Intraoperative fluorescence imaging in cancer
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INTRAOPERATIVE MULTISPECTRAL FLUORESCENCE IMAGING FOR THE DETECTION OF THE SENTINEL LYMPH NODE IN CERVICAL CANCER: A NOVEL CONCEPT

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ABSTRACT

PURPOSE
Real-time intraoperative near-infrared fluorescence (NIRF) imaging is a promising technique for lymphatic mapping and sentinel lymph node (SLN) detection. The purpose of this technical feasibility pilot study was to evaluate the applicability of NIRF imaging with indocyanine green (ICG) for detection of the SLN in cervical cancer.

PROCEDURES
In ten patients with early stage cervical cancer, a mixture of patent blue and ICG was injected into the cervix uteri during surgery. Real-time color and fluorescence videos and images were acquired using a custom-made multispectral fluorescence camera system.

RESULTS
Real-time fluorescence lymphatic mapping was observed in vivo in six patients; a total of nine SLNs were detected, of which one (11%) contained metastases. Ex vivo fluorescence imaging revealed remaining fluorescent signal in eleven of 197 non-sentinel LNs (5%), of which one contained metastatic tumor tissue. None of the non-fluorescent LNs contained metastases.

CONCLUSIONS
We conclude that lymphatic mapping and detection of the SLN in cervical cancer using intraoperative NIRF imaging is technically feasible. However, the technique needs to be refined for full applicability in cervical cancer in terms of sensitivity and specificity.
INTRODUCTION

Over the last decades, the sentinel lymph node (SLN) concept has proven feasible and safe in selected types of cancer such as vulvar cancer, breast cancer, early gastric cancer, and melanoma.\(^1\)\(^-\)\(^5\) Recently, intraoperative near-infrared fluorescence (NIRF) imaging was developed as a novel technique for SLN detection. By using a NIR fluorescent dye such as indocyanine green (ICG) and a sensitive fluorescence camera system, intraoperative lymph node mapping has proven feasible in breast cancer and skin cancer patients, with comparable or slightly better detection rates to conventional techniques.\(^6\)\(^-\)\(^9\) Advantages of this technique are the real-time feature for lymphatic mapping and the avoidance of radioactivity. Furthermore, the one-step procedure of NIRF imaging takes place entirely during surgery, thus improving patient comfort. To date, NIRF imaging has not been applied in cervical cancer.

The prognosis of early stage cervical cancer is substantially influenced by lymph node (LN) status, with positive LNs adding to an unfavorable prognosis.\(^10\)\(^,\)\(^11\) Standard surgical treatment therefore consists of radical hysterectomy combined with bilateral pelvic lymphadenectomy. Metastatic involvement is found in no more than 13-27% of the LNs,\(^1\)\(^,\)\(^11\) implying that radical pelvic lymphadenectomy can be regarded as overtreatment without clinical benefit in more than three quarters of the patients. Moreover, LN dissection may have the unfavorable consequence of lymphedema of the legs, occurring in 14-32% of patients.\(^12\)\(^,\)\(^13\) The SLN procedure could lead to more selective LN dissection, but its definitive value in cervical cancer is still matter of debate and subject of ongoing investigation.

The reliability in terms of sensitivity has improved markedly in the last two decades, mainly due to the combination of a radioactive tracer combined with a blue dye rather than using blue dye alone.\(^14\) This bimodal approach yields false negative rates of 0% in tumors <2 cm.\(^15\)\(^,\)\(^16\) Although these data are encouraging, SLN detection in the pelvic region remains complicated due the bilateral and not fully predictable lymphatic routing originating from the uterine cervix. About 80% of SLNs are found in the area of internal and external iliac nodes and in the obturator fossa, but involvement of lymph nodes in the parametrium and para-aortal regions has also been reported (Figure 1).\(^17\)\(^-\)\(^20\) Bilateral SLNs are found in 59-66% of patients.\(^15\)\(^,\)\(^16\) In particular, the blue dye can be difficult to trace in extrapelvic areas because of overlying muscle tissue or fat.

A great advantage of NIRF imaging over existing techniques is the high target-to-background ratio. When applied in cervical cancer, we hypothesized that real-time NIRF intraoperative imaging could be advantageous in visualization of the drainage pattern and, thus, dynamic detection of the SLN in the pelvic anatomy. We therefore initiated this pilot study to evaluate the technical feasibility and applicability of a clinical prototype intraoperative fluorescence camera system for detection of the SLN in cervical cancer.
PATIENTS AND METHODS

STUDY POPULATION
This pilot study was approved by the Institutional Review Board (IRB) of the University Medical Center Groningen and was performed in accordance with the ethical standards of the Helsinki Declaration of 1975 (Dutch Trial Register no. NTR1981; EudraCT no. 2009-010560-42). Consecutive patients with cervical cancer stage IA1, IB1 or IIA, eligible for abdominal hysterectomy and pelvic lymphadenectomy aged 21 years and older were included. They were recruited by the gynecologist and/or the research physician in the outpatient clinic. Informed consent was obtained after at least 1 week of reflection time. All patient data were depersonalized. Exclusion criteria included severe renal failure (serum creatinin ≥ 400 μmol/l), severe cardiac or pulmonary failure (ASA III-IV), past or present hyperthyroidism, pregnancy or a previous severe allergic or anaphylactic reaction to insect bites or medication.

IMAGING SYSTEM
The prototype intraoperative multispectral fluorescence camera system used in this study (Figure 2) was developed at the Helmholtz Zentrum of the Technical University Munich, Germany in close collaboration with SurgOptix (SurgOptix Inc, Redwood Shores, CA, USA). The system and its components were recently extensively described by Themelis et al. and will be pointed out briefly.

A white light source illuminates the operating field and a laser diode provides light of a desired wavelength for excitation of the fluorescent probe. Light passes through a system of optics and is separated into visible light (color), light at the emission (fluorescence) wavelength band and light at the excitation wavelength band (intrinsic). These bundles are detected by sensitive CCD-cameras. Multispectral signals from all three cameras are processed and combined in order to correct for artifacts and yield true quantitative fluorochrome biodistribution. Color and fluorescence signals can be displayed as separate images on external monitors or superimposed in one image, allowing for adequate anatomical positioning of the fluorescence signal. Images and real-time videos up to 12 frames per second are generated using customized software. During surgery, both the color image and the fluorescence signal are presented on external monitors, providing the surgeon with an unhindered view. Sterile drapes are used in the operating room to cover the camera system completely, making it applicable and safe for intraoperative use. The technical service of the University Medical Center Groningen performed necessary tests for leakage current and safety of the application of the camera in its current form during surgery.

IMAGING AGENTS
 Shortly prior to surgery, a vial of 25 mg ICG (Pulsion Medical Systems AG, Munich, Germany) was dissolved in 50 ml of water for injection (B. Braun Medical), yielding a concentration of 0.5 mg/ml. This concentration has previously been described adequate
for SLN imaging. For each patient, a new vial of ICG was used because of degradation of the fluorescence signal of ICG after dissolving. One ml of undiluted patent blue (Bleu patenté, Guerbet, France) was mixed with 1 ml of dissolved ICG in a syringe for injection into the cervix uteri and protected from light to prevent bleaching.

SURGICAL PROCEDURE
After laparotomy, the camera system was prepared for imaging and moved above the operating field. In this setting, the lens of the camera is situated approximately 25 cm above the patient. Real-time video capturing was started simultaneously with or shortly after the injection of the 2 ml mixture of ICG and patent blue in four quadrants of the cervix uteri. Propulsion of the ICG/patent blue mixture through the lymphatics was followed real-time. The first appearing lymph node (LN) was denominated as the SLN. The SLN or – in case of bilateral lymph flow - multiple SLNs were removed and the fluorescence signal was quantified ex vivo. Next, the remaining LNs in the obturator fossa and in the internal, external and communal iliac regions were dissected and evaluated for fluorescence and blue discoloration ex vivo. From this point on, surgery was carried out following the standard procedure. In case no fluorescent SLN was found in vivo, all LNs were dissected and evaluated ex vivo for a possible fluorescent signal and/or blue discoloration. All LNs were sent to the pathologist for histopathological examination. In case of a fluorescent hot spot in a cluster of LNs, the fluorescent node was excised and examined separately for tumor involvement.

RESULTS
STUDY POPULATION
Demographic and clinical characteristics are shown in Table 1. Ten patients were included in this pilot study (mean age 51.3, range 39-74; mean BMI 22.7, range 15-30). The majority of patients was diagnosed with stage IB1; five patients had a tumor <2 cm. No side effects were reported following the injection of ICG and patent blue. Histopathological examination revealed three patients with adenocarcinoma and seven with squamous carcinoma of the cervix. This is concordant with incidence rates in the population.

INTRAOPERATIVE FLUORESCENCE IMAGING
The fluorescent signal in the lymphatics became visible within 30 s upon injection of ICG and patent blue, with the SLN generally appearing within 1-2 minutes (Supplemental Video 1). Lymphatic mapping was clearly discernible in vivo in six patients (nos. 1, 3, 4, 5, 7 and 9). In one of these patients (no. 7), the lymph vessels were visible, but no SLN appeared. In contrast, no lymph vessels were visible in one patient (no. 10); nevertheless, the SLN could easily be detected in vivo. This case is illustrated in Figure 3. In total, one or more SLNs were detected in six patients (nos. 1, 3, 4, 5, 9 and 10). In these patients, a
total number of nine SLNs were detected. A bilateral SLN was detected in three patients. The SLNs were located in the left obturator fossa (three), right obturator fossa (two), left external iliac (one), right external iliac (one), right common iliac (one) and on the junction of the right internal iliac and obturator fossa (one, Figure 1). Of these nine fluorescent SLNs, six also showed a blue discoloration. Ex vivo fluorescence imaging confirmed the strong fluorescent signal in all excised SLNs. One of the SLNs turned out to contain metastatic disease on histopathological examination. A summary of the results is shown in Table 2.

In four patients, no lymphatic mapping could be observed in vivo. Three of these patients had tumors >2 cm. In patient 2, intra-abdominal fat impeded detection of fluorescence. However, upon dissection, a blue LN was discovered in the obturator fossa, also showing a strong fluorescent signal ex vivo. On histopathological examination, this LN turned out to contain metastatic tumor tissue. In patient 6, a smaller abdominal incision prevented maximal exposure of all LN basins to the camera, thus hampering detection. Leakage of ICG and patent blue into the pelvis as a result of a deep intracervical injection prevented SLN detection in patient 7. The last patient (no.8) in whom no SLN could be detected in vivo, was initially diagnosed with stage IB1 cervical cancer, but was upstaged to IB2 during surgery, a stage that does strictly not apply to the inclusion criteria. The size of the tumor could be the cause of hampered lymph flow and concomitant fluorescent and blue agents through the lymphatics.

**EX VIVO FLUORESCENCE IMAGING**

After LN dissection, all excised LNs were imaged for fluorescence ex vivo (Tables 2 and 3). A total of 197 non-sentinel LNs were taken out, with a mean number of 20 LNs per patient (range, 10-27).

In the six patients in whom SLN(s) were detected in vivo, a total of 116 non-sentinel LNs were excised. None of these contained metastases. In four of these six patients, a total of six LNs were found to show a fluorescent signal on ex vivo imaging; none of these were blue. The localization of these fluorescent LNs was in two cases contralateral from the SLN (patients 9 and 10; Table 3).

In the four patients in whom no SLN was detected in vivo, 81 LNs were taken out. Five LNs showed a fluorescent signal; three of these were also blue. An example of ex vivo fluorescence in a cluster of lymph nodes is shown in Figure 4. Bilateral fluorescent LNs were found in patient 2. Only one of these LNs, which was both fluorescent and blue, turned out to contain metastases. None of the non-fluorescent LNs contained metastases.

In summary, one of nine SLNs (11%) contained metastases. Eleven of 197 non-sentinel LNs (5.6%) showed a fluorescent signal ex vivo; three of these (1.5%) were also blue. One of these fluorescent non-sentinel LNs contained metastases.
DISCUSSION
This pilot study is, to our knowledge, the first clinical study to assess the technical feasibility of near-infrared fluorescence (NIRF) for the detection of the sentinel lymph node (SLN) in cervical cancer. We show that lymphatic mapping and SLN detection using the fluorescent contrast agent indocyanine green (ICG) is technically feasible. One or more SLNs were found in six out of ten patients; bilateral SLNs were detected in three of these patients. Overall, lymphatic mapping using fluorescence imaging was more straightforward in lean patients in whom the pelvic lymph basins could be more easily exposed; and somewhat more difficult in obese patients. Use of the intraoperative camera system in itself did not hinder the surgeon, and the surgical procedure was not extended with more than 30 minutes due to imaging. However, the limited range of movement of the camera head combined with smaller surgical incisions caused difficulties in detection of fluorescence in deep areas of the pelvis.
A number of summarized studies report detection of one or more SLNs in early stage cervical cancer in 84-97% of patients using radiocolloid in combination with blue dye.24-27 The lower detection rate of 60% in our pilot study can be explained by a number of factors, i.e., patient characteristics, the learning curve concerned with the new technique and technical features of the camera system and the fluorescent agent.
Several studies report that the SLN technique is less applicable in patients with cervical tumors >2 cm, with detection rates decreasing to 67-77%.24, 28 This could be the result of extensive tumor growth, causing either embolization or mass blocking of the lymph vessels.
Consistent with literature, we observed better detection rates in tumors smaller than 2 cm. SLNs were found in four out of five patients with tumors <2 cm (80%) and in only two out of five patients with tumors >2 cm (40%). Bilateral SLNs were detected in two tumors <2 cm (40%) and in one tumor >2 cm (20%). Furthermore, in one patient who presented with a large tumor (stage IB2; patient 8), neither the fluorescent nor the blue agent could be located in vivo in the pelvis of ex vivo in dissected LNs, indicating that the agents were not carried from the tumor into the lymphatics towards the SLN or LNs. Although these data suggest support for existing literature regarding the impact of tumor size on (bilateral) SLN detection, our numbers are too small to draw definitive conclusions from.
The detection rate of 60% observed in this pilot study may furthermore be influenced by the learning curve of working with the intraoperative camera system and correct injection of the fluorescent agent. As gynecologic oncologists got more acquainted with the technique and the injection, the procedure became more standardized. For example, in patient 2, no SLN was detected in vivo; however, when exposing the obturator lymph basin better, a blue LN was found, which turned out to be fluorescent as well. In retrospect, we believe this LN would have been detected as the SLN had we better exposed the lymph basin. In two of three patients with a unilateral SLN, we observed fluorescence ex vivo in a contralaterally located LN. These findings indicate that in these patients,
although the fluorescent agent was transported bilaterally from the uterine cervix; not all SLNs were detected. More experience with the technique may lead to a higher detection rate of SLNs in vivo.

Apart from patient selection and learning curve, we encountered several technical aspects influencing our results. Deep-lying LNs were in some cases difficult to visualize due to the limited range of movement of the camera head. A smaller, more flexible, preferably handheld camera system or even a laparoscopic system for intraoperative fluorescence imaging could overcome this problem. Technical progress will undoubtedly lead to more ergonomic and clinically-suited camera systems.

The most important limitation, however, appeared to be the penetration depth of ICG which lies around 1 cm. This is sufficient for superficial LNs, but generally not enough to visualize lymph nodes covered by adipose tissue or located deep in the pelvis hidden by the iliac vessels. In the current, still limited range of FDA-approved fluorescent agents, none has a larger penetration depth than ICG. In order to improve fluorescence imaging in the pelvis, future developments should be aimed at designing fluorescent agents with stronger tissue penetrating properties.

Apart from these pitfalls, fluorescence imaging also bears several advantages. The most important benefit for patients is that the fluorescent agent is injected in a one-step sequence during surgery, when the patient is already anesthetized, thus preventing four injections in the cervix several hours or even a day prior to surgery as is necessary when using a radiocolloid. Furthermore, we observed that the fluorescence signal is well discernable in the pelvic anatomy as long as the desired region of interest is projected within the field of view of the camera system. As mentioned earlier, more flexible camera systems with a greater range of movement will most certainly yield more optimized detection in deeper seated areas. Although the penetration depth of ICG will not exceed 1 cm, fluorescence is easy to locate in the pelvic region when compared to detection of the blue dye by visual inspection. Additionally, the real-time video properties aid the gynecologic oncologist in dynamic tracing of lymph flow towards the SLN within minutes after injection into the cervix. This unique feature of NIRF imaging provides a real-time image of the location and flow of the fluorescent agent in the lymphatic system, in close relation to the adjacent anatomical structures. This would, to our opinion, be the major advantage of NIRF imaging over conventional SLN detection methods. Future NIR fluorescent agents with better penetration properties are needed to improve detection rates.
CONCLUSION
We show that near-infrared fluorescence (NIRF) imaging using indocyanine green (ICG) for the detection of the sentinel lymph node (SLN) in early stage cervical cancer is technically feasible. Advantages of this approach are the real-time tracing of lymph flow and the one-step procedure. However, technical improvements and fluorescent dyes with larger penetration strength are needed to refine the technique.

ACKNOWLEDGEMENTS
The authors wish to thank Dr. M. van Oosten and Dr. K.T. Buddingh for assistance in data collection.
### Table 1 – Demographics and results

<table>
<thead>
<tr>
<th>Study nr.</th>
<th>Age</th>
<th>BMI</th>
<th>FIGO Stage</th>
<th>Tumor type(^a)</th>
<th>Tumor size</th>
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</thead>
<tbody>
<tr>
<td>1</td>
<td>41</td>
<td>19</td>
<td>IB1</td>
<td>Squamous</td>
<td>No tumor left after LEEP</td>
</tr>
<tr>
<td>2</td>
<td>70</td>
<td>26</td>
<td>IB1/IIA</td>
<td>Squamous</td>
<td>&gt; 2 cm</td>
</tr>
<tr>
<td>3</td>
<td>48</td>
<td>24</td>
<td>IB1</td>
<td>Squamous</td>
<td>&gt; 2 cm</td>
</tr>
<tr>
<td>4</td>
<td>40</td>
<td>24</td>
<td>IA2</td>
<td>Adeno</td>
<td>&lt; 2 cm</td>
</tr>
<tr>
<td>5</td>
<td>40</td>
<td>22</td>
<td>IB1</td>
<td>Adeno</td>
<td>&gt; 2 cm</td>
</tr>
<tr>
<td>6</td>
<td>40</td>
<td>20</td>
<td>IB1</td>
<td>Adeno</td>
<td>No tumor left after LEEP</td>
</tr>
<tr>
<td>7</td>
<td>70</td>
<td>23</td>
<td>IB1</td>
<td>Squamous</td>
<td>&gt; 2 cm</td>
</tr>
<tr>
<td>8</td>
<td>74</td>
<td>24</td>
<td>IB2</td>
<td>Squamous</td>
<td>&gt; 2 cm</td>
</tr>
<tr>
<td>9</td>
<td>39</td>
<td>15</td>
<td>IB1</td>
<td>Squamous</td>
<td>&lt; 2 cm</td>
</tr>
<tr>
<td>10</td>
<td>51</td>
<td>30</td>
<td>IB1</td>
<td>Squamous</td>
<td>&lt; 2 cm</td>
</tr>
</tbody>
</table>

\(^a\) Histologic subtype; squamous cell carcinoma (‘squamous’) or adenocarcinoma (‘adeno’)


LEEP – Loop Electrosurgical Excision Procedure conization

### Table 2 – Summary of in vivo and ex vivo results

<table>
<thead>
<tr>
<th>SLN detected in vivo</th>
<th>SLNs fluorescent (^a)</th>
<th>SLNs blue (^b)</th>
<th>SLN metastasis (^c)</th>
<th>LN dissected</th>
<th>LNs fluorescent (^d)</th>
<th>LNs blue (^e)</th>
<th>LN metastasis (^f)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Yes (n=6)</td>
<td>9</td>
<td>6</td>
<td>1 (11%)</td>
<td>116</td>
<td>6</td>
<td>0</td>
<td>0</td>
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<tr>
<td>No (n=4)</td>
<td>n/a</td>
<td>n/a</td>
<td>81</td>
<td>5</td>
<td>3</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>Total (n=10)</td>
<td>9</td>
<td>5</td>
<td>1 (0.5%)</td>
<td>197</td>
<td>11</td>
<td>3</td>
<td>1</td>
</tr>
</tbody>
</table>

\(^a\) Number of sentinel lymph nodes (SLNs) showing fluorescence in vivo.

\(^b\) Number of SLNs showing a blue discoloration.

\(^c\) Number of SLNs that were confirmed to contain metastatic tumor tissue.

\(^d\) Number of non-sentinel lymph nodes (LN) showing fluorescence ex vivo.

\(^e\) Number of LNs showing a blue discoloration ex vivo.

\(^f\) Number of LNs that were confirmed to contain metastatic tumor tissue.
Table 3 - Localization of fluorescent sentinel lymph nodes \textit{in vivo} and fluorescent non-sentinel lymph nodes \textit{ex vivo}

<table>
<thead>
<tr>
<th>Study nr.</th>
<th>No. SLN(s)</th>
<th>Localization SLN(s) a</th>
<th>SLNs with metastasis</th>
<th>No. fluorescent LN(s) \textit{ex vivo}</th>
<th>Localization fluorescent LN(s) b</th>
<th>LNs with metastasis c</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>2</td>
<td>Obturator left Obturator right</td>
<td>0</td>
<td>0</td>
<td>-</td>
<td>0</td>
</tr>
<tr>
<td>2</td>
<td>0</td>
<td>-</td>
<td>-</td>
<td>1</td>
<td>Obturator right</td>
<td>1</td>
</tr>
<tr>
<td>3</td>
<td>2</td>
<td>External iliac left External iliac right</td>
<td>0</td>
<td>1</td>
<td>Junction external / common iliac right</td>
<td>0</td>
</tr>
<tr>
<td>4</td>
<td>2</td>
<td>Obturator left Obturator right</td>
<td>0</td>
<td>2</td>
<td>External iliac right Obturator right</td>
<td>0</td>
</tr>
<tr>
<td>5</td>
<td>1</td>
<td>Obturator left</td>
<td>1</td>
<td>0</td>
<td>-</td>
<td>0</td>
</tr>
<tr>
<td>6</td>
<td>0</td>
<td>-</td>
<td>-</td>
<td>1</td>
<td>External iliac left</td>
<td>0</td>
</tr>
<tr>
<td>7</td>
<td>0</td>
<td>-</td>
<td>-</td>
<td>3</td>
<td>External iliac left (2) External iliac right</td>
<td>0</td>
</tr>
<tr>
<td>8</td>
<td>0</td>
<td>-</td>
<td>-</td>
<td>0</td>
<td>-</td>
<td>0</td>
</tr>
<tr>
<td>9</td>
<td>1</td>
<td>External iliac right</td>
<td>0</td>
<td>2</td>
<td>Obturator left Common iliac right</td>
<td>0</td>
</tr>
<tr>
<td>10</td>
<td>1</td>
<td>Common iliac right</td>
<td>0</td>
<td>1</td>
<td>External iliac left</td>
<td>0</td>
</tr>
</tbody>
</table>

a. Localization \textit{in vivo} where the sentinel lymph node(s) (SLN) was found
b. Initial localization of lymph nodes (LN) that showed fluorescence \textit{ex vivo} after dissection.
c. Number of LNs showing fluorescence \textit{ex vivo} that were confirmed to contain metastatic tumor tissue.
REFERENCE LIST

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