Degradation of 4-fluorophenol by *Arthrobacter* sp. strain IF1

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**Abstract** A Gram-positive bacterial strain capable of aerobic biodegradation of 4-fluorophenol (4-FP) as the sole source of carbon and energy was isolated by selective enrichment from soil samples collected near an industrial site. The organism, designated strain IF1, was identified as a member of the genus *Arthrobacter* on the basis of 16S ribosomal RNA gene sequence analysis. *Arthrobacter* strain IF1 was able to mineralize 4-FP up to concentrations of 5 mM in batch culture. Stoichiometric release of fluoride ions was observed, suggesting that there is no formation of halogenated dead-end products during 4-FP metabolism. The degradative pathway of 4-FP was investigated using enzyme assays and identification of intermediates by gas chromatography (GC), GC–mass spectrometry (MS), high-performance liquid chromatography, and liquid chromatography–MS. Cell-free extracts of 4-FP-grown cells contained no activity for catechol 1,2-dioxygenase or catechol 2,3-dioxygenase, which indicates that the pathway does not proceed through a catechol intermediate. Cells grown on 4-FP oxidized 4-FP, hydroquinone, and hydroxyquinol but not 4-fluorocatechol. During 4-FP metabolism, hydroquinone accumulated as a product. Hydroquinone could be converted to hydroxyquinol, which was further transformed into maleylacetic acid and β-ketoadipic acid. These results indicate that the biodegradation of 4-FP starts with a 4-FP monooxygenase reaction that yields benzoquinone, which is reduced to hydroquinone and further metabolized via the β-ketoadipic acid pathway.

**Keywords** Biodegradation . 4-Fluorophenol . Hydroquinone . Monooxygenase . Fluoride

**Introduction**

During the past decades, widespread application of fluoroaromatic compounds as agrochemicals and pharmaceuticals has lead to an increased occurrence of environmental contaminants containing fluorine (Key et al. 1997). Fluorinated compounds are rare in nature (Harper and O’Hagan 1999). The stability of the carbon–fluorine bond (116 kcal/mol in CH₃F, compared to 81 kcal/mol for the carbon-chlorine bond in CH₃Cl) makes most fluorine-containing compounds much more resistant to biodegradation than their unsubstituted analogs (Key et al. 1997). Furthermore, the van der Waals radius of fluorine is small (1.47 Å, in between that of a hydrogen and an oxygen). Yet, the electronegativity of fluorine causes strong polarization of C–F bonds, and fluorine substituents may be involved in biological interactions (Howard et al. 1996).

Regardless of the recalcitrance of most fluoroaromatics to biodegradation, several bacterial cultures have been described that aerobically degrade fluoroaromatic acids (Olthmanns et al. 1989; Engesser et al. 1980; Harper and Blakley 1971; Schloemann et al. 1990). Research on bacterial fluorophenol degradation has been limited to studies with whole cells, cell extracts, or purified enzymes from *Rhodococcus* species that were obtained by enrichment with other aromatic compounds as a growth substrate.
with Teflon-lined screw caps. 100-ml flasks filled to 25% of their volume and were closed at 30°C on a rotary shaker (180 rpm). Cultures were grown in a media obtained by adding 15 g/l of Difco agar. Strain IF1 was grown on a medium (Difco Laboratories). When required, the solid medium was transformed into a liquid medium by the addition of 0.5 g/l of yeast extract. The liquid medium was sterilized separately for growth on 1 mM 4-FP liquid medium. Growth and fluoride release were again monitored to verify 4-FP degradation. Strains capable of 4-FP degradation as a sole source of carbon and energy were used for further experiments. Strain IF1 was deposited at the Centraalbureau voor Schimmelcultures, Utrecht, The Netherlands, under accession number NCCB 100218.

Sequencing of the 16S rRNA gene For cloning of the 16S ribosomal ribonucleic acid (rRNA) gene, a single colony of strain IF1 was directly used for polymerase chain reaction (PCR) amplification. The primers 63f (5′-CAGGCCTAAACCAATGCAAGTC-GGT-3′) and 1387r (5′-GGGCGGWGTGTAACATGCAAGTC-3′) were used for PCR amplification. The PCR reaction mixture (50 μl) contained Taq PCR buffer, 2.5 mM MgCl₂, 20 pmol of each appropriate primer, 200 mM of each deoxyribonucleotide triphosphate, 1 U Taq DNA polymerase, and biomass of strain IF1. The PCR conditions were 94°C for 10 min followed by 1 min at 95°C, 1 min at 55°C, 1.5 min at 72°C, and 5 min at 72°C. The resulting fragments were cloned into the pCR4-TOPO vector (Invitrogen, Carlsbad, CA) and transformed into Top10 cells. The transformed cells were plated on LB plates containing 0.5 mg/ml of ampicillin, and the positive colonies were used for plasmid isolation and sequencing.

Phylogenetic analysis Alignments of the 16S rRNA gene were made using sequences downloaded from the Ribosomal database project II (RDP II; Cole et al. 2005), after searching for nearest neighbors using the sequence match tool. Further searches were conducted using BLAST and FASTA of the European Molecular Biology Laboratory (EMBL) database for 16S rRNA gene sequences that are closely related to the 16S rRNA gene of strain IF1 but not available from the RDP II. Alignments were subsequently made using the profile alignment option in ClustalX (Thompson et al. 1997), refined using BioEdit ver 7.0.5.2 (Hall 1999), and subsequently parsed through Gblock

Materials and methods

Media and growth conditions NB medium contained 8 g of Nutrient Broth (Difco) per liter. Mineral salts medium (MM) contained per liter 5.3 g Na₂HPO₄, 12H₂O, 1.4 g KH₂PO₄, 0.2 g MgSO₄ 7H₂O, 0.5 g (NH₄)₂SO₄, 5 ml trace metals solution (Janssen et al. 1985), and 10 mg of yeast extract (Difco Laboratories). When required, the solid medium was obtained by adding 15 g/l of Difco agar. Strain IF1 was grown on a Luria–Bertani medium (LB) at 37°C on a rotary shaker. cultivation was performed with a phase as the sole carbon and energy source. The cultures were incubated at room temperature on a rotary shaker (150 rpm), and 40% of the suspension was transferred to a new flask containing fresh medium every 15 days. During this time, growth (optical density at 600 nm) and liberation of fluoride were monitored. Samples of the enrichment culture were periodically spread onto minimal salts agar plates containing 1 mM 4-FP and onto NB plates as soon as growth on 4-FP was established. Pure cultures were obtained by repetitive streaking onto solid MM containing 4-FP and tested separately for growth on 1 mM 4-FP liquid medium. Growth and fluoride release were again monitored to verify 4-FP degradation. Strains capable of 4-FP degradation as a sole source of carbon and energy were used for further experiments. Strain IF1 was deposited at Centraalbureau voor Schimmelcultures, Utrecht, The Netherlands, under accession number NCCB 100218.

Enrichment and isolation of 4-FP-degrading cultures A variety of soil samples, collected from different sites in The Netherlands that are contaminated with halogenated aliphatic compounds (such as monochlorobenzene, hexachlorobenzene, and trichloropropane), were used as the initial inoculum for the 4-FP enrichments. The soil samples were used to inoculate flasks containing 30 ml of sterile minimal salts medium and 1 mM of 4-FP, supplied in the liquid phase as the sole carbon and energy source. The cultures were incubated at room temperature on a rotary shaker (150 rpm), and 40% of the suspension was transferred to a new flask containing fresh medium every 15 days. During this time, growth (optical density at 600 nm) and liberation of fluoride were monitored. Samples of the enrichment culture were periodically spread onto minimal salts agar plates containing 1 mM 4-FP and onto NB plates as soon as growth on 4-FP was established. Pure cultures were obtained by repetitive streaking onto solid MM containing 4-FP and tested separately for growth on 1 mM 4-FP liquid medium. Growth and fluoride release were again monitored to verify 4-FP degradation. Strains capable of 4-FP degradation as a sole source of carbon and energy were used for further experiments. Strain IF1 was deposited at Centraalbureau voor Schimmelcultures, Utrecht, The Netherlands, under accession number NCCB 100218.

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(Castresana 2000) to remove ambiguously aligned sections and increase the robustness of the data. Phylogenetic trees were determined using the neighbor-joining (NJ) method. Evolutionary distances for the global tree were calculated using the Kimura-2-parameter model with a transition/transversion ratio of 2 (Fig. 1). Further trees were constructed in Phylip version 3.6a3 (J. Felsenstein), using maximum parsimony and maximum likelihood methods. All NJ trees were tested statistically by means of bootstrap analysis.

Preparation of cell extracts Cells were grown in MM with 4 mM 4-FP, harvested by centrifugation at 5,000× for 15 min, washed with ice-cold TD buffer (0.1 M Tris–HCl, pH 7.5, and 0.1 M 1,4-dithiothreitol), and stored at −20°C until further use. The frozen cells were thawed, resuspended in TD buffer, and incubated with lysozyme (50 mg/ml) for 1 h at 30°C. A French press was used to disrupt the cells, and the crude extracts were centrifuged at 10,000×g for 15 min to separate the soluble from the particulate fraction. The supernatant was used for further experiments. Protein concentrations were determined with the Biorad protein assay kit.

Enzyme assays Catechol 1,2-dioxygenase and catechol 2,3-dioxygenase activities were measured spectrophotometrically at 260 and 375 nm, respectively. The reaction mixtures contained 0.1 mM catechol, TD buffer, and cell-free extract (0.1 mg of protein) in a final volume of 1 ml. 4-Fluorocatechol 1,2-dioxygenase was measured as catechol 1,2-dioxygenase but with 4-fluorocatechol instead of catechol as the substrate.

The 4-FP monooxygenase activity was measured spectrophotometrically by following the consumption of NADH at 340 nm in a reaction mixture containing cell-free extract (0.1 mg protein), 0.1 mM 4-FP, 0.1 mM NADH, and buffer in a total volume of 1 ml. The observed rates were corrected for substrate-independent NADH oxidation.

Hydroxymuconic semialdehyde dehydrogenase was measured at 375 nm. Reaction mixtures contained (in a final volume of 1 ml) about 0.05 mM freshly prepared hydroxymuconic semialdehyde, TD buffer, 0.1 mM NAD, and cell-free extract (0.1 mg of protein). Hydroxymuconic semialdehyde was obtained by incubation of catechol with the cell-free extract of Pseudomonas putida mt-2 as described previously (Mars et al. 1998).
Hydroquinone dioxygenase was assayed spectrophotometrically by monitoring the change in absorbance between 230 and 330 nm in a reaction mixture of 1 ml final volume containing 0.1 mM hydroquinone, TD buffer, and cell-free extract (0.1 mg of protein). Hydroquinone hydroxylase and hydroxyquinol dioxygenase were assayed by using a fiber optic oxygen sensor. Reaction mixtures contained 1 mM of substrate (4-FP, hydroquinone, 4-fluorocatechol, hydroxyquinol, or catechol), MM, and cell suspension (0.5 mg/ml protein) in a final volume of 1.25 ml.

Oxygen uptake measurements Strain IF1 was grown with glucose or 4-FP as the sole carbon source and harvested by centrifugation at 6,000×g for 15 min at 4°C. Cells were resuspended in MM, and O2 consumption was measured with a fiber optic oxygen sensor (MOPS-1, ProSense BV, Hannover, Germany). All reactions were performed in a stirred vessel at room temperature. The reaction mixtures contained 1 mM of substrate (4-FP, hydroquinone, 4-fluorocatechol, hydroxyquinol, or catechol), MM, and cell suspension (0.5 mg/ml protein) in a final volume of 1.25 ml.

Analytical methods For capillary gas chromatography (GC), 4 ml samples were extracted with 1 ml of diethyl ether. A model 6890 gas chromatograph (Hewlett-Packard) equipped with a flame ionization detector and a HP-5 column (Agilent 19091J-413, 30 m×0.25 mm×0.25 μm) were used for the analysis. GC-mass spectrometry (MS) analysis was carried out with a model 5973 mass selective detector (Hewlett-Packard) coupled to a HP 6890 series injector and a HP1 column (Agilent 19091Z-433; 30 m× 0.25 mm×0.25 μm). High-performance liquid chromatography (HPLC) was carried out using a Chrompack C18 column (10 cm×5 mm) connected to a Jasco UV-1575 detector, which monitored absorbance at 214 and 254 nm, and operated with Jasco PU-980 pumps and a Jasco AS-1555 sampler. The mobile phase was water/acetonitrile (70:30), 5 mM potassium phosphate (pH=3), and 340 mg/l sodium dodecyl sulfate, and the flow rate was 1 ml/min.

Liquid chromatography (LC)-MS was carried out with a ZMD Micromass spectrometer, equipped with a XTerra MS, Symmetry Shield C8 column (4.6×150 mm), a Waters 996 photodiode array detector, and a Waters 2690 separations module. Samples of 20 μl were analyzed, and compounds were isocratically eluted at a flow rate of 1 ml/min with a solution of water/acetonitrile (80:20) and 10 mM formic acid.

Concentrations of free fluoride in the culture supernatants were measured with a fluoride electrode (model 96-09, Thermo Russell, Scotland). Fresh sodium fluoride standards were prepared for calibration curves.

Chemicals 4-FP (>98%) was obtained from Sigma-Aldrich (Steinheim, Germany). All chemicals were of the highest purity grade available (Sigma-Aldrich; Acros Organics, Geel, Belgium). The compound 4-fluorocatechol was kindly provided by Dr. Erik de Vries. Purity of 4-FP, 4-fluorocatechol, and hydroquinone was checked by HPLC.

Nucleotide sequence accession numbers The 16S rRNA sequence of strain IF1 was deposited at GenBank with the accession no. DQ425093.

Results

Isolation of a 4-FP-degrading bacterium To obtain a bacterial culture that is able to use 4-FP as carbon and energy source for growth, enrichments were performed and followed over time. Two months of selective enrichment by repeated transfer of samples from a culture that displayed growth on 4-FP to fresh 4-FP containing media resulted in a microbial consortium that was capable of growth on 4-FP as a sole source of carbon and energy. Samples of the consortium were repeatedly plated onto NB agar plates and MM agar plates containing 4-FP. This procedure resulted in the isolation of three pure strains named IF1, IF2, and IF3. The strains were restreaked on MM supplemented with 4-FP and inoculated in liquid cultures with 1 mM 4-FP. Fluoride liberation was observed for all three strains but not in control incubations to which no bacterial inoculum was added. All the three strains showed growth, as monitored at 600 nm, and thus were able to use 4-FP as a sole carbon and energy source. After 5 days of incubation in liquid culture supplied with 1 mM 4-FP, strain IF1 reached 100% fluoride release and an optical density (OD) of 0.119 at 600 nm, while strain IF3 reached an OD of 0.076 and released 54% of the theoretical amount of fluoride ions. When a mixed culture of the three strains in liquid media was used, an OD600 of 0.217 and 100% fluoride release were measured after 4 days. Apparently, the mixed culture contained, even after prolonged adaptation, organisms with varying efficiencies of 4-FP utilization. Because strain IF1 showed the highest degradation rates combined with stoichiometric release of fluoride, this organism was chosen for further studies.

Microbiological characterization Strain IF1 is a Gram-positive motile bacterium with a rhodococcus lifecycle. The optimal temperature for growth is 30°C. The 16S rRNA gene sequence was determined. Initial searches against the RDP II and EMBL rRNA databases resulted in very close associations with members of the genus Arthrobacter, the closest of which were the quinaldine-degrading strain Arthrobacter sp. KA1-1 (Overhage et al. 2005) and the 4-
nitroguaiaicol-degrading actinobacterium *Arthrobacter nitroguajacolicus* sp. nov (Kotouckova et al. 2004). The 16S rRNA gene sequences of these organisms were greater than 99% identical to IF1’s. The phylogenetic analysis places the isolates in the phylum *Actinobacteria* and the genus *Arthrobacter* (Fig. 1). The subtree that were obtained shows the detailed relationship of isolate IF1 to other members of the genus *Arthrobacter*. The topologies of all the trees that were obtained during statistical analysis were in agreement and clearly placed this isolate in the genus *Arthrobacter* (data not shown). Furthermore, the change in form during the growth cycle between rod and coccus is typical of the lifecycle of the *Arthrobacter* genus.

**Catabolic activities of Arthrobacter sp. strain IF1** To study the degradation potential of strain IF1, a range of organic compounds were tested as growth substrates. Cells were inoculated into MM, and different organic substrates were added at a concentration of 1 mM. After 72-h incubation growth, substrate disappearance and halide release were measured.

Growth and substrate removal were found when catechol, hydroquinone, hydroxyquinol, benzoate, phenol, 4-fluorocinnamic acid, and 4-nitrophenol were used as substrates. The organism did not grow on 2-fluorophenol, 3-fluorophenol, 4-chlorophenol, 4-bromophenol, 4-iodophenol, fluoroacetate, trifluoroacetate, fluoroacetamide, trifluoroethanol, or on 2-bromoethanol. The fact that strain IF1 is capable of growth on catechol, hydroquinone, and hydroxyquinol but not on 4-fluorocatechol indicates that 4-fluorocatechol is not the most likely intermediate in the 4-FP pathway, although toxic effects could also play a role.

To test the range of 4-FP concentrations tolerated by strain IF1, experiments were conducted in sealed flasks with 4-FP at concentrations of 1 to 7 mM as a sole carbon and energy source. Control assays without 4-FP showed no growth or release of fluoride, and sterile controls showed no abiotic loss of 4-FP. Between 1 and 4 mM 4-FP, the substrate was completely consumed, stoichiometric release of fluoride was observed, and biomass increased linearly with the amount of 4-FP added (Fig. 2a). This indicates that the degradation of 4-FP by strain IF1 does not give large amounts of fluorinated dead-end products. Concentrations of 4-FP above 4 mM caused a toxic effect on the growth of IF1 (Fig. 2b). The use of 5 mM 4-FP promoted growth, but a longer lag time was observed. At 7 mM 4-FP, no growth occurred even after 15 days of incubation.

**Growth on 4-FP and formation of metabolites** A batch culture of strain IF1 supplied with 1 mM 4-FP as the only source of carbon and energy was monitored in time. Growth was accompanied by an increase in biomass, decrease in 4-FP, and formation of fluoride (Fig. 3). After 120 h, there was complete conversion of 1 mM 4-FP, and 1 mM fluoride was formed, indicating that there was no transient accumulation of fluorinated intermediates over the whole growth period. When cells grown on 4-FP were incubated in the presence of the iron chelator 2,2’-dipyridyl, no 4-FP degradation occurred, and no fluoride was released in the medium. This indicates that initial or further enzymes involved in 4-FP metabolism require ferrous ions for activity.

To isolate intermediates of the degradation of 4-FP, samples from a batch culture containing 4 mM 4-FP were taken at appropriate intervals and analyzed by GC, HPLC, and LC-MS. Metabolites that were detected were numbered in order of time of appearance in the culture. GC analysis indicated that at least four metabolites were formed and degraded over time (Table 1). Metabolite I, the earliest

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**Fig. 2** Fluoride release and biomass formation by strain IF1 with different concentrations of 4-FP. **a** Stoichiometric fluoride release. Symbols: *squares*, optical density at 600 nm; *circles*, fluoride concentration. **b** Growth in the presence of different levels of 4-FP. The initial concentrations used were: *triangles*, 1 mM 4-FP; *circles*, 4 mM 4-FP; *diamonds*, 5 mM 4-FP; *squares*, 7 mM 4-FP.
product that was observed, had the same retention time as an authentic hydroquinone standard and metabolite II had the same retention time as a standard of hydroxyquinol.

GC-MS analysis showed that the mass spectrum of metabolite I was indeed similar to that of the hydroquinone standard, with a molecular ion peak at 110 and 55 m/z. Metabolite II gave a molecular ion peak at 126 and 63 m/z, which is typical for hydroxyquinol, and the spectrum coincided with that of a standard. Metabolites V and VI were detected by GC but did not ionize in GC-MS under the conditions tested and were not identified.

When the supernatant of IF1 cells growing on 4 mM 4-FP were subjected to HPLC analysis, four peaks were detected (Table 1). Metabolite I had the same retention time as the hydroquinone standard, and metabolite II coeluted with an authentic standard of hydroxyquinol. Hydroquinone was detected in culture supernatants by HPLC between 7.5 and 51 h (Fig. 4). LC-MS analysis of the extracted samples of the supernatant revealed the presence of a compound (VII) of which the negative ion spectrum shows a molecular ion peak at 159 m/z and a peak at 80 m/z, which are expected for the negative ionization of 3-oxoadipate.

The above results suggest that 4-FP is initially converted to hydroquinone. The most likely enzyme involved in such a conversion is a 4-FP monoxygenase.

### Enzyme activities

To test inducibility of enzymes involved in 4-FP degradation, cells of *Arthrobacter* sp. strain IF1 were grown on glucose or 4-FP, washed, and tested for oxygen consumption in the presence of different substrates. Washed suspensions of cells grown on 4-FP rapidly oxidized 4-FP, hydroquinone, and 1,2,4-benzenetriol (hydroxyquinol) without a lag. Catechol and 4-fluorocatechol did not stimulate

### Table 1 Retention times in GC and HPLC analyses of metabolic intermediates formed by cells of strain IF1 growing on 4-FP and of some authentic standards

<table>
<thead>
<tr>
<th>Compound</th>
<th>GC retention time (min)</th>
<th>HPLC retention time (min)</th>
<th>m/z of observed fragment ions detected by GC-MS</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Metabolites</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>I</td>
<td>20.7</td>
<td>1.3</td>
<td>110, 81, 54</td>
</tr>
<tr>
<td>II</td>
<td>24.3</td>
<td>1.1</td>
<td>126, 80, 52</td>
</tr>
<tr>
<td>III</td>
<td>–</td>
<td>0.75</td>
<td>–</td>
</tr>
<tr>
<td>IV</td>
<td>–</td>
<td>2.6</td>
<td>–</td>
</tr>
<tr>
<td>V</td>
<td>16.5</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>VI</td>
<td>19.4</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>VII</td>
<td>–</td>
<td>3.1</td>
<td>159, 80</td>
</tr>
<tr>
<td><strong>Authentic standards</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>4-FP</td>
<td>15.7</td>
<td>3</td>
<td>112, 83, 57</td>
</tr>
<tr>
<td>4-Fluorocatechol</td>
<td>14.4</td>
<td>2</td>
<td>128, 82, 51</td>
</tr>
<tr>
<td>Hydroquinone</td>
<td>20.7</td>
<td>1.3</td>
<td>110, 81, 54</td>
</tr>
<tr>
<td>Hydroxyquinol</td>
<td>24.3</td>
<td>1.1</td>
<td>126, 80, 52</td>
</tr>
<tr>
<td>Catechol</td>
<td>19.1</td>
<td>–</td>
<td>–</td>
</tr>
</tbody>
</table>

*Elution followed at 254 nm

**Table 2 Substrates oxidized by 4-FP and glucose grown cells of *Arthrobacter* sp. strain IF1**

<table>
<thead>
<tr>
<th>Assay substrate</th>
<th>O2 uptake (μmol/min/mg of protein) by whole cells after growth on</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Fluorophenol</td>
</tr>
<tr>
<td>4-Fluorophenol</td>
<td>0.356</td>
</tr>
<tr>
<td>Hydroquinone</td>
<td>0.313</td>
</tr>
<tr>
<td>Hydroxyquinol</td>
<td>0.195</td>
</tr>
<tr>
<td>4-Fluorocatechol</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Catechol</td>
<td>&lt;0.001</td>
</tr>
</tbody>
</table>

* Oxygen consumption was measured with an oxygen sensor as described in Materials and Methods. All substrate were used at a concentration of 1 mM
Table 3  Specific activities of enzymes in crude extracts of strain IF1 pregrown in 4-FP or glucose

<table>
<thead>
<tr>
<th>Assay substrate</th>
<th>Enzyme tested</th>
<th>Specific activity (U/mg of protein) after growth on</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>4-Fluorophenol</td>
</tr>
<tr>
<td>Catechol</td>
<td>1,2-Dioxygenase</td>
<td>&lt;0.01</td>
</tr>
<tr>
<td></td>
<td>2,3-Dioxygenase</td>
<td>&lt;0.01</td>
</tr>
<tr>
<td>4-Fluorocatechol</td>
<td>Dioxygenase</td>
<td>&lt;0.01</td>
</tr>
<tr>
<td>4-Fluorophenol</td>
<td>Monoxygenase</td>
<td>0.12</td>
</tr>
<tr>
<td>Hydroquinone</td>
<td>Monoxygenase</td>
<td>&lt;0.01</td>
</tr>
<tr>
<td>Hydroxymuconic semialdehyde</td>
<td>Dehydrogenase</td>
<td>&lt;0.01</td>
</tr>
<tr>
<td>Hydroxyquinol</td>
<td>Dioxygenase</td>
<td>0.195</td>
</tr>
</tbody>
</table>

Discussion

This study reports the isolation and characterization of *Arthrobacter* strain IF1, an organism capable of growth with 4-FP as a sole source of carbon and energy. The biodegradation of 4-FP by strain IF1 was analyzed by GC, GC-MS, HPLC, LC-MS, oxygen uptake experiments, and measurements of enzymatic activities. A metabolic pathway is proposed on the basis of the results of this study (steps 1 and 2, *ortho*-cleavage) and by analogy with other systems.
We suggest that the degradation of 4-FP in strain IF1 starts with the conversion by a monooxygenase to benzoquinone, which is immediately reduced to hydroquinone. Hydroquinone then undergoes a further hydroxylation to form hydroxyquinol. This benzenetriol is the ring fission substrate, which is transformed by ortho-cleavage and yields maleylacetate, which is further converted to 3-oxoadipate.

Up to now, two main metabolic routes for the aerobic degradation of halogenated phenols have been described in the literature. In bacteria that degrade mono- and dichlorophenols, a degradation pathway is usually observed in which the substituted phenol is hydroxylated to the corresponding catechol, which is followed by ortho-cleavage of the aromatic ring (Haggblom 1992; Hollender et al. 1997; Wieser et al. 1994). On the other hand, biodegradation pathways in which the substituted phenol is converted via hydroquinone to maleylacetate have been found, mainly in organisms that grow on polyhalophenols or 4-nitrophenol (Kiyohara et al. 1992; Xun et al. 1992; Kadiyala and Spain 1998; Nordin et al. 2005; Perry and Zylstra 2007). Most studies have focused on the organisms and pathways that involve degradation via catechols.

A clear indication for the first step in the 4-FP catabolic pathway of strain IF1 was obtained by mass spectroscopic analysis of culture supernatants, which indicated the formation of hydroquinone as an early nonfluorinated intermediate. This result, in combination with the enzyme assays and oxygen uptake experiments, suggests the involvement of an inducible 4-FP monooxygenase that is NADH dependent, although the first expected product of this reaction, i.e., benzoquinone, was not detected. Monooxygenation of an aromatic substrate carrying an electron-withdrawing halogen group or a nitro-substituent can result in simultaneous hydroxylation and dehalogenation or nitrite removal. This has, for example, been described for the conversion of tetrafluoro-p-hydroxybenzoate by para-hydroxybenzoate hydroxylase (Husain et al. 1980) and the oxidation of 2,4,6-trifluorophenol by a monooxygenase fromRalstoniaeutrophaJMP134 (Xun and Webster 2004). The quinone that is produced is chemically reduced by NADH to a hydroxyquinol, leading to a net stoichiometry of two NADH oxidized per halide or nitrite that is released (Fig. 5). As in our case, no transient accumulation of a benzoquinone is usually observed, indicating that benzoquinone reduction is rapid. Other examples of halophenol degradation with formation of hydroquinone derivatives are the degradation of trichlorophenol to 2,6-dichlorohydroquinone by a strain of Pseudomonas pickettii (Kiyohara et al. 1992), the degradation of pentachlorophenol by a Sphingobium sp. through hydroxylation by a flavoprotein monooxygenase (Xun and Orser 1991), and the degradation of 4-chlorophenol by an Arthrobacter sp. (Bae et al. 1996).

Our results further indicate that hydroquinone is not the ring fission substrate because no enzymatic activities for hydroquinone dioxygenase or hydroxymuconic semialdehyde dehydrogenase were found. Instead, upon conversion of hydroquinone by strain IF1, we observed the formation of a transient metabolite that was identified as hydroxyquinol, indicating that a second hydroxylation takes place before ring fission. The production of hydroxyquinol could be due to the action of a separate hydroquinone monooxygenase, or it could be caused by a second hydroxylation step by the 4-FP monooxygenase. Conversion of hydroquinone to hydroxyquinol before ring fission was also suggested for strains Arthrobacter that degrade 4-nitrophenol (Perry and Zylstra 2007) and 4-chlorophenol (Nordin et al. 2005). Ring fission of hydroxyquinol would produce maleylacetate, which we did not detect as an intermediate in 4-FP degradation, but a transient metabolite was found with mass properties identical to those of 3-oxoadipate, which is expected to be the next intermediate in hydroxyquinol degradation.

Most cometabolic transformations of fluorophenols described in the literature involve the initial action of a phenol hydroxylase that results in the formation of fluorocatechols, which are subsequently transformed into fluoromuconates through a catechol 1,2-dioxygenase. The conversion usually proceeds with lactonization and elimination of fluoride (Boersma et al. 2001; Bondar et al. 1999; Finkelstein et al. 2000; Boersma et al. 1998; Bondar et al. 1999). Our results suggest that Arthrobacter strain IF1, which grows on 4-FP, possesses a new pathway for the degradation of 4-FP that involves immediate defluorination by a monooxygenase. Strain IF1 avoids the accumulation of possible toxic metabolites such as 3- or 4-fluorocatechol by this initial dehalogenation.

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