Hyperthermic isolated limb perfusion with TNF and cisplatin in the treatment of osteosarcoma of the extremities: A feasibility study in healthy dogs

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Abstract
The feasibility of hyperthermic isolated limb perfusion (HILP) with tumor necrosis factor-α (TNF) and cisplatin for the management of osteosarcoma was studied in the canine model. During seven perfusions in six healthy mongrel dogs (weight 32±2 kg) the technical aspects of HILP under mild hyperthermia (39-40°C) were studied. In five experiments HILP was performed with TNF alone (0.5 mg/L extremity volume), and in two experiments TNF was combined with cisplatin (25 mg/L extremity volume). During the perfusions physiological parameters were monitored and TNF and total cisplatin concentrations were determined. Perfusion conditions (pH, PCO₂, PO₂, flow and pressure) remained within physiological ranges. Three dogs died within 24 hours despite a sublethal systemical concentration of TNF that leaked from the perfusion circuit. Three dogs were terminated; one dog after the second experiment in accordance with Dutch ethical rules; one dog because it showed an invagination of the small bowel resulting in an ileus; one dog because of necrosis of the perfused limb. This feasibility study in healthy dogs demonstrated that HILP with TNF and cisplatin was associated with a high mortality rate and therefore does not allow us to treat dogs with spontaneous osteosarcoma with TNF and cisplatin HILP. Therefore, an alternative model should be used in the search for the ideal combination of perfusion agents for the limb sparing treatment in human osteosarcoma.

Introduction
Osteosarcoma is the most frequent primary malignant bone tumor in humans. Until the 1970s the most common approach to the management of localized osteosarcoma was surgical resection, amputation or radiation therapy.1 During the last decades a definite role for neoadjuvant high dose methotrexate and cisplatin based polychemotherapy was established.1-4 The potential local tumor effect of systemically administered cisplatin, however, is limited due to the nephrotoxicity and ototoxicity of cisplatin. Therefore an attempt was made to increase the local effect of cisplatin without increasing systemic toxicity by using hyperthermic isolated regional limb perfusion (HILP) with cisplatin in dogs with spontaneous osteosarcoma.5 These studies showed an acceptable locoregional toxicity, improved functional outcome at 6 and 12 weeks, and a steadily improving radiological picture. However, the histological results were modest, with none of the dogs showing a complete response at 6 weeks after perfusion. The same experience was found in patients with sarcomas of soft tissue and bone treated with cisplatin HILP.5 Results of recent publications and of our own experience with a new perfusion modality, which combines tumor necrosis factor-α (TNF) and melphalan in patients with recurrent melanoma or soft tissue sarcoma, are very promising.7,8 However, in 6 of 8 evaluable patients with
unresectable osteosarcoma of the lower limb treated with TNF and melphalan HILP, histological evaluation revealed moderate results with \( \geq 80\% \) necrosis in 3 patients, 50%-60% necrosis in 2 patients and < 50% necrosis in one patient. After TNF and melphalan HILP, limb sparing surgery was possible in 6 patients. As cisplatin is one of the most active chemotherapeutics in the treatment of osteosarcoma, it seems worthwhile to investigate the results of HILP with TNF and cisplatin. With the high frequency of occurrence in dogs, canine osteosarcoma is a useful model for evaluation of new treatment regimens in humans as rapid case accrual and rapid time to reach measurable end points are possible. The canine osteosarcoma therefore appears to be a valid model for studying the potential treatment of HILP with TNF and cisplatin in the local treatment of osteosarcoma of the extremity in humans. To establish optimal HILP conditions using TNF and cisplatin for local tumor control in dogs bearing osteosarcoma, a feasibility study in healthy dogs was undertaken.

**Materials and methods**

**Dogs**

During 7 experiments in 6 healthy mongrel dogs with a mean average weight of 32 ± 2 kg and a mean age of 6 ± 1 years different aspects of HILP with TNF and cisplatin were studied. Preoperatively, all dogs were thoroughly clinically evaluated at the Central Animal Facility of the University of Groningen. The study was approved by the Animal Welfare Committee of the Faculty of Medicine of the Groningen University.

**Anesthetics**

The dogs fastened for 12 hours and were anaesthetized with thiopental (30mg/kg BW, i.v.) (Pentothal, Abbott, Amstelveen, The Netherlands) and after muscle relaxation with pancuroniumbromide (0.08 mg/kg BW, i.v.) (Pavulon, Organon, Oss, The Netherlands), the dogs were ventilated (Ohmeda Modulus 2) with a mixture of O\(_2\) and isoflurane. The oxygen concentration in the gas mixture was continuously measured by means of an oxygen analyzer (Ohmeda Modulus 2) and minute volumes (4-6 L/min) were adjusted to maintain an end-expiratory CO\(_2\) concentration of 4-5% (Siemens CO\(_2\) - analyzer 930). The dogs were placed in the supine position on a heated mattress to maintain their normal body temperature of 38 °C. During the operations all dogs were given about 2 L of glucose 5% through a cephalic or internal jugular vein. Central arterial pressure was recorded as well as an ECG and diuresis.
Operation and Perfusion Techniques

During anaesthesia the volume of the extremity was measured using Archimedes rule (1.7-2 L). The iliac vessels were exposed under sterile conditions and collateral vessels were clipped. Cannulas were inserted into the artery (Bardic, 14-18 F) and vein (Bardic, 14-18 F). Both cannulas were connected to an extracorporeal circuit consisting of an occlusive roller pump, a cardiotomy reservoir and a bubble oxygenator with heat-exchanger. A nylon tourniquet was placed around the base of the extremity using, a pin in the bone and bandage around the middle to complete the isolation of the limb from the systemic circulation. The perfusate consisted of 350 ml 5% dextran 40 in glucose 5% (Isodex, Pharmacia AB, Uppsala, Sweden), 250 ml red blood cells (canine blood donors), 250 ml plasma, 30 ml sodiumbicarbonate 8.4% and 0.5 ml 5000 IU/ml heparin (Thromboliquine, Organon B.V., Oss, The Netherlands). The mixture of oxygen, air and carbondioxide through the oxygenator was adjusted to maintain the blood gas values within the physiological range and when necessary, bicarbonate was added to adjust the pH value.

All perfusions were performed under mild hyperthermic conditions (39-40°C) and optimal physiologic conditions. Thermistor probes (Electrolaboriet, Copenhagen, Denmark) were inserted into the subcutaneous tissues and into a muscle of the thigh just above the knee for continuous monitoring of the temperatures during perfusion. In the first 5 experiments TNF was the sole perfusion agent, in the last 2 experiments TNF was combined with cisplatin. The dosage of TNF (0.5 mg/L extremity volume) (Boehringer, Ingelheim, Germany) was calculated not to exceed ten times acceptable systemic levels (Systemic: 10 µg/kg bodyweight). The dosage of cisplatin (25 mg/L extremity volume) (Platinol 0.5 mg/ml, Bristol Myers Squibb, Weesp, The Netherlands) used in the perfusion had been established in a previous study and was based on a maximum tolerable dose of 30 mg/L extremity volume. Cisplatin was added to the circulated perfusate in 10 minutes. During perfusion, serum TNF and total cisplatin levels were determined in the regional and systemic circulation at 0, 5, 15, 30, 45, 60, 75 and 90 minutes by ELISA and flameless atomic absorption spectrophotometry (FAAS), respectively. The perfusion time was 1 hour, followed by washout of the extremity with 3L of Isodex. Tourniquet, cannulas and clips were then removed and the incisions in the vessels repaired. Protamine hydrochloride (Hoffman La Roche, Mijdrecht, The Netherlands) was administered, to neutralize heparin, in a ratio of 1:1 to the initial dose of heparin. All dogs were closely observed during at least 24 hours. No anti-inflammatory or analgesic drugs were administered during follow-up. All dogs were followed for local and systemic side effects of TNF and cisplatin perfusion, as well as survival.
Limb perfusion with TNF and cisplatin in healthy dogs

Results

Table 1 shows the characteristics of the 7 experiments in 6 dogs. During the experiments conditions for perfusions (pH, PCO₂, PO₂) were kept within the physiological ranges as in human perfusions. Figure 1 shows the flow, blood pressure, perfusion pressures, weight gain or loss of the extra corporeal circuit and temperature during 60 minutes of perfusion in the 7 experiments. In the first 5 experiments only TNF was administered to the perfusion circuit. In the last two experiments cisplatin was added. Figure 2 illustrates the TNF concentrations (mean ± SEM) in the perfused limb as well as in the systemic circulation of the dog during perfusion and afterwards. Peak TNF concentrations in the perfused limb were 650 ± 158 ng/ml and in the systemic circulation of the dog 37 ± 15 ng/ml. The peak systemic concentrations in the dog were in the same range as those of in TNF and melphalan HILP used in the treatment of humans at our institute.\textsuperscript{16} Figure 3 shows the measured total cisplatin values in the last two experiments. During the experiments we were not able to perform any leakage monitoring by means of radionuclear detection techniques which are used in the clinical perfusion setting. Therefore leakage was calculated afterwards according to Stehlin with the amount of blood in the dogs estimated at 69 ml/kg bodyweight.\textsuperscript{17} Calculated leakage values are summarized in Table 1.

Three dogs died within 24 hours: the first two during the TNF experiment, the third after TNF and cisplatin perfusion. Postmortem examination of these animals did not provide any macroscopic or microscopic evidence to explain the cause of death. Three dogs were terminated; two due to treatment complications. One dog showed an invagination of the small bowel resulting in an ileus and another was terminated 1

<table>
<thead>
<tr>
<th>Exp. Nr.</th>
<th>Body weight (kg)</th>
<th>Limb volume (L)</th>
<th>TNF dose (mg)</th>
<th>Cisplatin dose (mg)</th>
<th>Leakage (%)</th>
<th>Limb toxicity</th>
<th>Follow-up</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>35</td>
<td>2.7</td>
<td>1.3</td>
<td>0</td>
<td>8.7</td>
<td>II</td>
<td>Dead &lt; 24 h</td>
</tr>
<tr>
<td>2</td>
<td>31.5</td>
<td>1.4</td>
<td>0.6</td>
<td>0</td>
<td>0.3</td>
<td>II</td>
<td>Dead &lt; 24 h</td>
</tr>
<tr>
<td>3</td>
<td>26.5</td>
<td>2.3</td>
<td>1.15</td>
<td>0</td>
<td>5.1</td>
<td>II</td>
<td>Ileus, terminated &lt; 1 wk</td>
</tr>
<tr>
<td>4</td>
<td>33.5</td>
<td>2.3</td>
<td>1.15</td>
<td>0</td>
<td>4.9</td>
<td>I</td>
<td>Alive, experiment 1</td>
</tr>
<tr>
<td>5</td>
<td>31</td>
<td>1.9</td>
<td>1</td>
<td>0</td>
<td>6.1</td>
<td>n.a.</td>
<td>Terminated, experiment 2</td>
</tr>
<tr>
<td>6</td>
<td>29.5</td>
<td>2.0</td>
<td>1</td>
<td>50</td>
<td>33.0</td>
<td>V</td>
<td>Necrotic limb, terminated&lt;1wk</td>
</tr>
<tr>
<td>7</td>
<td>28.5</td>
<td>1.8</td>
<td>0.8</td>
<td>50</td>
<td>10.8</td>
<td>II</td>
<td>Dead &lt; 24 h</td>
</tr>
</tbody>
</table>

Exp. Nr. = experiment number; n.a. = not applicable, dog 4 underwent 2 experiments; limb toxicity according to Wieberdink;\textsuperscript{26} Grade I, no reaction, objectively and subjectively; Grade II, slight erythema, edema or loss of sensation; Grade III, considerable erythema or edema with some blistering, slight functional disturbances; Grade IV, extreme epidermolysis and/or obvious damage to the deep tissues causing definite functional disturbances; Grade V, reaction that might necessitate amputation.
Fig. 1  Perfusion characteristics (flow, systemic blood pressure of the dog (BP); arterial catheter pressure (P-art); venous catheter pressure (P-ven); extra corporeal circulation (ECC) weight gain (+) or loss (-) and temperature of the perfused limb (°C)) in time during 60 minutes of perfusion in 7 experiments
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Discussion

In the treatment of osteogenic sarcoma a distinction can be made between systemic therapy and locoregional treatment. High dose methotrexate based systemic chemotherapy is primarily administered in order to eradicate possible micrometastatic disease and its use was a major breakthrough in the clinical treatment of osteosarcomas in the 1970s.1,2 Today about 60% of patients with resectable primary tumors and no metastases at the time of the initial diagnosis will be cured.1 The primary objective in locoregional treatment is to prevent local recurrence and allow limb salvage procedures in an attempt to preserve limb function. New surgical techniques and the development of endoprosthetic materials, coupled with the systemic neoadjuvant chemotherapy,
have offered less radical surgery for 40-80% of patients with osteosarcoma since the 1980s.\textsuperscript{1,18} Procedures that increase tumor necrosis of the primary tumor, with reduction of viable tumor cells and tumor volume, could contribute to limb preservation strategies. Since its first use, cisplatin has been one of the most effective chemotherapeutic agents and has been incorporated in most systemic treatment regimens for osteosarcoma. A recent attempt to overcome its nephrotoxic and ototoxic limitations by administering cisplatin in HILP in the treatment of spontaneous canine osteosarcoma was histologically modest.\textsuperscript{5} Promising results of recent publications and our own experience with a new combination perfusion modality (TNF and melphalan) for recurrent melanoma or soft tissue sarcoma, but moderate histological results in patients with osteosarcoma, prompted us to investigate the combination of TNF and cisplatin in HILP for osteosarcoma.\textsuperscript{7-9} Since the endothelial cells are supposed to play a key role in the working mechanism of TNF, osteosarcomas with a high extent of tumor vessels, are of particular interest.

Before application of TNF and cisplatin HILP in humans and client owned osteosarcoma bearing dogs, the present feasibility study was performed in normal healthy dogs. Despite sufficient experience in HILP in dogs as well as in humans, an unexpected high mortality rate was encountered. Although there was no mortality related to the operation, 3 dogs died within 24 hours after perfusion (50%). This direct postoperative mortality could not be explained by a surplus of systemical leakage of TNF. In the experiment, the dog with the highest leakage and, as a consequence the highest systemical TNF concentrations, survived immediately postoperatively, and the dog with the lowest leakage (lowest systemical TNF concentrations) died within 24 hours after perfusion. No correlation between leakage and mortality rate could be established. Maximal leakage encountered in these experiments was 33%, this corresponds with 330 \textmu g TNF given systemically per dog; since the average dog weighs 33 kg, the dose of TNF that reaches the systemical circulation of the dog is sublethal (10 \textmu g/kg).\textsuperscript{14} Although only sublethal doses of TNF leaked to the systemical circulation, the clinical picture resembled responses observed in lethal doses (>100\textmu g/kg), characterized by progressive hypotension, shock and death within 24 hours.\textsuperscript{19} Due to the lack of facilities, we were not able to support the dogs with intensive postoperative care, as is the case after human TNF HILP. In part this could explain the observed direct postoperative mortality and supports the need for intensive treatment after TNF HILP in the dog.

Three dogs survived the first days after perfusion, however, one dog developed an ileus and was terminated within 1 week after perfusion. One dog that underwent two experiments survived the first without morbid effects, but was terminated after the second experiment according to the Dutch ethical rules. Leg toxicity consisted in
slight erythema and edema in all dogs except one in the cisplatin treated group. In this dog, necrosis of the perfused limb was encountered, necessitating termination. We have never observed necrosis of the perfused limb with the cisplatin dose used (25 mg/L extremity volume) in experiments were cisplatin was the sole perfusion agent. This observation may indicate that TNF might enhance the effect of cisplatin to the local tissues of the perfused limb. The in vitro anticancer potential, and overcoming cisplatin resistance with the combination of TNF and cisplatin in different cell lines, has been established by others. Buell et al. demonstrated an increased cellular cisplatin accumulation and DNA adduct formation as the possible cellular basis for the augmented cisplatin cytotoxicity in the presence of TNF and hyperthermia. Recently, Anda et al demonstrated that TNF selectively promoted the in vitro permeability of the blood-brain barrier to CDDP without disrupting the tight junctions. An improved penetration of cisplatin in the interstitial space due to a higher permeability of the vascular wall, combined with an increased cellular cisplatin accumulation and DNA adduct formation, could explain the observed necrosis of the limb in this in vivo model with the cisplatin dose used, which was previously non-toxic.

The observed mortality and morbidity that we encountered in this canine study was in similar to the experience of Withrow and colleagues (unpublished observations). The present results in normal elderly mongrel dogs indicate that treatment of dogs with spontaneous osteosarcoma using TNF and cisplatin HILP is not appropriate. Future research could focus on postoperative monitoring and care in dogs after TNF HILP; perhaps a better alternative for testing the effect of TNF with cisplatin HILP, is the use of the rat osteosarcoma model described by Manusama et al., since rats are much less susceptible to TNF than dogs.
References


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