevention and treatment of eclamptic convulsions. Clin Obstet Gynecol. 1999;42(3):507-18.
- 117. Moran NF. Preventing and treating eclamptic seizures. Will magnesium sulphate for preeclampsia really help? BMJ. 2003 4;326(7379):50
- 118. Loureiro R, Leite CC, Kahhale S, Freire S, Sousa B, Cardoso EF, Alves EA, Borba P, Cerri GG, Zugaib M. Diffusion imaging may predict reversible brain lesions in eclampsia and severe preeclampsia: Initial experience. Am J Obstet Gynecol 2003;189:1350-1355
- 119. Bellamy L, Casas JP, Hingorani AD, Williams DJ. Pre-eclampsia and risk of cardiovascular disease and cancer in later life: systematic review and meta-analysis. BMJ 2007;335:974

Part I

Hoe als je je met zorgeloosheid kon omringen en dat dat je ruimte was

(Bert Schierbeek)