Amoeboid Cells Use Protrusions for Walking, Gliding and Swimming
van Haastert, Petrus; Hotchin, Neil A.

Published in:
PLoS ONE

DOI:
10.1371/journal.pone.0027532

IMPORTANT NOTE: You are advised to consult the publisher’s version (publisher’s PDF) if you wish to cite from it. Please check the document version below.

Document Version
Publisher's PDF, also known as Version of record

Publication date:
2011

Link to publication in University of Groningen/UMCG research database

Citation for published version (APA):

Copyright
Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

Take-down policy
If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Downloaded from the University of Groningen/UMCG research database (Pure): http://www.rug.nl/research/portal. For technical reasons the number of authors shown on this cover page is limited to 10 maximum.
Amoeboid Cells Use Protrusions for Walking, Gliding and Swimming

Peter J. M. Van Haastert*
Department of Cell Biochemistry, University of Groningen, Groningen, The Netherlands

Abstract
Amoeboid cells crawl using pseudopods, which are convex extensions of the cell surface. In many laboratory experiments, cells move on a smooth substrate, but in the wild cells may experience obstacles of other cells or dead material, or may even move in liquid. To understand how cells cope with heterogeneous environments we have investigated the pseudopod life cycle of wild type and mutant cells moving on a substrate and when suspended in liquid. We show that the same pseudopod cycle can provide three types of movement that we address as walking, gliding and swimming. In walking, the extending pseudopod will adhere firmly to the substrate, which allows cells to generate forces to bypass obstacles. Mutant cells with compromised adhesion can move much faster than wild type cells on a smooth substrate (gliding), but cannot move effectively against obstacles that provide resistance. In a liquid, when swimming, the extending pseudopods convert to side-bumps that move rapidly to the rear of the cells. Calculations suggest that these bumps provide sufficient drag force to mediate the observed forward swimming of the cell.

Introduction
Many cells have a mode of migration known as amoeboid movement that is characterized by frequent changes in cell shape due to the extension of protrusions [1,2]. The protrusions of an amoeboid cell, often termed pseudopods or lamellipods, are crucial for cell movement, as they determine the speed, direction and trajectory of the cell. An important aspect of cell motility is the ability of cells to respond to directional cues with oriented movement. Gradients of diffuse chemicals give rise to chemotaxis [3,4]. Other directional cues that can induce oriented movement are temperature gradients or electric fields [5,6]. These signals somehow modulate the direction of pseudopods such that, on average, cells move in the direction of the positional cues.

Recently, cell migration has been investigated using a ‘pseudopod-centred’ approach, in which large data sets are collected on the spatiotemporal properties of pseudopods that are extended by cells in the absence or presence of directional cues [7–13]. In the absence of external cues, cells are more likely to extend a new pseudopod in the direction of the existing pseudopods, and their directions are alternating to the left and right. With these characteristics cells move with strong persistence to cover a large distance in a short period [11,14]. A shallow gradient of chemoattractant induces a small positional bias, such that cells are more likely to start a new pseudopod at the side of the cell facing the highest chemoattractant concentration [8,9]. In addition, cells with multiple pseudopods often retract the pseudopod that is positioned in the worse direction relative to the cAMP gradient [7]. As pseudopods are generally extended perpendicular to the cell surface, this positional bias where pseudopods begin will direct the cell towards the attractant [15]. Cell migration and chemotaxis generally are studied in two dimensions as the cells crawl over various solid substrates. However, in vivo, cells move in a complex three dimensional environment [16,17]. Such cells may experience obstacles such as other cells, soil particles, cavities, or liquids. Movement in a complex environment may require the ability to generate substantial force to resist obstacles, as well as the ability to swim [18,19].

It has been suggested that pseudopods are self-organizing structures, which means that their organization is largely intrinsically controlled; although external signals can trigger the formation and location of a pseudopod, the pseudopod otherwise follows a typical life cycle [20]. In our studies on the movement in Dictyostelium cells we observed that wild type and many mutant cells extend a new pseudopod every ~15 seconds. After ~12 seconds, the pseudopod suddenly stops growing. In wild type cells on a solid support about 75% of the pseudopods make contact with the substrate, followed by outward expansion of the pseudopodia thereby contributing to the forward movement of the cell [9,14]. Well after the pseudopodia cease to expand, they remain recognizable as convex extensions at the side of the cell. We have made some investigations to such “side-bumps”, because likely they are places of attachment of the cell to the substrate [21–24] and therefore may contribute to movement in an environment with obstacles providing enhanced resistance. Surprisingly, cells in suspension have side bumps that move to the rear of the cell, potentially providing a drag force that may contribute to the forward movement of cells in suspension. Previously it has been suggested that traveling waves of surface deformations may explain the movement of cyanobacteria [25]. Here I report on the conversion of pseudopods to side-bumps and their potential role in movement of amoeboid cells on solid supports and in suspension.
Results and Discussion

Walking of cells on a solid support

The algorithm Quimp for pseudopod analysis describes the cell boundary as a polygon of ~100 nodes [26]. Each node has an address, and therefore its speed and convexity can be determined in subsequent images. Pseudopods are identified as extending convex areas, with the central node of this convex area assigned as the tip of the pseudopod. We followed the tip node during and after pseudopod extension. The speed of the pseudopod tip relative to the substrate during the extending phase of the pseudopod is very high (~30 μm/min). At the end of the extension period, the speed of the tip node declines abruptly to nearly zero (Fig 1A). Interestingly, these stationary tip nodes are still present in convex areas, as identified by the computer algorithm [27] and by visual inspection of individual cells (Fig 1B). It thus appears that pseudopods frequently convert to convex bump at the side of the cell. Since the cell moves forward, the stationary bumps are found after about 1 minute in the rear of the cell, where they are retracted.

Pseudopods are formed at the front alternating to the right and left [11,14], and therefore the sideway extensions are also alternating right/left from front to tail (Fig 1B). Previous experiments suggest that positions where pseudopods convert to side bumps are places of cell adhesion to the substrate [21–24]. Therefore, cell movement on a solid medium has the appearance of walking, because cells have sideway stationary “feet”; cells retract these feet in the rear while cells extend pseudopod at the front that become new feet.

Traveling waves of convexities in tail-attached cells

Dictyostelium cells occasionally appear to be attached to the substrate exclusively at the tail region, whereas other parts of the cell move freely in suspension (Fig. 1C). This interpretation is based on the observations that i) the tail touches the surface in the focal plane of the tail ii) the position of the tail does not change (thus the tail does not move), and iii) the rest of the cell can move; sometimes the cell is relatively stationary as in figure 1C (and therefore pseudopod and bump data can be easily collected for a few minutes) and sometimes cells move actively to the right and left (‘‘waving’’). In wild type cells, about 5–10% of the cells show this behavior. In some mutants, such as talinB-null cells, tail-attached movement is very common (83% of the cells). Tail-attached cells extend pseudopods at nearly the same frequency compared to surface-attached cells (Table S1). However, pseudopods grow for a shorter period and are also smaller. As in cell body-attached cells, the convex pseudopod tips frequently convert to side bumps. Interestingly, these side bumps travel to the rear of the cell (Fig. 1C) at a speed of about -13 μm/min; the minus sign is to indicate that side-bumps travel in a direction opposite to extending pseudopods. After approximately 1 minute the side bumps have arrived at the tail of the cell.

Swimming cells use pseudopods that convert to paddles

Occasionally the tail-attached cells detach from the substrate. Although such cells will slowly sink, they can be followed while completely free in suspension during a few minutes (Fig. 2A). We followed 8 tail-attached cells after they detach from the substrate. Such cells continue to form pseudopods and side bumps that travel to the rear of the cell, with essentially the same properties as tail-attached cells. The swimming cells move forward (i.e. in the direction of the extended pseudopods) at a slow speed of ~ 3 μm/ min (Table S1). The trajectories of swimming cells reveal persistent directional movement (Fig. 2C).
cytosol flows into the pseudopod and the old pseudopod tip merges with the cell body. Interestingly, these cells have enhanced speed of movement, 17 μm/min, compared to 10 μm/min for wild type cells. The enhanced speed was also observed for other mutants with reduced adhesion: *talinA*-null cells with defects in the cytoskeleton [29] and *sadA*-null cells with defects in adhesion molecules [30]. The adhesion-defective mutant cells move nearly with the speed of the forward moving pseudopods, suggesting that cells in the absence of strong adhesion glide over the substrate. Can cells with reduced adhesion move against obstacles? Wild-type and *gbpD*-null cells were covered with a block of agar to provide resistance. Whereas movement of wild type cells under agar is only slightly slower than movement on agar, the speed of *gbpD*-null cells is strongly reduced from 17 μm/min to 5 μm/min under agar.

How fast can cells glide, walk and swim?

The maximal speed of walking or gliding cells on a substrate depends on the frequency \( F \) and size \( l_p \) of extending pseudopods according to

\[
v_c = F l_p a b \cos \alpha,
\]

where \( a \) = fraction of pseudopods that contribute to movement, \( b \) = positional overlap of pseudopods, and \( \alpha \) = the mean angle between pseudopod and forward cell movement. The experimental data (Table S2) predict that wild type cells can walk at a maximal speed of 11.4 μm/min, which is close to the observed speed of 10.4 μm/min.

Gliding *gbpD*-null cells extend pseudopods at nearly the same frequency as wild type cells, but with some subtle differences: pseudopods are larger and rarely retracted, thereby providing more forward movement to the cells. Using these measured pseudopod properties, the predicted speed of gliding cells is 17.8 μm/min, which is close to the observed speed of 17.3 μm/min.

Apparently, the lower speed of wild type cells is attributed to the shorter pseudopods and the more frequent retraction of new pseudopods. Why? Movement of substrate-attached cells can be regarded as the transport of material from behind the attachment zone at the rear of the cell to before the attachment zone at the front of the cell. Materials to be transported are cytoplasm with organelles and plasma membrane. Cytoplasm may flow freely to the front by hydrostatic pressure [31], but transport of membrane may be restricted by adhesion of the cell to the surface. The reduced flow of membrane may lead to increased membrane tension at the front of the cell, which could impair pseudopod growth and induce retraction of newly formed pseudopods, as is observed in wild type cells.

How fast can cells swim in liquid? An object moving in a fluid will experience a drag force that, according to Newton’s third law, will induce an equal counterforce on the object. For small objects the drag occurs at low Reynolds numbers [32] and therefore the drag force \( \mathbf{F}_D \) is given by the Stokes equation:

\[
\mathbf{F}_D = -6 \pi \rho R v,
\]

where \( \rho \) is the viscosity of the fluid, \( R \) is the radius of the frontal cross sectional area of the object and \( v \) is the speed of the object. A swimming *Dictyostelium* cell may experience three forces: the drag induced by the extending pseudopod, the drag induced by the rearward moving bumps, and the drag induced by the total movement of the cell. Assuming that these three drag forces experience the same viscosity, the balanced force equation yields:

\[
v_c R_c n_c \bar{v}_b + n_p R_p \bar{v}_p = 0,
\]

where the subscripts indicate cells (c) bumps (b) or pseudopods (p), \( \bar{v} \) indicates the mean speed of the object in the direction of the cell and \( n \) indicates the mean number of moving bumps or bumps.
pseudopods. We measured these properties for cells in suspension (Table S2), predicting an average swimming rate of \(3.5\ \text{mm/min}\), which is close to the observed rate of \(3\ \text{mm/min}\) for freely swimming *Dictyostelium* cells.

**Conclusions**

Polarized amoeboid cells may move by three modes, walking, gliding and swimming (Fig. 4). Investigations to swimming cells are complicated because cells in suspension will sink. In isodense suspensions of ficol cells can be observed easily [18], but ficol may induce an osmotic response or it may provide some form of structure to move on, as ficol may not be a Newtonian solution [33]. We investigated cells that detached from the surface by which they move in suspension, and obtained a swimming speed of \(3\ \text{mm/min}\), about 4-times slower that the crawling speed of \(12\ \text{mm/min}\). In ficol, cells swim and crawl at about the same speed of \(4.2\ \text{mm/min}\) and \(3.8\ \text{mm/min}\), respectively [18]. The current observations suggest that the three moving modes, walking, gliding and swimming, are all based on pseudopods, which are extending convex areas of the cell boundary. At the end of the extension period the convex pseudopods often convert to convex bumps at the side of the cell, which move in about 1 minute to the rear of cells in suspension. The \(43\) rearward moving bumps provide sufficient drag force to explain the observed forward movement of swimming cell. Pseudopods of cells attached to a substrate also convert to side bumps that end up in the rear of the cell after about 1 minute. However, these bumps do not move, but are fixed to the substrate, presumably because they form attachment sides of the cell to the substrate [21–24]. Sur-

![Figure 3. Gliding cells.](image)

**Figure 3. Gliding cells.** A. Adhesion of wild type and gpdD-null cells, expressed as the fraction of cells that detach from a plastic surface after shaking in buffer for 1 hour [28]. B. Speed of wild-type and mutant cells on a solid support and under a block of agar.

doi:10.1371/journal.pone.0027532.g003

![Figure 4. Model of gliding, walking and swimming.](image)

**Figure 4. Model of gliding, walking and swimming.** All cells extend pseudopods at a frequency of \(4\) per minute. In gliding cells, pseudopods are large and all contribute to forward moving; side bumps are rare. In walking cells, pseudopods are smaller and only \(75\%\) of pseudopods contribute to forward movement; these pseudopods convert to side-bumps that are stationary relative to the substrate. In swimming cells, pseudopods are small and convert to side-bumps that move to the rear of the cell. On average walking and swimming cells have \(3\) side-bumps (see table S1).

doi:10.1371/journal.pone.0027532.g004
prisingly, adhesion-defective mutant cells move at a much higher speed than wild type cells on a smooth surface, but exhibit a much lower speed then wild type cells when exposed to strong resistance, such as movement under a block of agar. *Dictyostelium* cells live in a heterogeneous environment composed of soil particles and surrounding liquid. Cells move probably most of the time on 2D surfaces of soil particles, but may also experience clfts and obstacles. Cells have the ability to walk on these complicated surfaces with stronger adhesion, and to swim in water, all using essentially the same cycle of pseudopod formation with conversion to sideway extensions. This may allow the cells to effectively move optimally in its physically complex habitat.

**Methods**

The strains used are wild type AX3 and *gbdD*-null cells lacking gene encoding a Rap-GEF [28]. Cells were grown in HG5 medium (contains per liter: 14.3 g oxoid peptone, 7.15 g bacto yeast extract, 1.36 g *Na2HPO4·12H2O*, 0.49 g *KH2PO4*, 10.0 g glucose), harvested in PB (10 mM *KH2PO4*/*Na2HPO4*, pH 6.5), and allowed to develop in 1 ml PB for 5 hours in a well of a 6-wells plate (Nunc). Movies of starved cells in PB on an objective glass were recorded with an inverted light microscope (Olympus Type CK40 with 20x objective) and images were captured at a rate of 1 frame/second with a JVC CCD camera. For movement under agar, cells were covered with an approximately cubic (length 3 mm) block of 1.5% agar in PB. Excess buffer was removed and movies were recorded as described above.

Images were analyzed with the fully automatic algorithm Quimp3 [26]. In short, the program uses an active contour analysis tools: PVH. Wrote the paper: PVH.

**Supporting Information**

**Table S1** Properties of pseudopods, bumps and cells for three modes of movement.

**Table S2** Three modes of pseudopod-based movement

**Author Contributions**

Conceived and designed the experiments: PVH. Performed the experiments: PVH. Analyzed the data: PVH. Contributed reagents/materials/analysis tools: PVH. Wrote the paper: PVH.

**References**