In vivo labeling of rabbit cholinergic basal forebrain neurons with fluorochromated antibodies

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Cholinergic basal forebrain neurons (CBFN) expressing the low-affinity neurotrophin receptor p75 (p75NTR) were previously selectively labeled in vivo with carbocyanine 3 (Cy3)-tagged anti-p75NTR, but the applied 192IgG-conjugates recognized p75NTR only in rat. The antibody ME 20.4 raised against human p75NTR had been shown to cross-react with the receptor in monkey, raccoon, sheep, cat, dog, pig and rabbit. Hence, for in vivo labeling of rabbit CBFN in the present study, ME 20.4 was fluorochromated with Cy3-N-hydroxysuccinimide ester and purified Cy3-ME 20.4 was injected intracerebroventricularly. Two days post-injection, clusters of Cy3-ME 20.4 were found in CBFN displaying choline acetyltransferase-immunoreactivity. Following photoconversion, electron microscopy revealed fluorochromated antibodies in secondary lysosomes. In conclusion, Cy3-ME 20.4 might become an appropriate marker for CBFN in live and fixed tissues of various mammalian species.

Key words: Choline acetyltransferase; Cholinergic system; Low-affinity neurotrophin receptor; Oxygen-enriched photoconversion

INTRODUCTION

Severe damage to cholinergic neurons of the basal forebrain (BFB) is a major neuropathological hallmark of Alzheimer’s disease, and this has prompted investigations focused on this brain region in various mammalian species [1]. Neuroanatomical studies have achieved the identification of cholinergic nerve cells by the detection of their marker enzyme choline acetyltransferase (ChAT, EC 3.2.1.6) as well as of vesicular acetylcholine transporters. An additional selective marker of cholinergic BFB neurons is the low-affinity neurotrophin receptor p75 (p75NTR), as was demonstrated in the rat [2], raccoon [3], monkey [4] and human [5]. Targeting of p75NTR led to the development of cholinergic immunolesion models. In particular, the in vivo application of antibodies directed against an extracellular epitope of rat p75NTR, as demonstrated in the rat [2], raccoon [3], monkey [4] and human [5]. Targeting of p75NTR led to the development of cholinergic immunolesion models. In particular, the in vivo application of antibodies directed against an extracellular epitope of rat p75NTR, as demonstrated in the rat [2], raccoon [3], monkey [4] and human [5].

A further derivative of 192IgG, namely its conjugate with the red fluorochrome carbocyanine 3 (Cy3-192IgG), was introduced as a tool for the selective labeling of rat cholinergic, p75NTR-bearing neurons both in vivo [7,8] and in vitro [9]. 192IgG-conjugates are commonly infused intracerebroventricularly and bind to p75NTR on cholinergic terminals, become internalized as ligand/receptor complexes and are transported retrogradely to the perikarya of cholinergic BFB neurons [6]. Internalized Cy3-192IgG molecules form clusters in secondary lysosomes [8]. Whereas ribosome inactivation by saporin causes neuronal death [6,10], Wu et al. [11] demonstrated that pre-labeling of cholinergic neurons with Cy3-192IgG does not alter their electrophysiological and pharmacological properties in ex vivo brain slice preparations containing the medial septum/diagonal band complex (MSDB). Concordantly, identification of cholinergic nerve cells with Cy3-192IgG in brain slices led to new insights in the synaptic organization of neurons in the MSDB with implications for learning and memory [11,12].

However, 192IgG conjugates recognize p75NTR only in the rat precluding their application in other mammalian species (e.g. monkeys) with a BFB chemoarchitecture resembling that in humans [13]. In contrast, the monoclonal antibody ME 20.4 raised against an extracellular epitope of the human p75NTR [14] was demonstrated to cross-react with p75NTR also in monkey, raccoon [3], rabbit, pig and cat [15]. Accordingly, the ME 20.4-saporin conjugate was shown to be a selective cholinotoxin in the common marmoset [16], rhesus monkey [17], rabbit [18] and sheep [19].

To overcome the restriction of in vivo labeling in the rat, the present study investigated whether Cy3-ME 20.4 can be employed as a marker of living cholinergic neurons in a wide range of mammalian species expressing analogues of the human p75NTR. Encouraged by recent data on ME 20.4-saporin-induced immunolesions of cholinergic cells and...
subsequent β-amyloid deposition in the rabbit brain [18], the potential of Cy3-ME 20.4 as a label for cholinergic nerve cells was exemplified in studies on BFB neurons of this animal species.

MATERIALS AND METHODS

Preparation of Cy3-ME 20.4: Cy3-ME 20.4 was prepared by coupling the antibody with a chemically activated ester of Cy3 under reaction conditions proved to maintain the antigen-binding properties of ME 20.4 [20]. Four hundred micrograms of ME 20.4 (pure IgG; AB-N07, Advanced Targeting Systems, San Diego, CA, USA) were reacted with a monofunctional Cy3-N-hydroxysuccinimide ester (Antibody Cy3 Labeling Kit, Amersham Pharmacia Biotech, Freiburg, Germany) according to the instruction of the manufacturer at pH 9.3 for 1 h, but with doubled dye:protein ratio. Thereafter, the reaction mixture was dialyzed against phosphate-buffered saline (3 x 4 h) and the concentration of Cy3-ME 20.4 was adjusted to approximately 0.5 mg/ml prior its use for intracerebroventricular (i.c.v.) injections.

Surgical procedure: New Zealand white rabbits of either sex (n = 5; 2.3–5.1 kg; 4–21 months of age) were used in this study. The rabbits were anesthetized with a mixture of ketamine (50 mg/kg; Exalgan, Merck, Hallbergmoos, Germany) and xylazine (4 mg/kg; Rompun, Bayer AG, Leverkusen, Germany), and their heads were mounted in a stereotaxic frame. Two animals received unilateral i.c.v. injections, and three animals were injected bilaterally with 16 μl Cy3-ME 20.4 at an infusion speed of 0.5 μl/min with a Hamilton microsyringe (Hamilton, Bonaduz, Switzerland). Injection co-ordinates were applied according to Beach et al. [18] at AP = 0.0 mm, L = 2.2 mm and D = 9.0/7.5 mm relative to bregma. All efforts were made to minimize animal suffering throughout the experiments including administration of the analgetic Metamizol (150 mg/kg; Berlozin, Berlin-Chemie, Germany) immediately after operation. The antibiotic Enrofloxacin (10 mg/kg daily; Baytril, Bayer AG) was given orally throughout the post-injection period. The care and treatment of the animals were in accordance with the European Communities Council Directive (86/609/EEC) and approved by the Laboratory Animal Care and Use Committee of the University of Leipzig (TVV-Nr.15/00).

Immunochemistry and electron microscopy: Two days post-injection rabbits were transcardially perfused with 4% paraformaldehyde containing either 0.5% glutaraldehyde (three animals) or 0.1% glutaraldehyde (two animals) in 0.1 M phosphate-buffer (pH 7.4). Whole brains were removed from the skulls, divided into fore- and hindbrain regions and post-fixed overnight in 4% paraformaldehyde. Two forebrains were coronally sectioned on a vibratome at 50 μm and the remaining tissue was cryoprotected and cut at 30 μm on a cryostat microtome. Sections spanning the MSDB and the anterior subdivision of the nucleus basalis magnocellularis were washed with Tris-buffered saline (TBS, 0.1 M, pH 7.4), rinsed in distilled water, mounted, air-dried and coverslipped with Entellan (in toluene; Merck, Darmstadt, Germany).

To determine the specificity of in vivo labeling, Cy3-ME 20.4 pre-labeled sections were applied to the concomitant immunolabeling of ChAT and the calcium-binding protein calretinin. Non-specific binding sites for the immunoreagents were primarily blocked with 5% normal donkey serum in 0.1 M Tris-buffered saline (TBS) containing 0.3% Triton X-100 for 1 h. Subsequently, the sections were incubated with a mixture of affinity-purified goat anti-ChAT (1:25; AB144P, Chemicon International; Hofheim, Germany) and rabbit anti-calretinin (1:300; AB149, Chemicon) diluted in the blocking solution for 16 h. Next, rinsed sections were reacted with a cocktail of Cy2-tagged donkey anti-goat IgG and Cy5-conjugated donkey anti-rabbit IgG (20 μg/ml; Dianova, Hamburg, Germany) diluted in TBS containing 2% bovine serum albumin for 1 h. Omission of the primary antibodies in control experiments led to the absence of any cellular Cy2- and Cy5-labeling. After the cytochemical procedures the sections were rinsed, mounted, air-dried and coverslipped in Entellan. Fluorescence microscopy was performed using an Axiosplan microscope (Zeiss, Oberkochen, Germany) and selected sections were inspected with a Zeiss 510 confocal laser-scanning microscope.

To quench the autofluorescence caused by lipophilic lipofuscin-like compounds in sections from 21-month-old rabbits, tissue was treated with Sudan Black B according to Schnell et al. [21] but prior to the immunofluorescence labeling of cholinergic neurons. All Sudan Black B-pre-treated sections were embedded in glycerol/gelatin (Sigma, Deisenhofen, Germany).

The subcellular distribution of Cy3-ME 20.4 was determined following photoconversion of Cy3-ME 20.4 into an electron-dense reaction product applying oxygen-enriched photoconversion in a closed conversion chamber as described previously [8]. Specimens were analyzed with a Zeiss 900 electron microscope.

RESULTS

Two days following the i.c.v. infusion of Cy3-ME 20.4 there was strong punctate labeling in many cells of the MSDB (Fig. 1) that became less intense in the nucleus basalis magnocellularis. This staining pattern was observed irrespective of the concentration of glutaraldehyde in the fixative. In comparison to rabbits unilaterally infused with Cy3-ME 20.4, bilaterally injected animals appeared to contain more in vivo-labeled nerve cells in both hemispheres.

In vivo labeling of Cy3-ME 20.4 was exclusively found in cholinergic neurons, as revealed by their immunoreactivity for ChAT (Fig. 1a) and p75NTR (not shown). In contrast, in vivo labeling was never observed in non-cholinergic, calretinin-immunoreactive nerve cells (Fig. 1b) that are known to be intermingled with cholinergic neurons in the border zone of the medial and lateral septum [22] and in magnocellular, parvalbumin-containing GABAergic BFB neurons (not shown).

It is noteworthy that treatment of the sections from 21-month-old rabbits with Sudan Black B quenched the autofluorescence caused by lipophilic lipofuscin-like compounds. Importantly, Sudan Black B treatment only slightly diminished the intensity of in vivo labeling with hydrophilic
Fig. 1. The confocal laser-scanning micrograph (a) shows red punctate Cy3-ME 20.4 in vivo labeling in cholinergic neurons of the rabbit medial septum. Selectivity of Cy3-ME 20.4 for cholinergic neurons was demonstrated by co-localization with choline acetyltransferase (ChAT) that appears green (Cy2). (b) Merged carbocyanine triple fluorescence staining of Cy3-ME 20.4 (red clusters) and immunoreactivities for ChAT (Cy2, green) and calretinin (Cy5, color-coded in blue) in the border zone between the medial and lateral septum confirmed the specificity of in vivo labeling for cholinergic neurons and its absence in non-cholinergic, calretinin-containing nerve cells. Bar = 25 μm.

Fig. 2. Electron micrographs of cholinergic neurons in the medial septum (a) and, at higher magnification, in the diagonal band of Broca (b) following photoconversion of internalized Cy3-ME 20.4. The in vivo label was predominantly present in secondary lysosomes (arrows) after two days of survival. In comparison to the photoconverted material, non-labeled primary lysosomes (arrowheads) are different in their size, shape and electron density. Prior to print, the brightness and contrast of the electron micrographic images were digitally adjusted and shading correction was performed to reduce the effects of uneven illumination. Bar = 500 nm (a), 250 nm (b).
Cy3 and of the subsequently performed Cy2-immunostaining of cholinergic neurons. Electron microscopic analysis elucidated that Cy3-ME 20.4 was predominantly located intracellularly in secondary lysosomes (Fig. 2a,b).

DISCUSSION
The granular cellular labeling of neurons in the rabbit BFB with i.c.v. applied Cy3-ME 20.4 resembled that of Cy3-192IgG in the rat BFB [7]. A subpopulation of cholinergic neurons in the MSDB was unlabeled by Cy3-ME 20.4 that also parallels the incomplete Cy3-192IgG labeling in the rat [7,11]. Recently, the counting of Cy3-192IgG-positive cells in the rat BFB revealed a medial septum > horizontal limb of the diagonal band > nucleus basalis magnocellularis gradient of labeling efficacy [23]. An apparently similar gradient was observed after the in vivo labeling with Cy3-ME 20.4; that, however, remains to be quantified. Future studies should also elucidate whether the in vivo labeling with Cy3-ME 20.4 is long-term stable or undergoes gradual clearance from the neurons, as was previously shown for Cy3-192IgG in the rat [8].

The predominant location of Cy3-ME 20.4 in secondary lysosomes, as revealed by electron microscopy, is identical to the ultrastructural localization of Cy3-192IgG in rat basal forebrain neurons [8]. This finding also corresponds with the observation that lysosomes are preferred targets for forebrain neurons [8]. This finding also corresponds with the ultrastructural localization of Cy3-192IgG in rat basal lysosomes, as revealed by electron microscopy, is identical to the ultrastructural localization of Cy3-192IgG in rat basal forebrain neurons [8]. These studies showed that muscarinic receptor antagonists assumed diminished hippocampal acetylcholine release, and confirmed neuroanatomical data on collaterals of cholinergic neurons contacting parvalbumin-immunopositive GABAergic neurons within the MSDB [25].

CONCLUSION
The high interspecies variability of the chemical nature, structure and topography of the cholinergic components in magnocellular BFB nuclei hampers the development of general functional concepts based on investigations of only one or a few animal species. Therefore, the use of ME 20.4-saporin conjugates both in primates [16,17] and in sheep [19] suggests that Cy3-ME 20.4 might be successfully employed also in primates and ungulates, and become a useful cholinergic marker to determine morphological, electrophysiological and pharmacological parameters of cholinergic neurons under physiological or pathological conditions.

REFERENCES