The beta-adrenergic system and airway reactivity. Investigations in vivo and in vitro
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CHAPTER VI

SUMMARY AND CONCLUSIONS

This thesis deals with some aspects of airway hyperreactivity. This phenomenon is clinically defined as a condition of increased responsiveness of the bronchi i.e., the occurrence of bronchial obstruction after a stimulus in a dose which does not provoke such a reaction in most individuals. The mechanism underlying airway hyperreactivity has only partially been solved. In this thesis we started from the hypothesis that the main cause of airway hyperreactivity is a disturbed balance between bronchoconstricting and bronchodilating forces. On the one hand, this imbalance may be caused by an increase in bronchoconstricting forces due to enhanced vagal activity and/or mediator release by mast cells. On the other hand, an imbalance could arise from a disturbance in the beta-adrenergic system, thus leading to a decreased bronchodilating activity. This imbalance theory is extensively reviewed in Chapter I, with special reference to the beta-adrenergic system.

The aims of the presented investigations were:

1. to evaluate the role of a disturbed beta-adrenergic system in well-defined groups of asthmatic patients with airway hyperreactivity;
2. to investigate the effects of several exogenous factors (allergen challenge, bronchodilator treatment) on the beta-adrenergic system and/or airway hyperreactivity, in order to obtain more insight into the pathogenic process of asthma and the control of this process by drugs.

Methods

In a number of studies, the beta-adrenergic system in the asthmatic patient was investigated by parallel measurements in vivo and in vitro (see Chapters III and IV). In vitro, we assessed beta-adrenergic receptor numbers and beta-adrenergic receptor function of isolated peripheral lymphocytes, as an approach to investigate the molecular "status" of the beta-adrenergic receptor system under various clinical conditions. In this approach it is assumed that a change in the beta-adrenergic receptor function of lymphocytes may reflect an altered beta-adrenergic function of the (less accessible) airways (Parker and Barnett, 1982). In vivo, we determined static and dynamic airway hyperreactivity by gas dilution studies, in order to assess the clinical relevance of a disturbed beta-adrenergic system to oppose bronchoconstriction.

In Chapter II.1, we described the methodological approach. In Chapter II.2, a new method of preparing broken lymphocyte preparations for beta-adrenergic receptor assay, using chaotropic denaturing agents, was introduced. This method is time-saving and easy to perform, in contrast to previously reported methods (see Chapter II.2).
The reactivity. This phenomenon is the nonsensiveness of the bronchi i.e., a disturbed balance between bronchial responsiveness and beta-adrenergic system in well-defined groups of patients (allergic challenge, bronchial asthma, and/or airway hyperreactivity, process of asthma and the consequences reviewed in Chapter I).

In the asthmatic patient was studied (see Chapters III and IV), and beta-adrenergic receptor approach to investigate the molecular mechanisms underlying various clinical conditions, the beta-adrenergic receptor function of the accessible) airways (Parker and Smith, 1973; Szentivanyi, 1979, 1980; Nahorski and Barnett, 1982). In vivo, we determined the sensitivity of the airways to propranolol, as a parameter of airway hyperreactivity which may indicate the capacity of the beta-adrenergic system to oppose bronchial obstructive forces. This parameter was used to assess the clinical relevance of the in vitro studies.

Chapter II described three methods which were developed to accomplish this approach.

In Chapter II.1 a new method for the extraction and the determination of cAMP from human peripheral lymphocytes is presented. Usually, cAMP is extracted from cells using the chaotropic denaturing agent trichloroacetic acid (TCA). Since TCA interferes in the competitive protein binding assay for cAMP, this agent needs to be removed first by multiple extractions with diethyl ether, which procedure is laborious and time consuming. In the new method the cells are extracted with diluted HCl in a boiling water bath. After centrifugation of precipitated protein, the samples are neutralized by the addition of excess CaCO₃, which dissolves rapidly until a neutral pH is reached, thus avoiding an exact titration of the samples with alkali. The excess of undissolved CaCO₃ is removed by centrifugation and cAMP can be determined in the supernatant. This method has proven to be reliable, rapid and convenient, especially when many samples have to be determined.

In Chapter II.2 the binding of (−) ³H-dihydroalprenolol (³H-DHA) to intact and broken lymphocyte preparations was characterized, in order to obtain a reliable beta-adrenergic receptor assay. Although a beta-adrenergic receptor determination in membrane preparations was described already in 1976 (Williams et al., 1976), considerable differences in binding characteristics have since been reported. It was demonstrated that this may be largely due to differences in the definition of non-specific binding. In the literature, non-specific binding has been defined by using a wide range of propranolol concentrations (1-50 μM) and millimolar concentrations of isoprenaline. By inhibition studies it was shown that concentrations of propranolol exceeding 1 μM may inhibit low affinity, non-receptor binding of ³H-DHA, both in membrane preparations and in intact cells, thus leading to aberrant characteristics and overestimation of the supposed 'specific' binding. By contrast, isoprenaline appeared to compete for specific binding only. Recent studies indicated that non-specific binding of ³H-DHA, in part, reflects partitioning of this ligand into membranes. This partitioned ligand is relatively unaffected by hydrophilic agents such as isoprenaline, whereas more lipophilic agents such as propranolol may displace this binding (Mendel
Intact lymphocyte measurements were additionally complicated by a very high degree of non-specific binding due to intracellular accumulation of the ligand. As in other cell types, this part of the non-specific binding could effectively be inhibited by inclusion of 0.1 mM phentolamine in the incubation medium, thus providing a valuable method to determine beta-adrenergic binding sites on intact viable lymphocytes. The mechanism of phentolamine action has not yet been fully established, but it presumably inhibits uptake of the radioligand into the lysosomes (Dulis and Wilson, 1981).

Under optimal conditions, $^3$H-DHA binding to both intact cells and membranes appeared to be rapid and reversible, saturable, of high affinity, stereoselective and of beta$_2$-adrenergic specificity. In both preparations, a similar beta-adrenergic receptor density was found, indicating that identical beta-adrenergic receptor sites were being labelled. Nevertheless, some differences were found in the dissociation constants of $^3$H-DHA and isoprenaline. Membrane receptors showed a reduced affinity for $^3$H-DHA and an increased affinity for isoprenaline when compared to intact cells. The cause and the physiological meaning of the reduced affinity for the radiolabelled antagonist are still unclear. However, the higher affinity for isoprenaline may at least partially be explained by depletion of GTP during cellular disruption and fractionation. Thus, it may be concluded that with respect to binding affinities care must be taken when extrapolating binding data obtained with membranes to intact cell physiology. This is particularly true for the physiologically meaningful agonist binding, which is often used to monitor the degree of coupling between beta-adrenergic receptor and adenylate cyclase in a variety of (patho-)physiological conditions (Lefkowitz and Michel, 1983).

In Chapter II.3 we presented the measurements of the degree of airway hyperreactivity in a group of 39 asthmatic patients by inhalation-provocation tests with histamine, acetylcholine, and propranolol, respectively. In this study we attempted to investigate the involvement of the beta-adrenergic system in the regulation of the bronchial smooth muscle tone. An increased bronchial responsiveness to inhaled propranolol, a non-selective beta-blocker, was taken as an indication that the beta-adrenergic system was involved. The results suggested that propranolol (in the concentrations used) is less sensitive in determining the presence of airway hyperreactivity than histamine or acetylcholine. The airway reactivity to propranolol was predominantly found in patients with a relatively high degree of hyperreactivity to histamine and acetylcholine dependent on beta-adrenergic.

The beta-adrenergic system is demonstrable in well-defined allergic and non-allergic asthmatic patients. In the latter, patients with severe asthma show increased blood eosinophil counts, while the former show increased blood eosinophil counts, while the latter patients show increased blood eosinophil counts and a high degree of airway hyperreactivity. This relationship might be explained by the presence of a high degree of airway hyperreactivity in allergic asthma and a high degree of airway hyperreactivity in non-allergic asthma.

In a first study (Chapter III) we investigated the effect of allergen-induced airway hyperreactivity in atopic asthmatic patients. Since it is known that the airway hyperreactivity in atopic asthmatic patients is dependent on beta-adrenergic dysfunction, we attempted to determine the degree of airway hyperreactivity in atopic asthmatic patients before and after allergen challenge. Twenty-four hours after allergen challenge, the airway hyperreactivity in atopic asthmatic patients was significantly reduced compared to the pre-challenge measurement. These findings support the hypothesis that allergen-induced airway hyperreactivity is dependent on beta-adrenergic dysfunction.
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The patients who showed a bronchoconstriction with propranolol had, in general, a more severe asthma as indicated by a lower FEV₁ as a percentage of predicted FEV₁, increased blood eosinophils, more positive skin tests, and lower histamine and ace-

tylcholine thresholds than those who did not respond. Furthermore, in the proprano-

lol responders group more positive allergen challenges were observed. This result is

hat one would expect in asthmatics who have positive skin tests to common allergens in combination with a high degree of hyperreactivity. The results suggest a relation-

ship between propranolol sensitivity and the occurrence of allergic phenomena. How-

ever, this relationship might be simply due to the fact that patients with allergic

ptoms usually have the highest degree of airway hyperreactivity.

The beta-adrenergic system in allergic asthmatic patients and its modulation by al-

gen-induced asthmatic reactions

Chapter III describes some studies on the functioning of the beta-adrenergic

system in well-defined allergic asthmatic patients who were not treated with beta-

nergic agonist drugs. We assessed the beta-adrenergic receptor function in lymphocytes of these patients in relation to parameters of airway hyperreactivity. In this respect, special attention was focussed on the airway reactivity to the beta-

blocker propranolol.

In a first study (Chapter III.1), beta-adrenergic responsiveness of lymphocytes and propranolol thresholds were determined before and after inhalation challenge with house dust mite allergen. These studies were performed to investigate the influence of an allergen-induced asthmatic reaction on the beta-adrenergic receptor system, since it is known that these reactions may modulate airway hyperreactivity (Göke-

meyer, 1976; Cockcroft et al., 1977). It was demonstrated that lymphocytes of rela-
tively stable asthmatic patients with increased airway reactivity to histamine had a normal cAMP response to increasing doses of isoproterenol. This indicates that air-

way hyperreactivity in asthma is not primarily caused by an intrinsic, generalized beta-adrenergic dysfunction, as initially proposed by Szentivanyi (1968, 1980). Twenty-four hours after an allergen-induced asthmatic reaction, however, the beta-

nergic response in these patients was significantly reduced by about 50%, whereas that of normal controls remained unchanged 24 hours after inhalation of allergen.

These findings support previous observations that a reduced beta-adrenergic receptor
function in lymphocytes of asthmatic patients is most pronounced during active and severe symptoms (Parker and Smith, 1973; Brooks et al., 1979; Kariman, 1980), and they suggest that the reduced beta-adrenergic responsiveness is a consequence of an active disease state rather than its cause. In addition, we observed that the reduced beta-adrenergic responsiveness of the lymphocytes was accompanied by an enhanced airway reactivity to propranolol in these patients. This might indicate that the reduced lymphocyte adrenergic receptor function indeed reflects a reduced beta-adrenergic receptor function of the airways. More direct evidence for this assumption was presented by Szentivanyi (1979), who found parallel changes in beta-adrenergic binding sites of lymphocytes and lung tissue. However, from our study there appeared to be no significant correlation between the extent of the changes in lymphocyte cAMP response and in propranolol sensitivity. This lack of correlation might be due to the low sensitivity of the propranolol threshold determination and the low number of experiments. Moreover, other factors such as increased vagal activity, possibly induced by the inflammatory process during the late asthmatic response, might contribute to the enhanced airway reactivity.

The purpose of a second study (Chapter III.2) was to localize the alteration in the beta-receptor-adenylate cyclase system of the lymphocytes. In this investigation, \(^3\)H-DHA binding characteristics and adenylate cyclase responses to isoproterenol, GppNHp, and NaF were determined in lymphocyte membranes of asthmatic patients and healthy control subjects, again before and after house dust mite challenge. In accordance with the above-mentioned intact lymphocyte experiments, the lymphocyte membranes of the patients showed a normal beta-adrenergic receptor number, a normal dissociation constant of \(^3\)H-DHA and normal adenylate cyclase responses to isoproterenol, GppNHp and NaF, when determined in the non-acute phase. After house dust mite challenge, however, all parameters, except the dissociation constant of \(^3\)H-DHA, were significantly reduced in the patients but not in the controls. The adenylate cyclase response to isoproterenol was reduced by about 40%, a decrease similar to that found in the intact cells. The beta-adrenergic receptor number showed a small but significant change from 1048 ± 77 to 829 ± 57 receptors/cell (20% decrease), indicating that a reduced receptor density may have contributed to the reduced beta-adrenergic responsiveness. However, the change in agonist-induced adenylate cyclase activity was better correlated with changes in GppNHp- and NaF-induced adenylate cyclase responses (about 40%), indicating that alterations distal to the receptor (regulatory protein or catalytic unit) may play a dominant role in reduced lymphocyte beta-adrenergic responsiveness as four.

Uncoupled receptors, with the antagonist \(^3\)H-DHA, changes in receptor number and be stressed that \(^3\)H-DHA antagonists will be blunted. Thus in this study it was demonstrated in lymphocyte membranes of determined after allergen provocation.

Thus, altogether the regional dysfunction of the lymphocytes and catecholamines may be emphasized. Keijzer et al., unpublished data, agonists could be involved in homologous or heterologous mechanisms. They tested this hypothesis by inclusion of isoprenaline and histamine. It was found that isoprenaline caused a decrease in lymphocyte cAMP responses to isoprenaline, histamine and that both agonists could contribute to the spasm of isoprenaline, histamine and no effect on the beta-adrenergic receptors. It was concluded that both catecholamine and histamine levels of lymphocytes in lymphocytes to histamine could be cause-